

Conceptual and empirical advances in Neotropical biodiversity research

Alexandre Antonelli^{1,2,3,4}, María Ariza^{1,2,5}, James Albert⁶, Tobias Andermann^{1,2}, Josué Azevedo^{1,2}, Christine Bacon^{1,2}, Søren Faurby^{1,2}, Thais Guedes^{1,2,7,8}, Carina Hoorn^{9,10}, Lúcia G. Lohmann^{11,12}, Pável Matos-Maraví^{1,2}, Camila D. Ritter^{1,2}, Isabel Sanmartín¹³, Daniele Silvestro^{1,2,14,15}, Marcelo Tejedor^{1,2,16}, Hans ter Steege^{17,18}, Hanna Tuomisto¹⁹, Fernanda P. Werneck²⁰, Alexander Zizka^{1,2} and Scott V. Edwards^{2,4,21}

- ¹ Department of Biological and Environmental Sciences, University of Gothenburg, Gothenburg, Sweden
- ² Gothenburg Global Biodiversity Centre, Gothenburg, Sweden
- ³ Gothenburg Botanical Garden, Gothenburg, Sweden
- ⁴ Department of Organismic Biology and Evolutionary Biology, Museum of Comparative Zoology, Harvard University, Cambridge, MA, USA
- ⁵ Laboratoire Ecologie et Biologie des Interactions, Team “Ecologie, Evolution, Symbiose”, Université de Poitiers, Poitiers, France
- ⁶ Department of Biology, University of Louisiana at Lafayette, Lafayette, LA, USA
- ⁷ Federal University of São Paulo, Diadema, Brazil
- ⁸ Museum of Zoology, University of São Paulo, São Paulo, Brazil
- ⁹ Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, Amsterdam, Netherlands
- ¹⁰ Universidad Regional Amazonica IKIAM, Napo, Ecuador
- ¹¹ Instituto de Biociências, Departamento de Botânica, Universidade de São Paulo, São Paulo, Brazil
- ¹² Integrative Biology, University of California, Berkeley, CA, USA
- ¹³ Real Jardín Botánico, Madrid, Spain
- ¹⁴ Department of Computational Biology, University of Lausanne, Lausanne, Switzerland
- ¹⁵ Swiss Institute of Bioinformatics, Lausanne, Switzerland
- ¹⁶ Instituto Patagónico de Geología y Paleontología, Puerto Madryn, Guatemala
- ¹⁷ Naturalis Biodiversity Center, Leiden, Netherlands
- ¹⁸ Systems Ecology, Free University, Amsterdam, Netherlands
- ¹⁹ Department of Biology, University of Turku, Turku, Finland
- ²⁰ Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil
- ²¹ Gothenburg Centre for Advanced Studies in Science and Technology, Chalmers University of Technology and University of Gothenburg, Gothenburg, Sweden

Submitted 21 July 2017
Accepted 27 August 2018
Published 4 October 2018

Corresponding author
Alexandre Antonelli,
alexandre.antonelli@bioenv.gu.se

Academic editor
Tomas Hrbek

Additional Information and
Declarations can be found on
page 28

DOI 10.7717/peerj.5644

© Copyright
2018 Antonelli et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

ABSTRACT

The unparalleled biodiversity found in the American tropics (the Neotropics) has attracted the attention of naturalists for centuries. Despite major advances in recent years in our understanding of the origin and diversification of many Neotropical taxa and biotic regions, many questions remain to be answered. Additional biological and geological data are still needed, as well as methodological advances that are capable of bridging these research fields. In this review, aimed primarily at advanced students and early-career scientists, we introduce the concept of “trans-disciplinary biogeography,” which refers to the integration of data from multiple areas of research in biology (e.g., community ecology, phylogeography, systematics, historical biogeography) and Earth and the physical sciences (e.g., geology, climatology, palaeontology), as a means to reconstruct the giant puzzle

of Neotropical biodiversity and evolution in space and time. We caution against extrapolating results derived from the study of one or a few taxa to convey general scenarios of Neotropical evolution and landscape formation. We urge more coordination and integration of data and ideas among disciplines, transcending their traditional boundaries, as a basis for advancing tomorrow's ground-breaking research. Our review highlights the great opportunities for studying the Neotropical biota to understand the evolution of life.

Subjects Biodiversity, Biogeography, Ecology, Evolutionary Studies, Genomics

Keywords Biogeography, Biotic diversification, Landscape evolution, Phylogeny, Scale, Biodiversity, Community ecology, Phylogeography, Phylogenetics, Tropics

INTRODUCTION

The Neotropical region (also referred to as tropical America or the American tropics) extends today from central Mexico to Argentina, including the Caribbean ([Morrone, 2013](#)). The region encompasses a vast range of biomes and habitat types, each with a particular history of landscapes and biotic evolution ([Fig. 1; Hughes, Pennington & Antonelli, 2013](#)). For many groups of organisms, the Neotropics are home to outstandingly high levels of species richness, when compared to other major biotic realms ([Lundberg et al., 2000; Antonelli & Sanmartín, 2011](#)). As such, understanding Neotropical biodiversity patterns and the processes associated with its origin and maintenance represents a major scientific priority.

Biodiversity refers to the diversity of life across all levels of biological organization ([Purvis & Hector, 2000; Gaston & Spicer, 2004](#)). Biodiversity is unevenly distributed across Earth, and varies among and within geographic regions, between terrestrial and aquatic ecosystems, and among different groups of organisms. Biodiversity increases from the poles to the equator, reaching the highest values in tropical regions, a pattern termed the latitudinal diversity gradient ([Willig, Kaufman & Stevens, 2003; Field et al., 2009](#)). However, this pattern is complex, with numerous non-diverse tropical or diverse non-tropical areas and taxa constituting exceptions. In addition, there are still numerous uncertainties in the underlying data used to generalize overall patterns. Most importantly, these patterns remain far from properly understood, and we are still struggling to identify their main determinants. As a result, researchers tend to focus on different aspects of biodiversity such as taxonomic, phylogenetic, and functional diversity (FD) ([Swenson, 2011; Tables 1–3](#)). Each of these aspects of biodiversity may vary among regions and taxa, and each must therefore be assessed by independent criteria ([Strecker et al., 2011; Magurran, 2013](#)).

SURVEY METHODOLOGY

In April 2017, we gathered scholars from several countries and scientific backgrounds to discuss Neotropical biodiversity during the “Origins of Biodiversity Workshop” organized by Chalmers University of Technology and the University of Gothenburg (Sweden), under the auspices of the Gothenburg Centre for Advanced Studies. We spent

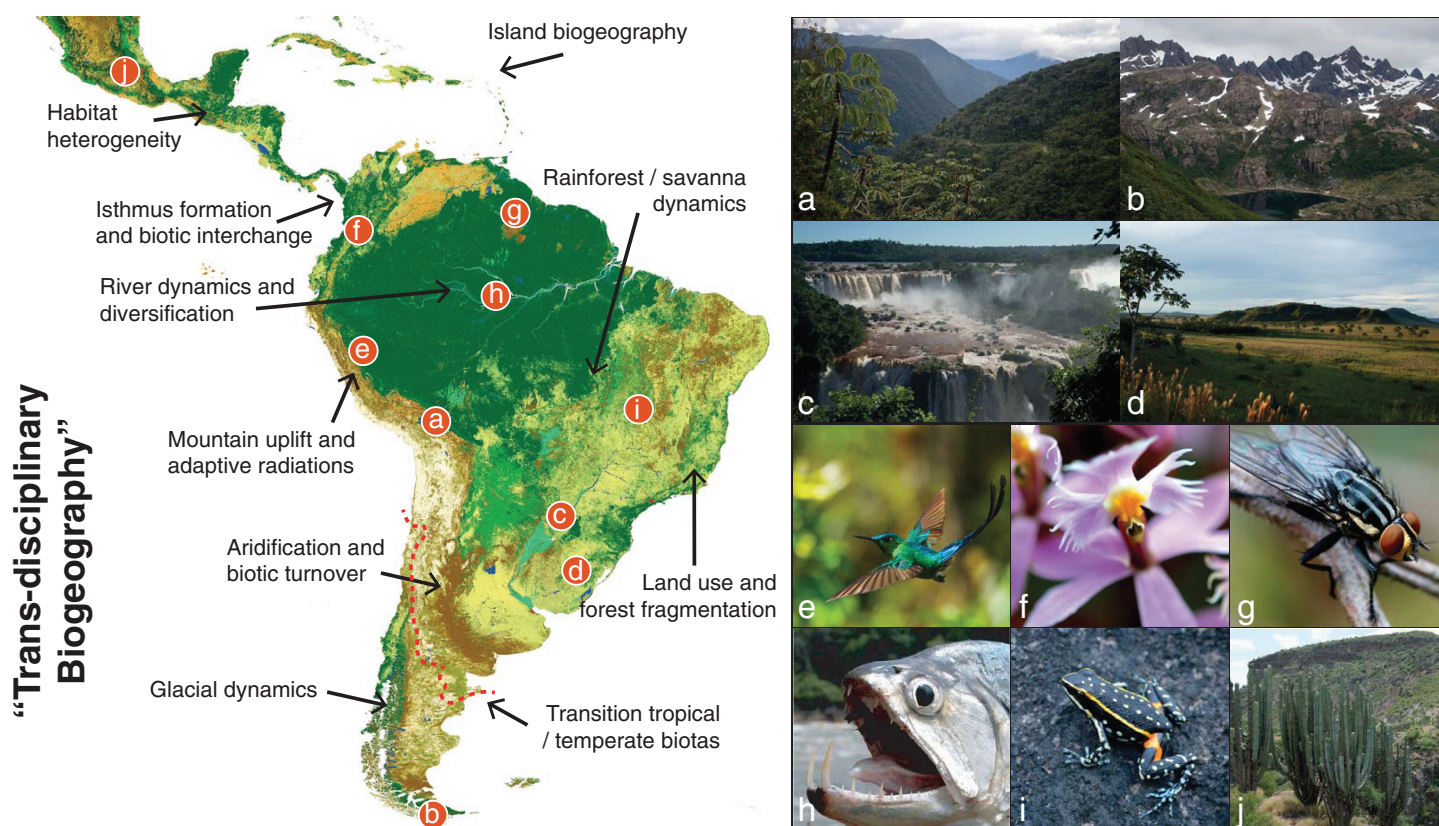


Figure 1 The giant Neotropical puzzle. Map of the Neotropical region, spanning from Central Mexico to central Argentina (red dashed line) and including all Caribbean Islands. The figure shows examples of the large diversity of Neotropical habitats and the taxa that inhabit those habitats. We also outline a few of the many topics in Neotropical biodiversity that can be studied in the “trans-disciplinary biogeographic approach” advocated here. (A) Eastern slopes of the Bolivian Andes, where the Amazonian and Andean biotas meet; (B) Patagonian mountains of southern Chile, which despite being in the temperate zone of South America is home to many Neotropical-derived lineages; (C) Iguazu waterfalls, where increased humidity create gallery forests within the South American open diagonal; (D) Southern grasslands of the Pampas, a naturally open habitat now largely influenced by human activity; (E) One of the ca. 338 known species of hummingbirds, a conspicuous clade currently restricted to the American continent and particularly diverse in the Andes; (F) *Epidendrum ibaguense*, a widespread species in the orchid family in which many new Neotropical species are discovered each year; (G) An unidentified fly in the inselbergs of southern French Guiana, where basaltic rocks emerge several hundred meters above the surrounding Amazonian rainforest; (H) The large dogtooth characin fish *Hydrolycus scomberoides*, exemplifying the world’s richest ichthyofauna in the Amazon drainage basin; (I) *Ameerega flavopicta*, a rock-dwelling frog species adapted to a region of high seasonality of precipitation; (J) A columnar cactus of central Mexico, near the northwestern limits of the Neotropical region where low-canopy forests and succulent vegetation build vegetation mosaics across the landscape. Map generated through the remote-sensing ESA GlobCover 2009 project and colored by biome assignments (©ESA 2010 and UCLouvain; http://due.esrin.esa.int/page_globcover.php). Photo credits: A–G, I and J: A.A.; H: J.A.

Full-size DOI: 10.7717/peerj.5644/fig-1

one week outlining the topics presented here, focusing primarily on recent advances and the future of the field. We continued to work remotely toward the conclusion of this publication. The overall goal of this review is to summarize the knowns and unknowns about Neotropical biodiversity, with focus on terrestrial taxa and ecosystems, and to discuss the many opportunities and challenges of this research field. We acknowledge that the wide breadth of the topics discussed here are in part due to this being a summary of ideas produced at a workshop of diverse participants, and that each topic contains a depth that cannot be simply synthesized. For instance, more comprehensive reviews on the theories proposed to explain the origin of Neotropical diversity can be found elsewhere (Moritz *et al.*, 2000; Antonelli &

Table 1 The concept of taxonomic diversity, and its use and challenges in the Neotropical context.

Definition. Taxonomic diversity refers to how many taxa can be found within a given area or higher clade, and how individuals are distributed among these taxa. Taxonomic diversity can be quantified at different taxonomic ranks (e.g., species, genera, families), with the species rank being the most popular by far. Species richness is widely viewed as a fundamental measure of overall biodiversity (Gotelli & Colwell, 2001). This is due to the fact that the species boundary defines the limits of genetic variation, natural selection, and adaptation (Sexton et al., 2009). While individual organisms live and die, the stable phenotypes recognized as species may persist for millions of years, serving as predictable components of the ecosystems in which all species function and evolve (Eldredge, 1989). As result, species are thought to constitute the basic structural and functional units in ecology and evolution (Tilman & Downing, 1994; Worm et al., 2006).

Generic and family-level taxonomic ranks are occasionally used in comparative studies, especially when species identification or delimitation is difficult (Bertrand, Pleijel & Rouse, 2006). However, the ranks that taxonomists must assign to higher-level taxa are often considered to be arbitrary constructs, reflecting little biological organization, and incorporating further biases and artefacts when compared, although opposing views exist (Humphreys & Barraclough, 2014). In general, species are seen as the “fundamental category of biological organization” despite the multitude of species definitions available (De Queiroz, 2005).

Metrics and usage. Taxonomic diversity is most commonly measured using taxon richness, that is, the number of taxa in a given area. However, relative abundance distributions can differ greatly among areas, and an area where taxon abundances are equal has intuitively higher diversity than an area with the same number of taxa but a high degree of dominance by one or a few taxa. Abundance differences can be taken into account by quantifying diversity as the effective number of species (also known as Hill number or true diversity; Hill, 1973; Jost, 2006; Tuomisto, 2010, 2018). Because its values are easier to interpret and compare than those of traditional diversity indices (e.g., Brillouin, Shannon, and Simpson indices), the effective number of species is emerging as the best general measure of diversity by a broad consensus. Quantitative abundance data are rather rare, though, and few studies have included abundance when discussing diversity in the Neotropics (but see Valdujo, Carnaval & Graham, 2013; Ter Steege et al., 2013; Tuomisto, Zuquim & Cárdenas, 2014; Jenkins et al., 2015; Moura et al., 2016; Azevedo, Valdujo & Nogueira, 2016; Gómez et al., 2018).

Observed taxonomic diversity is sensitive to sampling effort, especially at the species rank. Since communities typically contain many species that are locally rare, observed species richness provides only an underestimate of the number of species actually present, unless the community is very thoroughly sampled. The accuracy of estimates of taxonomic diversity depends on the number of individuals sampled, the size of the local species pool, the evenness of species abundances in the community, size and environmental heterogeneity of the area, and the status of taxonomic knowledge of the groups surveyed. When comparing estimates of local taxonomic diversity among areas, it is therefore important that they are based on quantitative and standardized sampling (Chao & Jost, 2012; Tuomisto, 2018).

Beta diversity and species turnover, reflecting heterogeneity in species composition among sites, are also of interest (Tuomisto, 2010, 2018; Higgins et al., 2011; Leprieur et al., 2011; J. M. Craig et al., 2018, unpublished data). However, quantifying these requires data where species identifications have been done consistently using a standard taxonomy, and such data are only available in some areas for some vascular plants (e.g., trees, ferns; Tuomisto, Ruokolainen & Yli-Halla, 2003; Arellano et al., 2016; Tuomisto et al., 2016), and some vertebrates (e.g., birds, primates, some fishes; Arrington & Winemiller, 2009). For these same organisms, a general understanding of species richness gradients has emerged (Kier et al., 2005; Albert et al., 2011; Rosauer & Jetz, 2014; Tuomisto, Zuquim & Cárdenas, 2014). For most other organisms, too few data are available to allow accurate circumscriptions of taxa and reasonable estimates of species richness gradients and species turnover (Andújar et al., 2015). Indeed, the smaller and less conspicuous the organism, the poorer the state of knowledge. For instance, very little is known about microbial and fungal diversity, and insect diversity is similarly understudied (Basset et al., 2012). However, even among well-studied and charismatic Neotropical taxa—such as birds and mammals, even river dolphins—there are still new species to discover (Hrbek et al., 2014).

Sanmartín, 2011; Leite & Rogers, 2013; Table 4). We focus here on topics that we think require further development within this research field.

WHAT DO WE KNOW ABOUT PATTERNS OF NEOTROPICAL BIODIVERSITY?

Taxonomic diversity

Relatively good estimates of taxonomic diversity are only available for well-studied Neotropical taxa, as in other parts of the world. These estimates have been used to identify the best predictors of diversity at large scales (Jenkins et al., 2015; Moura et al., 2016). Although sampling across taxa is comparable or even greater in the Neotropics than in other tropical regions (Fig. 2; Table 1), taxonomic diversity is generally underestimated within the Neotropics, especially for poorly sampled organisms such as fungi, invertebrates, and micro-organisms.

Table 2 The concept of phylogenetic diversity, and its use and challenges in the Neotropical context.

Definition. Phylogenetic diversity (PD) assesses cumulative evolutionary distinctiveness within and among areas and taxa. We do not contest the usefulness of species as entities in the assessment of biodiversity patterns, conservation, and many other disciplines, from medicine to bioengineering. However, species are not universally comparable units, given differences in species concepts, operational criteria of delimitation, and circumscriptions among areas, taxa, and taxonomists. In addition, species differ widely in their evolutionary ages, geographic distributions, habitat tolerances, and degree of genetic structure. Species also differ in the biological attributes of their constituent organisms, and therefore, in the effects that these traits may have on ecological and evolutionary processes. Furthermore, species are really just the tips of larger phylogenetic trees evolving through time. PD is therefore a useful measure to directly compare the degree of phylogenetic divergence among groups and regions.

Metrics and usage. The basic idea of PD is to measure the total amount of lineage evolution through time (i.e., branch length) observed among all members of a clade or area (Faith, 1992). Overall, PD has been shown to provide a better estimate of “feature divergence” than species richness alone (Forest et al., 2007), because PD multiplies the species (tips of the tree) and multi-species clades (tree branches with multiple tips) by their phylogenetic age, usually measured in millions of years. However, there are many ways of deriving and applying such metrics from phylogenies. As such, researchers should try to choose the most appropriate index for each situation, as well as acknowledge these differences in cross-taxonomic comparisons (Tucker et al., 2016). Phylogenetic diversity (or more explicitly: divergence), although not “visible” constitutes a potentially powerful concept for increasing the standardization of biodiversity analyses, the recognition of areas for conservation, and our understanding of evolutionary history of clades, among others.

Complementary to phylogenetic diversity based on the relationships among taxa, patterns of genetic variation within species also represent a vital but often under-appreciated component of biodiversity. Knowledge of intraspecific genetic variation may also improve the prediction of a species ability to adapt to changing climates, as well as improving understanding of the process of speciation. This type of information is particularly important in the light of ongoing anthropogenic climate change. However, our current knowledge of species genetic diversity is restricted to a few selected species, and overall patterns of intraspecific genetic diversity remain poorly understood. Even among well-studied groups (e.g., mammals), spatial patterns of genetic diversity are effectively unknown within the tropics.

Table 3 The concept of functional diversity, and its use and challenges in the Neotropical context.

Definition. Functional diversity (FD) measures differences in the physiological, behavioral, and ecological characteristics of organisms, and how biological trait values (such as body mass of animals, and life form or habit of plants) affect ecological and evolutionary processes. Knowledge about species traits and ecological functions (such as a species trophic level, and including the variation in traits within and among species) is a crucial component of biodiversity. However, this is one of the major shortcomings in current biodiversity knowledge, especially in tropical areas. Few studies to date have mapped large-scale patterns of functional diversity, although efforts in this direction are underway (see for fishes e.g., Arbour & López-Fernández, 2014; Toussaint et al., 2016).

Metrics and usage. Apart from the lack of data, the theory behind functional diversity is not yet well consolidated. We still do not know which traits are ecologically and evolutionarily important for different groups, how to compare traits for different sets of organisms, and how functional diversity affects ecosystem productivity, stability, and resilience, especially in the tropics. An additional shortcoming is associated with biotic interactions. Apart from basic information on pollination and dispersal syndromes, we know surprisingly little about most biotic interactions. Very few species interaction networks are available to date (Toju et al., 2017).

Several examples of species-rich, yet incompletely-documented faunas are available, including large clades of freshwater fishes, amphibians, and some groups of reptiles. Although about 5,600 species of freshwater fishes are currently known in the Amazon, the Orinoco, and adjacent river basins of tropical South and Central America, more than 100 new species are described every year (Van Der Sleen & Albert, 2017). In other words, approximately two new species are described per week, although an even higher number of new species would be expected if more trained taxonomists were available. This rapid pace of species description is not slowing down, and recent estimates for the total number of Neotropical freshwater fishes exceed 8,000 species (Reis et al., 2016). This estimate is remarkable, implying that more than 2,400 fish species might remain to be described in the Neotropics alone, a number that exceeds the combined number of rodent species currently known on Earth. This large number of expected, but still hidden, lineages represents an example of the “known unknowns” of Neotropical biodiversity (Table 5),

Table 4 Some of the many theories proposed to explain the high levels of (Neo)tropical biodiversity.

Theory	Key proponent(s)	Summary	Comment
Riverine barrier hypothesis	Wallace (1889)	The formation of large lowland Neotropical rivers like the Amazon resulted in genetic isolation and speciation in taxa ecophysiologically restricted to non-flooded rain forests.	Dynamic river capture is even more effective at isolating and reuniting populations than is the static geometry of dendritic river basins.
Pleistocene refugium theory	Haffer (1969)	Most Amazonian birds, and probably other taxa, originated recently in response to Pleistocene climate changes. The repeated contraction of forests in relation to savannas led to the isolation of populations and inability of breeding once they came into secondary contact during inter-glacials.	From initial support for plants and other taxa, this theory has been heavily criticized based on lack of geophysical evidence for savanna expansions, old divergence times from phylogenies, etc.
Time-area integrated hypothesis	Fine & Ree (2006)	Diversity can be predicted by the amount of time that species spend in a region, multiplied by the total area of that region.	A modification of this model is a strong predictor of dispersal events across the Neotropics (Antonelli et al., 2018).
Phylogenetic niche conservatism	Wiens & Donoghue (2004)	Tropical biotas are more diverse because many lineages of the modern biosphere evolved in the super-greenhouse world of the Mesozoic and early Cenozoic 140–50 Ma and remained in their original environment.	Most clades have origins in warm and wet tropical climates. Most clades at higher latitudes adapted to cold and dry conditions in the Neogene and Quaternary.
Out of the Tropics	Jablonski, Roy & Valentine (2006)	Tropical biotas are more diverse because lineages have higher speciation rates, lower extinction rates, and higher net emigration over immigration than lineages in extra-tropical regions.	This is just one popular theory among several others attempting to explain the latitudinal diversity gradient in species richness.
Metabolic theory of ecology	Brown & Svenning (2013)	Higher metabolic rates translate into higher rates of speciation and extinction at low latitudes.	Incompletely developed mechanistic links between kinetics at the metabolic, ecophysiological, and evolutionary scales.
Tropical productivity	Gillman et al. (2014)	Species richness is positively correlated with net primary productivity because larger populations are less likely to stochastically fluctuate to a population size of zero, which is a sticky boundary.	Metanalyses have shown a unimodal relationship is more common than a monotonic between productivity and species richness (Fraser et al., 2015).
Sea-level fluctuations	Nores (2002)	Repeated sea-level rises during the late Cenozoic led to the allopatric speciation of Amazonian species in true islands.	Model based on current topography, lacking other geophysical evidence.
Museum hypothesis	Stebbins (1974)	Tropical lowlands act as “museums” of diversity, in which species of different origins gradually accumulate.	The Neotropics is now considered both “museum” and “cradle” of diversity (McKenna & Farrell, 2006).

which may have different underlying explanations (such as lack of taxonomic training, uneven distribution of resources across taxa and habitats, morphological complexity, among others).

Current knowledge of taxonomic limits of Neotropical amphibians and reptiles is gradually improving. Several molecular studies have detected high levels of cryptic diversity, that is, the existence of two or more lineages within a known species ([Bickford et al., 2007](#)), indicating that the known taxonomic diversity is still underestimated in many orders ([Funk, Caminer & Ron, 2011](#); [Fouquet et al., 2012](#)). Even

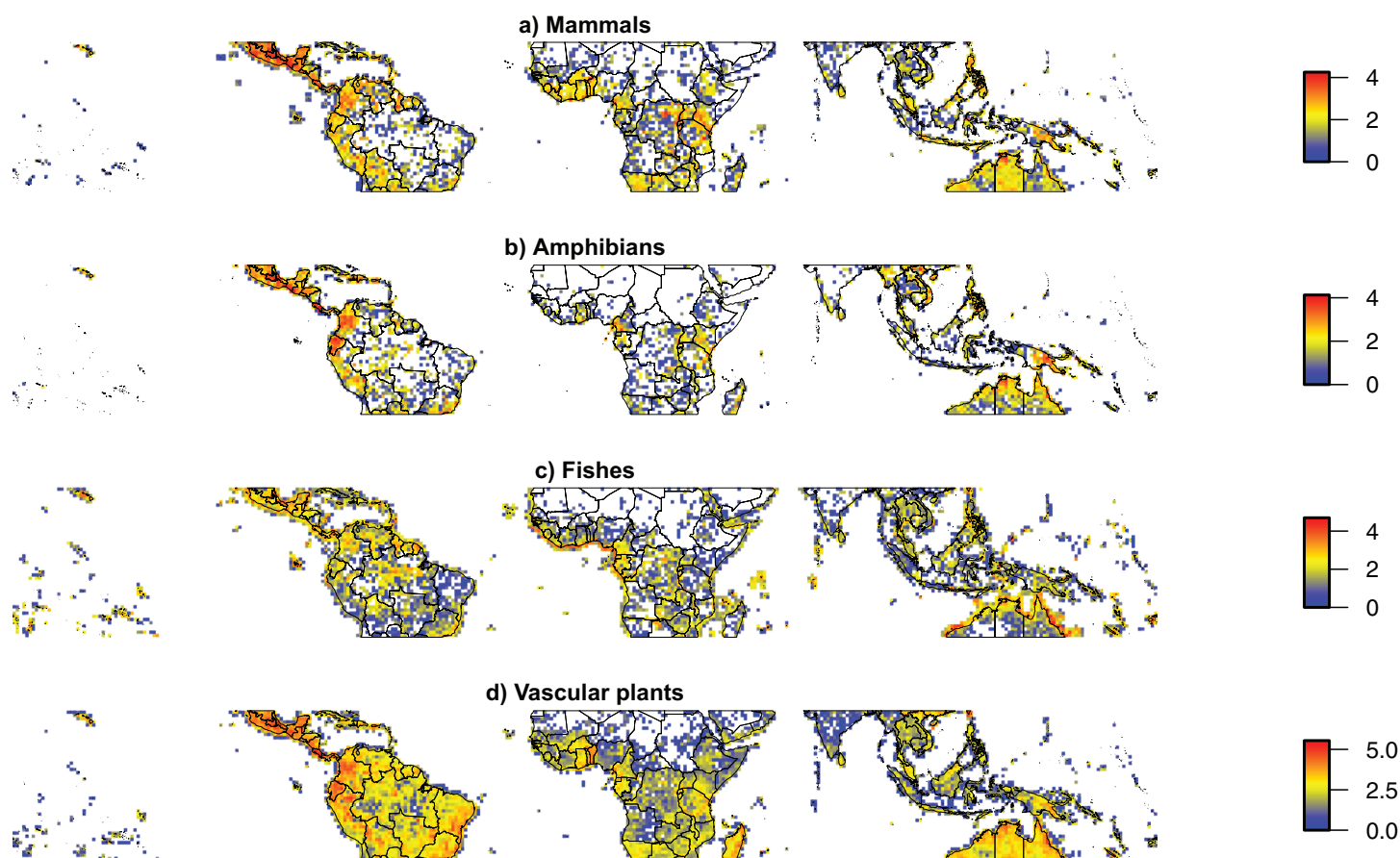


Figure 2 Taxonomic sampling across the world's tropics. Density maps for geo-referenced species occurrences available from the Global Biodiversity Information Facility for (A) Mammals, (B) Amphibians, (C) Fishes, (D) Vascular plants between the Tropics of Cancer and Capricorn (23.5°S–23.5°N), showing the main spatial biases of taxonomic sampling. All datasets were cleaned for automatically detectable errors using SpeciesGeoCoder (Töpel et al., 2016). The figure is shown on a cylindrical equal area projection with standard parallels of 11.75°S and 11.75°N. The width of each cell is consistently 1°, while the height of each cell is 1° at the standard parallels, slightly lower at the equator and slightly higher at the Tropics of Cancer and Capricorn. Colors indicate 10-based logarithm of the number of records. [Full-size](#) DOI: 10.7717/peerj.5644/fig-2

in the much more densely sampled and well-studied Atlantic Rainforest of Brazil, charismatic species of frogs are still being discovered. For instance, seven new species of *Brachycephalus* were recently described for this region (Ribeiro et al., 2015). Likewise, intraspecific analyses of Neotropical lizards show that the occurrence of cryptic diversity is often manifested across biomes. This subdivision of broadly distributed taxa into multiple cryptic species with more restricted geographic distributions increases the perception of biological diversity of a given region, and has numerous implications for biogeography (Werneck et al., 2012b) and conservation (Simões et al., 2014).

For plants, a quantitative assessment on the discovery of Amazonian trees during the last three centuries was compiled by Ter Steege et al. (2016), showing clear peaks in herbarium collections and new species descriptions. Although the data show a drop in the collection of unknown taxa after the 1980s (Ter Steege et al., 2016), there are still enormous discoveries to be made. For example, in a few years of increased collection efforts, the

Table 5 The various components of Neotropical biodiversity, examples of major aspects known about them, and some of the key topics that remain to be understood.

Biodiversity components	Known knowns	Known unknowns	Unknown unknowns
Taxonomic diversity	Approximate species numbers for macroscopic organisms; human impacts tend to decrease overall diversity	Large portions of biodiversity are unexplored (i.e., microbes, invertebrates, fungi)	Taxonomic units used in biodiversity studies may not represent comparable ecological or evolutionary units
Genetic variation (within species)	Patterns of genetic variation known for very selected taxa	Overall patterns of genetic variation	How generalizable are conclusions drawn by such limited patterns of genetic diversity
Phylogenetic diversity	General understanding of the tree (or network) of life	Drivers of diversification	Potential biases in phylogeny reconstruction and time-calibration
Spatial patterns of diversity	Hotspots and general patterns of species richness and diversity; broad species ranges for charismatic taxa	Areas of endemism; known patterns of biodiversity are biased; ecological preferences of species; drivers of diversity	Human impact to overall spatial patterns
Functional diversity	Large scale productivity patterns	Biotic interactions	Relevance of current functional diversity measures; equivalency in functional traits; relationship between current and future functional diversity

Guide of the Ducke Reserve (Brazil; [Ribeiro, 1999](#)), which covers one of the most thoroughly studied areas of Amazonian forest, increased the number of known vascular plants from 825 ([Prance, 1990](#)) to 2,079 ([Hopkins, 2005](#)).

One difficulty in assessing taxonomic diversity is that taxonomic units may vary according to the preference of the taxonomist revising a particular group (e.g., whether a “splitter” or a “lumper”), and by the data and methodologies underlying taxonomic revisions and species circumscriptions. This issue becomes obvious when taxonomic treatments of the same group are produced by different researchers independently. For example, the Neotropical palm genus *Attalea* included 29 species in one monograph ([Henderson, Galeano & Bernal, 1995](#)), and 65 species in another taxonomic treatment published just 4 years later ([Glassman, 1999](#)). Similarly, the Caribbean palm genus *Coccothrinax* included 14 species in one taxonomic treatment ([Henderson, Galeano & Bernal, 1997](#)) and 53 species in another ([Dransfield et al., 2008](#)). Personal preferences to “lumping” vs. “splitting” among taxonomists may have large consequences for biodiversity estimates and have been shown to strongly affect diversification rate estimates ([Faurby, Eiserhardt & Svenning, 2016](#)). Taxonomic practices should therefore be considered when comparing taxonomic biodiversity at any scale, and whenever adequate, researchers should take advantage of explicit and reproducible criteria for species delimitations.

Besides lumping vs. splitting, species lists may vary among authorities depending on the inclusion criteria, such as whether or not to include rare occurrences of a species common elsewhere, and how to classify the life forms of species (e.g., primarily herbaceous plants rarely recorded as trees). For large regions such as Amazonia, these are some of the reasons why the number and contents of species lists may differ substantially ([Cardoso et al., 2017](#); H. Ter Steege et al., 2018, unpublished data).

Phylogenetic diversity

Many Neotropical clades are known from just one or a few species that may represent relictual survivors of ancient and otherwise extinct groups (Table 2). This phenomenon is known from most organism groups, from Neotropical fishes (Albert *et al.*, 2011) to plants (Wilson *et al.*, 2012). To study how differences in diversity arise among taxa, some researchers have turned their attention to the study of early-branching, low-diversity clades. Examples of such clades include the leafy cacti (*Pereskia* and *Leuenbergeria* spp.; Cactaceae), the South American lungfish (*Lepidosiren paradoxa*; Lepidosirenidae), the hoatzin (*Opisthocomus hoazin*; Opisthocomidae), and the coral pipe snake (*Anilius scytale*; Aniliidae). In contrast, other species are members of species-rich Neotropical clades still in the full bloom of their diversification, like the lianas of tribe Bignonieae with more than 400 species (Lohmann & Taylor, 2014), palms with over 730 species (Dransfield *et al.*, 2008), armoured catfishes (Loricariidae) with 680 species (Armbruster, Van Der Sleen & Lujan, 2018), and tanagers (Thraupidae) with 371 species (Burns, Unitt & Mason, 2016).

The first attempts to map phylogenetic diversity (PD) over continental and global scales were conducted for select vertebrate groups for which phylogenies were available and for which distribution patterns are relatively well known, such as amphibians, birds, and mammals (Safi *et al.*, 2011). Other than these, large-scale phylogenetic and FD studies with focus and dense sampling in the Neotropics are scarce. Some progress has been made in mapping PD patterns in the Neotropics for specific clades (Lehtonen *et al.*, 2015; Lovejoy *et al.*, 2010; Rossatto, 2014; Fenker *et al.*, 2014) or at the intraspecific level in the search for areas of high phylogeographic diversity and endemism (Carnaval *et al.*, 2014; Smith *et al.*, 2017; Melo *et al.*, 2018). Several ongoing studies by independent research groups are now working to broaden our knowledge on the spatial distribution of Neotropical PD.

BIASES AND GAPS IN NEOTROPICAL BIODIVERSITY KNOWLEDGE

There are two main sorts of biodiversity biases and gaps: taxonomic (also called the “Linnaean shortfall”) and spatial (the “Wallacean shortfall”). Taxonomically, detailed information on the richness and geographical distribution of species is restricted to certain well-studied taxa (e.g., primates and birds). We also know much more about organisms on land than those living in freshwater systems, including lakes, rivers, and swamps, leaving a large gap in the knowledge of Neotropical aquatic diversity. Spatially, our knowledge is concentrated to few well-studied areas (e.g., La Selva in Costa Rica, Barro Colorado Island in Panama, the Ducke Reserve in Brazil, Manu National Park in Peru, Yasuni National Park in Ecuador). Some regions stand out as having the lowest levels of sampling, including some parts of Central America, the central Andes, the Caatinga, and large parts of Amazonia (Fig. 2), where we have almost no occurrence records available (Hopkins, 2005; Feeley, 2015; Ter Steege *et al.*, 2016; Tedesco *et al.*, 2017). Clearly, for the vast majority of taxa, regions, and ecosystems, biodiversity knowledge is still scarce.

In general, knowledge of species distributions and diversity patterns are strongly biased toward areas that are more easily accessible by roads, rivers, and research stations (Hopkins, 2005; Meyer *et al.*, 2015). Interestingly, at least Amazonian trees, there seems

Table 6 The commonness of rarity in Neotropical diversity.

Most Neotropical species are rare, narrowly distributed, and endemic to particular regions or biomes (see [Albert et al., 2011](#) for fishes; [Ter Steege et al., 2016](#) for plants). Species with low abundances and narrow geographic ranges, as well as those confined to special habitats or areas, represent a sizable portion of Neotropical diversity. Indeed, a recent study extrapolating population size for Amazonian trees suggests that most species in the region are represented by comparatively few individuals ([Ter Steege et al., 2013](#)). Another study suggests that a considerable fraction of the rare species in the region may actually have relatively large distribution ranges ([Zizka et al., 2017](#)). However, many apparently widespread species in most taxonomic groups have been shown to contain multiple phylogenetic species, a possibility that remains poorly explored in the Neotropics but has important consequences for our understanding of diversity patterns and conservation priorities ([Bickford et al., 2007](#); [Colli et al., 2016](#)). The contributions of rare species to diversity patterns are difficult to quantify and remain largely obscure ([Coddington et al., 2009](#)), partly because most truly rare species will be completely unknown, and partly because rareness in the ecological sense is hard to define, depending on a variety of aspects, including the species concept adopted and the taxonomic preferences.

In both fish and plant taxa, areas of endemism separated by prominent biogeographic barriers, such as Amazonian and Mesoamerican rainforests currently separated by the Andes, arise from dispersal limitation, and differential environmental tolerances ([Bemmels et al., 2018](#)). In contrast, for some tree genera of rainforest trees, dispersal does not seem to be a constraining factor, meaning that community assemblages either represent random draws from the possible species pools available ([Dexter et al., 2017](#)), or functional differences arising from different habitat tolerances. The geographic distributions of many riverine and floodplain taxa are limited by river basin watersheds, and opportunities for dispersal include river capture events ([Albert et al., 2017](#)). Finally, it is not enough to know where particular species occur; we also need to know where these species do not occur ([Soria-Auza & Kessler, 2008](#)). It is, therefore, difficult to reliably say if the biodiversity patterns known to date really reflect true patterns or biases in collection effort. Further, patterns of species richness are usually discerned relatively early in the documentation of a newly explored biota, whereas patterns of species endemism are more difficult to discern as they require positive knowledge of both where species are present and absent ([Soria-Auza & Kessler, 2008](#)).

also to exist a bias toward reporting rare species ([Table 6](#)), as most scientific collectors tend to over-collect rare or uncommon trees ([Ter Steege et al., 2011](#)), although this pattern may not exist for other taxa. Although bioinformatic solutions may now assist in cleaning, predicting, and validating species occurrence data, taxonomic expertise is still essential but limited ([Maldonado et al., 2015](#); [Töpel et al., 2016](#)). As a result of our limited knowledge on species distributions patterns, and large gaps in knowledge about climatic and edaphic conditions for large portions of the Neotropics, the ecological requirements for most species remain unknown ([Table 5](#)).

INTRODUCING “TRANS-DISCIPLINARY BIOGEOGRAPHY”

Here, we propose that the best way to fully understand the complexity of Neotropical biodiversity is by conceptualizing and implementing a novel holistic framework. We define **trans-disciplinary biogeography** as “a holistic framework that takes advantage of the methods and data in multiple disciplines, in order to solve complex questions about the evolution, maintenance, and distribution of biodiversity through time and space. By doing so, each individual discipline transcends its traditional borders.”

The idea and need of combining data from different sources in biogeography has been advocated before (e.g., [Ribas et al., 2012](#); [Weeks, Claramunt & Cracraft, 2016](#)) but we propose a major expansion. Some examples of the constituent disciplines in this pursuit include biology (e.g., community ecology, phylogeography, systematics, taxonomy, historical biogeography; [Lomolino, Riddle & Whitaker, 2017](#)), geology (e.g., palaeontology, sedimentology, geomorphology), and climatology (e.g., modeling, speleology), amongst others. Successful examples of trans-disciplinary research include the archaeology-ecology synergy that led to the elucidation of pre-Columbian effects on the distribution of Amazonian plants ([Levis et al., 2017](#)); the genetic-geology synergy that led to the discovery of an earlier and more prolonged biotic interchange between South and

North America since the Miocene ([Bacon et al., 2015](#); [De Baets, Antonelli & Donoghue, 2016](#)), and the geology-biogeography-systematics synergy that led the discovery of a Miocene origin for the modern transcontinental Amazon river ([Hoorn et al., 2010b](#); [Albert, Val & Hoorn, in press](#)). Some of these interactions are already recognized as new sub-disciplines, such as “community phylogenetics” ([Swenson, 2011](#)), “geogenomics” ([Baker et al., 2014](#)), and “geodiversity” ([Gray, 2004](#)). We envision the integration of a high number of additional synergistic sub-disciplines.

In practice, we want to encourage young students and researchers to invest time in learning more about disciplines that might fall outside their general curriculum, but which could contribute to creating fruitful synergies. Obviously, not every project or publication has to (or should) be trans-disciplinary, and this pursuit should not decrease the depth of a researcher’s skills in her or his topic of expertise. But without trans-disciplinary frameworks that are defined in early stages of new research projects, there is a risk that important perspectives are missed out.

To showcase the benefit of these interactions, we provide some background on the emergence of Neotropical biogeography as a research focus—which was integrative from its early days, but successively lost much of its cross-disciplinarity. We then discuss how trans-disciplinary biogeography may help address the interactions between landscape evolution, climate change and biotic diversification at its multiple levels (see also [Hoorn, Perrigo & Antonelli, 2018](#)).

Early ideas about Neotropical biogeography

The Prussian naturalist Alexander von Humboldt was among the first to realize that biotic and abiotic processes interact to constrain species distributions, and to place these influences into a geological and climatic framework. He came to this notion in the Neotropics, most famously during his study of the Chimborazo volcano in Ecuador, where he carefully documented the location of different species along elevational zones ([Humboldt & Bonpland, 1805](#)). It was in this trip that he first observed that physical parameters such as topography and climate were key for geographic distributions.

A century later, [Wegener \(1912\)](#) advanced the incipient field of historical biogeography with the theory of continental drift, based in part on past geographic distributions of biotas linked by previously connected continental plates. The striking fit between the coastlines of South America and Africa was one of the pieces of evidence inspiring Wegener’s theory of dynamic, non-static landmasses. In the 1960s, a geophysical mechanism for plate tectonics was proposed ([Vine & Matthews, 1963](#)), placing studies of plant and freshwater fish biogeography into a plate tectonic framework, where vicariance was assumed as a major biogeographic force ([Raven & Axelrod, 1974](#); [Rosen, 1975](#)).

At first, the explanatory power of vicariance biogeography was the ability to predict biogeographic distributions of individual taxa and that of whole biotas from knowledge of how landscapes changed through time ([Rosen, 1978](#)). The paradigmatic example is the geological fragmentation of the Gondwana supercontinent, and the resulting fragmentation of the resident Gondwanan biotas. The vicariance biogeography approach

satisfies the scientific impulse of systematists and biogeographers for general explanations of organismal distributions, rather than ascribing each distribution to the vagaries of idiosyncratic evolutionary histories ([Humphries & Parenti, 1999](#)).

Soon after, the challenge to vicariance biogeography as a general theory was the commonplace observation that vicariant cladogenesis (i.e., allopatric speciation) is only one of three general macroevolutionary processes, along with dispersal and extinction ([MacArthur & Wilson, 1967](#); [Ree & Smith, 2008](#)). Indeed, ecologists have long understood dispersal to be a pervasive process influencing biogeographic distributions ([Cowie & Holland, 2006](#)). Long-distance dispersal has been documented in the formation of many biotas worldwide ([Sanmartín & Ronquist, 2004](#); [Bacon, Baker & Simmons, 2012](#)) including those in the Neotropics ([Smith et al., 2014](#); [Tagliacollo et al., 2015b](#); [Hawltischek, Ramírez Garrido & Glaw, 2017](#); [Antonelli et al., 2018](#)).

Inferring landscape evolution in the Neotropics

Neotropical historical biogeography increasingly relies on geological models that specify the landscape configurations on which species originate, disperse, and go extinct. Understanding phylogeny and biogeography in the context of landscape evolution requires assessment of geological data, including sedimentary environments, sedimentation rates, palaeontological records, and geochronological ages, among others ([Salo et al., 1986](#); [Räsänen et al., 1995](#); [Räsänen, Salo & Kalliola, 1987](#); [Hoorn et al., 1995](#); [Lundberg et al., 2000](#); [Figueiredo et al., 2009](#); [Hoorn et al., 2010b](#); [Lagomarsino et al., 2016](#); [Sanín et al., 2016](#); [Jaramillo et al., 2017](#); [Hoorn et al., 2017](#)).

Some recent reconstructions of the Neogene landscape in Amazonia are based on dynamic topography, in which mantle movements through time are quantified ([Shephard et al., 2010](#)). The effects of these movements are estimated on surface subsidence and are then related to environmental and landscape changes, such as the model applied to explain the genesis of the Pebas wetland in western Amazonia ([Hoorn et al., 2010a](#)). Another approach is to use numerical modeling and create reconstructions from physical parameters such as rates of erosion and mountain uplift. An example is the reconstruction of the flow of the Amazon River which incorporates surface processes, flexural isostasy and crustal thickening due to orogeny into a mathematical model to explain the drainage reversal in the Miocene ([Sacke, 2014](#)). However, this study did not incorporate the synergic effects of plate movements and surface dynamics, that are known to have impacted on the formation of mega-wetlands and ecosystems ([Horton, 2018](#)).

Landscape evolution models (LEMs) can be useful in a biological context but often lack spatial and temporal precision. Biological data can help to infer past landscapes, by testing alternative geological models and increasing their precision. However, we caution that the evolutionary history of one clade might represent an idiosyncratic story, rather than inform the general evolution of an entire landscape in which the clade occurs ([Cruz-Neto, Garland & Abe, 2001](#); [Ribas et al., 2012](#)).

In recent years, integrated approaches have built integrative LEMs based on geological, climatic and biodiversity data ([Craw et al., 2016](#); [Badgley et al., 2017](#); [Costa et al., 2017](#)). Some studies make use of geographic information systems and combine these with

well-dated palynological databases, such as Neotoma (<https://www.neotomadb.org/>). These models are mainly applied to reconstruct landscapes across Quaternary time scales (i.e., the past 2.6 million years). For example, reconstruction of changes in connectivity across the northern Andes enabled the inference of cyclic phases of biotic dispersal and speciation vs. extinction ([Flantua et al., 2015](#)). Molecular phylogenetic data can be used to statistically evaluate the likelihood of competing geological models on longer time scales, such as the closure of the Central American Seaway dividing South and Central America ([Bacon, 2013](#)), and the roles of the Caribbean plate margins as dispersal corridors between South and Central America ([Tagliacollo et al., 2015a](#)). Similar approaches based on both terrestrial ([Baker & Couvreur, 2012](#)) and aquatic taxa ([Hrbek, Seckinger & Meyer, 2007](#)) may provide important insights when geological data are insufficient or ambiguous.

The impact of the Andean uplift on Neotropical diversification

Neotropical biodiversity can only be properly understood when considering the Andean uplift and the effects of this orogeny on the landscape ([Fig. 3](#)) and regional climate ([Räsänen, Salo & Kalliola, 1987](#); [Gentry, 1982](#); [Hoorn et al., 2010b](#)). Although the Andes are entirely confined to South America, their formation has led to far-reaching effects across the Neotropics, and there are clear links with orogenies in Mesoamerica caused by plate tectonics.

The Andes today

The 7,000 km long Andes is positioned perpendicular to the principal global atmospheric currents and traps the humid air of the Intertropical Convergence Zone. This configuration enhances precipitation along the Andean slopes and in western Amazonia, making them wetter than they would be in a low Andes setting. Moreover, the high Andes also redirects the atmospheric flow inducing the southward deflected South American low-level jet ([Garreaud & Muñoz, 2005](#); [Insel, Poulsen & Ehlers, 2010](#); [Rohrman et al., 2016](#)).

The situation is reversed in southern and in northwestern South America. In these regions, the Andes trap the humid air of the Southern Hemisphere westerlies ([Garreaud & Muñoz, 2005](#)). In contrast to the Amazonian settings, the eastern margin of the Andes at its northern and southern extremes forms a rain shadow where semi-desert conditions prevail, and on the western flank there is increased precipitation with more humid conditions ([Blisniuk et al., 2005](#); [Palazzesi et al., 2014](#)), although this situation is partially reverse during El Niño events. The monumental Andean barrier has thus imposed strong impacts on both the climate and landscapes of South American lowlands, resulting in the modification of river systems and drastic changes in the climate and habitats of many Neotropical regions.

Impact on biotic diversification

The rise of the northern Andes had a major impact on Neotropical biodiversity, as documented for many taxa ([Cruz-Neto, Garland & Abe, 2001](#); [Hughes & Eastwood, 2006](#); [Santos et al., 2009](#); [Tagliacollo et al., 2015b](#); [Sanín et al., 2016](#); [Chazot et al., 2016](#); [Diazgranados & Barber, 2017](#); [Bacon et al., 2018](#)). Recent studies that explicitly

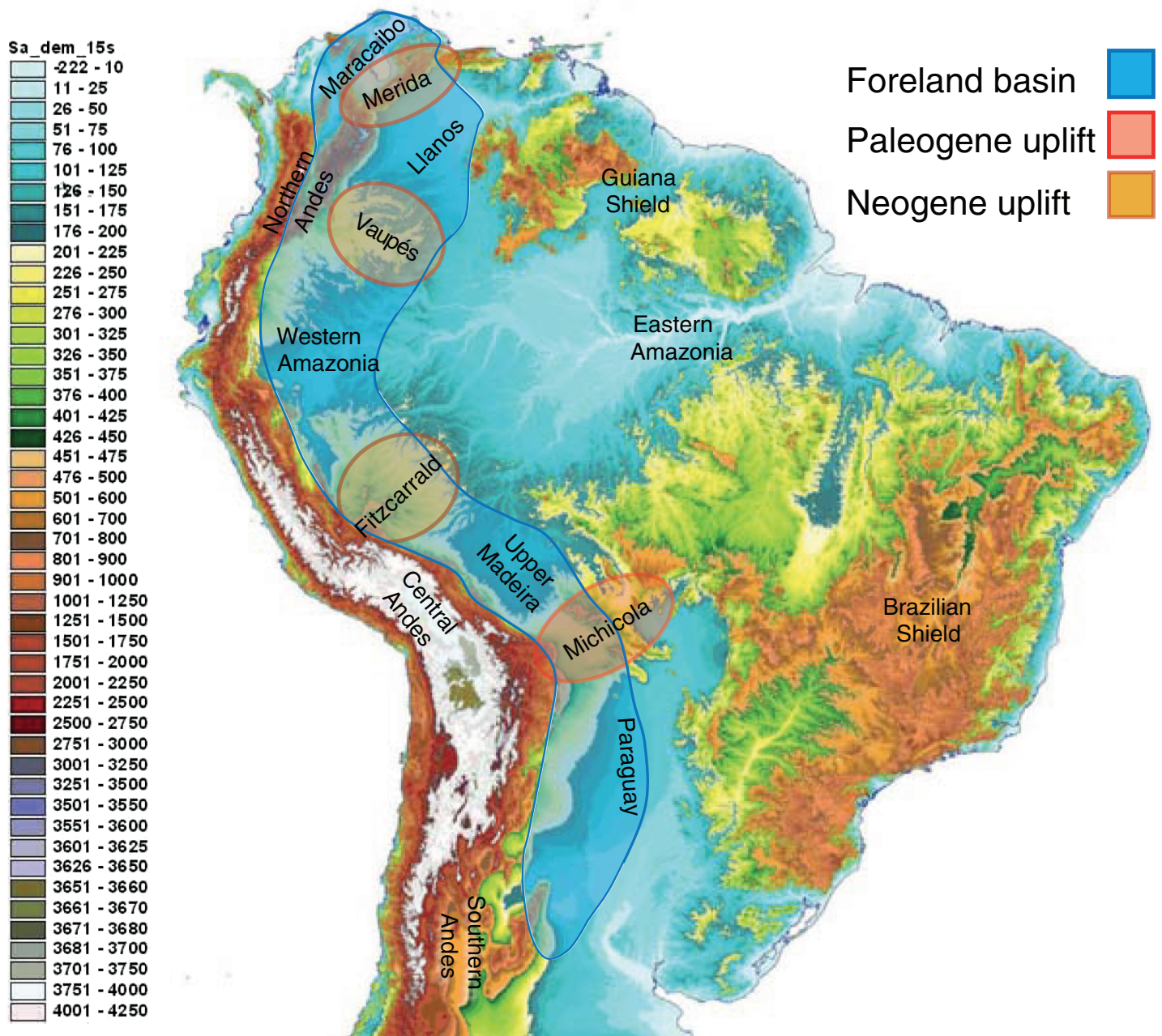


Figure 3 The complex topography and geology of South America. This map highlights the topographic differences across the continent, including the Precambrian and Paleozoic upland shields, and the Andean cordilleras and structural arches that uplifted during the Cretaceous and Cenozoic. The Sub-Andean foreland basin constituted the main drainage axis of South America for most of the past 100 million years, serving as the main arena of evolutionary diversification for the mega-diverse biota of lowland Amazonia. Uplift of structural arches during the Paleogene and Neogene resulted in the formation of the modern continental drainage configuration. Base map created by Paulo Petry from the Shuttle Radar Topography Mission with elevations in meters. Note that the scale exaggerates differences at lower elevations. Adapted from *Albert, Petry & Reis (2011)*. Full-size [DOI: 10.7717/peerj.5644/fig-3](https://doi.org/10.7717/peerj.5644/fig-3)

integrate surface uplift and climatic changes as a function of speciation and extinction include work on the Andean bellflowers (*Lagomarsino et al., 2016*), Neotropical orchids (*Pérez-Escobar et al., 2017*), and Neotropical hummingbirds (*Condamine et al., 2018*).

The Andean uplift affected Neotropical regions in different ways. Over the course of the Miocene, it led to a humidification of Amazonia and aridification of Patagonia (Cione *et al.*, 2005; Blisniuk *et al.*, 2005; Palazzesi *et al.*, 2014; Rohrmann *et al.*, 2016). This contrast is reflected by the history of New World monkeys (Platyrrhini), whose geographic expansion and morphological and taxonomic diversification is tightly linked with climatic changes (Silvestro *et al.*, *in press*). Platyrrhines were once widely distributed in Patagonia from early to middle Miocene, including the southernmost non-human primates that have ever lived (Tejedor *et al.*, 2006; Novo *et al.*, 2017). However, those primates were later extirpated during regional aridification and global cooling after the Middle Miocene. The platyrrhine record of the high Chilean Andes indicates that the connection between Patagonia and the northern Neotropics possibly persisted on the western part of South America, as the southern Andean cordillera was not an important barrier in the Middle Miocene (Flynn *et al.*, 1995). This scenario provided primates and other animals with a migration route to the north, facilitating faunal turnover (Tejedor & Muñoz-Saba, 2013). This connection may also have contributed to the subsequent Amazonian diversification of crown platyrrhines, including some Patagonian lineages (Rosenberger *et al.*, 2009).

Advances on climatic reconstructions via historical records and climatic modeling (Cheng *et al.*, 2013; Wang *et al.*, 2017) or biome palaeo-distribution modeling (Carnaval & Moritz, 2008; Werneck *et al.*, 2011, 2012a; Ledo & Colli, 2017) allow for direct hypothesis testing based on independent biodiversity data.

THE FOUR SCALES OF BIODIVERSITY RESEARCH

In the previous section, we urged for a broader integration *across* the scientific disciplines. We exemplified “trans-disciplinary biogeography” by showcasing the strong links that exist, for instance, between geological and biological fields. Now, we wish to deepen the discussion *within* biodiversity research in a more traditional sense. We do this by discussing and contrasting the current and potential levels of interaction across four sub-disciplines that span the taxonomic, temporal and spatial scales: (1) community ecology, (2) phylogeography, (3) phylogenetics, and (4) historical biogeography (Fig. 4).

COMMUNITY ECOLOGY

A major question in the study of biodiversity in general, and Neotropical research in particular, is how ecological communities have been assembled over time and how abiotic factors and species interactions have influenced this process. Approaches for the study of biodiverse communities have employed a wide range of models with diverse conceptual roots. Over the last 20 years, there has been an expansion from studies focusing on contemporary community structure and spatial patterns of physical properties of ecosystems (Tuomisto, Ruokolainen & Yli-Halla, 2003; Heithaus, 1979; Gentry, 1988; Duellman, 1989; Tuomisto *et al.*, 1995; Kalko & Handley, 2001; Garnier *et al.*, 2004), to studies focusing on historical aspects of community structure and evolution (Leite & Rogers, 2013; Smith *et al.*, 2017). Early approaches (Margalef, 1963) focused on indices of diversity, descriptions of community membership, as well as flow charts of energy and

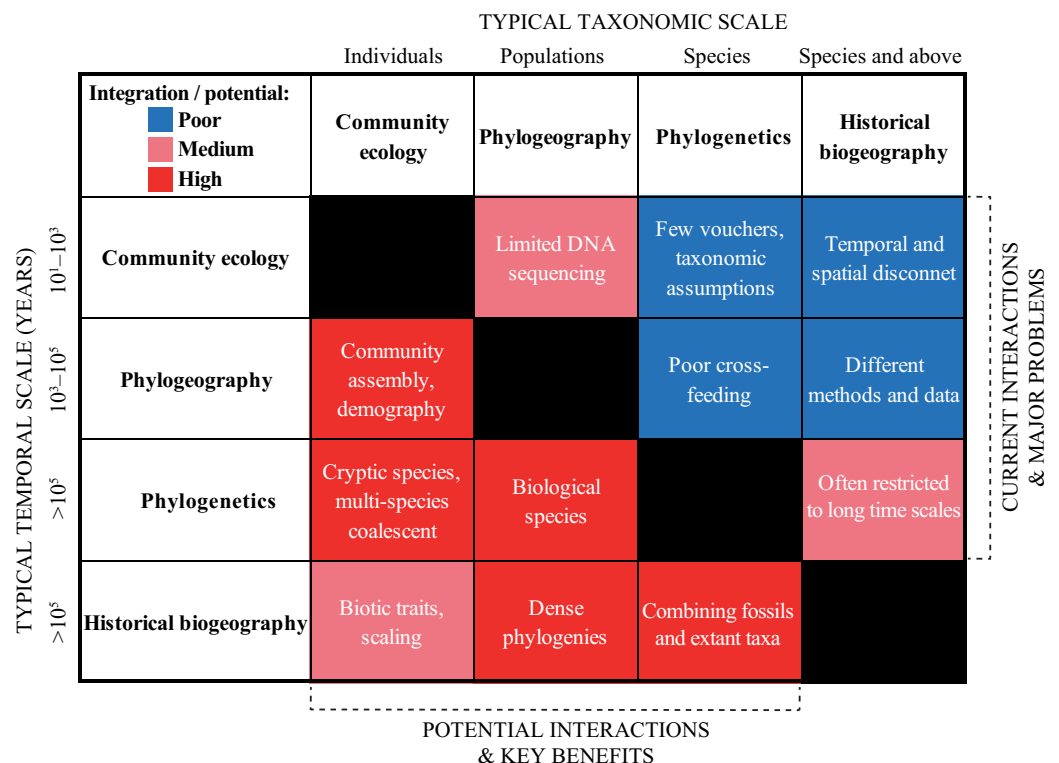


Figure 4 Heat map summarizing the current (right upper boxes) and potential (left bottom boxes) interactions across the biodiversity disciplines in the Neotropics. The X- and Y-axes indicate the typical taxonomic and temporal scales covered by each discipline, respectively. The white text in each box provides some short examples of why the disciplines are not yet successfully integrated, and some of the key benefits that will be gained by a further integration. See text for a discussion.

Full-size DOI: 10.7717/peerj.5644/fig-4

nutrients through the community. Key variables influencing community structure consistently emphasized classic Hutchinsonian processes such as resource use, competition, and niche partitioning. However, as ecologists adopted new techniques, the resolution of the niche increased from simple variables to also include high-resolution data on climate, soil chemistry, microbiomes, and other physical and biological properties. These approaches have guided several recent analyses of tropical groups, including microorganisms and plants (Tuomisto *et al.*, 2003; Costa *et al.*, 2009a; Mendes *et al.*, 2015; Arellano *et al.*, 2016; Tuomisto *et al.*, 2016). Additionally, the availability of spatially explicit online global datasets of climate and environmental parameters has helped spawn a generation of studies using large-scale spatial biodiversity surveys and inventories. When analyzed with statistical approaches, these data allow the interpolation between sampled sites and estimation of diversity in non-sampled areas (Costa *et al.*, 2007, 2009b; Ter Steege *et al.*, 2011, 2013).

Niche-based studies

The general idea that species are adapted to their environment (i.e., have different niches) has two important consequences. First, species distributions are expected to reflect the

distribution of suitable habitats. Second, species composition in local communities should reflect the environmental characteristics of the site, as unsuitable environmental characteristics or biotic interactions make it impossible for a species to establish and/or survive. Along these lines, many studies have aimed to characterize the edaphic associations of tropical plant species (Tuomisto & Poulsen, 1996; Phillips et al., 2003; Costa, Magnusson & Luizao, 2005; Roncal, 2006; Zuquim et al., 2009; Kristiansen et al., 2012; Tuomisto et al., 2016; Cámara-Leret et al., 2017; Figueiredo et al., 2017) and the elevational ranges of many taxa (Kluge, Bach & Kessler, 2008). If there are more species adapted to some environmental conditions than others and dispersal is generally not a limiting factor, a species richness gradient should result. However, it is also possible that some environmental conditions may allow more species to coexist than others. Several studies have analyzed species richness gradients along environmental gradients such as elevation (Kluge, Kessler & Dunn, 2006; Brehm, Colwell & Kluge, 2007), rainfall (Clinebell et al., 1995; Esquivel-Muelbert et al., 2017), and soil fertility (Costa, Magnusson & Luizao, 2005; Ter Steege et al., 2006; Tuomisto, Zuquim & Cárdenas, 2014). In general, these studies have shown that Neotropical plant species richness tends to be highest in warm, humid, and aseasonal environmental niches at low to middle elevations.

Neutral and non-neutral perspectives

In contrast to niche-based processes, spatial patterns in the abundance of anurans from Central Amazonia have been shown to conform to the expectations of Hubbell's neutral theory of biodiversity and biogeography (NTBB; Hubbell, 2001; Diniz-Filho et al., 2011). More recently, a study demonstrated that the incorporation of population genetic dynamics into NTBB support the hypothesis that biodiversity dynamics are out of equilibrium (Manceau, Lambert & Morlon, 2015). Additional research is needed to assess the relative roles of niche constraints, neutral, and non-neutral processes, in explaining and predicting Neotropical biodiversity.

Ecological interactions

Early theoretical ecologists conceived the role of ecological interactions (e.g., herbivory, pollination, frugivory) in shaping natural communities, mainly through mechanisms of competition and predation (Hazen, 1964; Boucher, 1988). Later, theoretical ecology shifted to a broader perspective, when facilitation (i.e., positive interactions such as mutualism and symbiosis) was envisioned as a mechanism that affects processes in both population and community levels (Bruno, Stachowicz & Bertness, 2003). This broad spectrum linked many ecological concepts and type of data (such as geo-referenced occurrences and DNA sequences), creating a multi-layer framework for investigating macro-evolutionary processes and patterns.

The use of multi-layered data have shed light on the role of climate gradients in pollinator turnover (Correa Restrepo et al., 2016), the role of frugivory traits in palm diversification (Onstein et al., 2017) and demographic and spatio-temporal distribution of species interactions (Beck, 2006). Coupling time-calibrated phylogenetic and ecological data of ant-plant interactions in the Neotropics also allowed the reconstruction of the

geographical origin of the Acacia-ant interaction ([Gómez-Acevedo et al., 2010](#)) and the identification of ecological and macro-evolutionary patterns in ant symbioses ([Chomicki, Ward & Renner, 2015](#)). In addition, phylogenetic and network analyses disclosed that specialized pollination interactions can display asymmetrically dependent diversification ([Ramírez et al., 2011](#)), revealing that specialized interactions might dilute the ecological signal in macro-evolutionary processes. Furthermore, comparative phylogenetic analyses using multi-layered data suggest that mutualistic interactions drive the relative higher diversification rates of frugivorous bats in some Neotropical regions ([Rojas et al., 2012](#)) and highlight the putative role of bat seed dispersal in shaping species-rich meta-communities.

PHYLOGEOGRAPHY

Phylogeographic research based on dense geographic and molecular sampling at the intraspecific level (subspecies and populations) has advanced significantly in the Neotropics. Specifically, phylogeographic studies of widespread species has improved our understanding of genetic diversification across various biomes. It might be intuitive to assume that species are confined to particular biomes, so that most phylogeographic studies should be done at such spatial scale, but this might not necessarily be the case. We addressed this possibility by synthesizing the recent data presented in [Antonelli et al. \(2018\)](#), where all available Neotropical species in six major clades were codified as present or absent in ten broadly defined biomes or regions: Andean Grasslands, Amazonia, Atlantic Forests, Caatinga, Cerrado and Chaco, Dry Northern South America, Dry Western South America, Mesoamerica, Patagonian Steppe, and West Indies. Our compilation shows that a substantial proportion of species in each clade occurred in more than a single biome or region: angiosperms (27,875 species or 36% of the total number of species analyzed), frogs (232 spp., 17%), birds (1,440 spp., 43%), ferns (1,529 spp., 40%), mammals (530 spp., 42%), and squamates (482 spp., 23%). Clearly, cross-biome transitions have taken place at various taxonomic levels, including populations and species.

Examples of phylogeographic studies across biomes include investigations of major vegetation transitions between Amazonia and the Cerrado ([Gehara et al., 2014](#); [Melo et al., 2018](#)) and across biomes with closer functional affinities, such as those of the dry diagonal ([Werneck et al., 2012a](#)). Another recent study revealed phylogeographic patterns of disjunctly distributed taxa, which led to inferences on the past connectivity among biomes ([Thomé et al., 2016](#); [Prates et al., 2016b, 2016a](#)). It should be noted, however, that at least some of the species of apparently widespread distribution across multiple biomes may in fact not have left their preferred habitat—such as Amazonian species being found along gallery forests in the Cerrado (see Discussion in [Antonelli et al., 2018](#)).

Comparative phylogeographic approaches can lead to robust inferences of lineage diversification, and even challenge traditional allopatric scenarios, as has been shown for Neotropical rainforest birds ([Smith et al., 2014](#)), or for the synchronous demographic expansion detected for the xeric Caatinga herpetofauna ([Gehara et al., 2017](#)). In the Neotropics, hierarchical approximate Bayesian computation analyses ([Hickerson, Stahl &](#)

Lessios, 2006) have been a popular option to reconstruct patterns of shared phylogeographic history (*Carnaval et al., 2009; Prates et al., 2016a; Gehara et al., 2017*). More recently, a paradigm shift has been proposed for the field of phylogeography, arguing for a focus placed on trait-based hypothesis testing rather than the more traditional approach of concordance amongst taxa (*Papadopoulou & Knowles, 2016*). Although this is a relatively new advance in the field, it may gain popularity, considering the general lack of concordance in the distribution of many taxa and the possible range of underlying causes. For instance, a recent study of Neotropical fishes showed that population structure is not concordant amongst palaeodrainages, but rather reflects the fundamental differences in riverine history (*Thomaz, Malabarba & Knowles, 2017*).

PHYLOGENETICS

The explosion of molecular phylogenetics and dating analyses allow the inference of time-calibrated trees, where branch lengths are measured as units of time or rates of molecular evolution. Fortunately, a massive increase in the availability of genetic information is being driven by high throughput sequencing technologies. Novel genomic data are likely to significantly improve our understanding of genetic diversity and evolutionary relationships among and within species (*Jin & Chakraborty, 1994*). Furthermore, these data will also greatly improve our understanding of largely under-studied groups, such as soil microbes (*Mahé et al., 2017*).

The integration of time-calibrated trees into phylogeographic and biogeographic analyses now enables the establishment of links with external sources of temporal information such as landscape evolution, geological history, fossil record, and climate history. Therefore, phylogenies constitute the strongest and most concrete “bridge” across scales and disciplines as outlined here (*Fig. 4*). Given the complexity and challenges with the reconstruction of reliable phylogenetic trees, we refer to an accompanying review in this volume (*Bravo et al., 2018*).

HISTORICAL BIOGEOGRAPHY

Single clade approaches

Detailed reconstructions of the temporal and spatial evolution for individual clades are obtained through “single clade” approaches. These approaches focus on contingencies or events that are idiosyncratic to the group under study, instead of generalities across groups. Methodological advances in single lineage approaches have undergone major developments with parametric methodologies (*Ree & Smith, 2008; Lemey et al., 2009; Landis et al., 2013; Matzke, 2014; Landis, 2017; Table 7*).

Typical biogeographic analyses use time-trees and parametric models of biogeographic evolution to estimate ancestral ranges of lineages (branches) and speciation events (nodes), and to infer rates of biogeographic processes (e.g., dispersal, speciation, and extinction). To date, probably hundreds of studies have examined the biogeographic history of particular Neotropical clades in this way. Biogeographic hypotheses or models about the relative role of biogeographic processes in the geographic evolution of particular groups can be compared statistically using methods for model selection in phylogenetics,

Table 7 Methodological challenges and advances for estimating biogeographic histories.

Inferring the spatial and temporal dimensions of evolution are fraught with difficulties, especially due to a lack of abundant and evenly sampled biological and geological data. This is particularly critical for the Neotropics, due to the region's immense size, relatively limited access, extraordinary biodiversity levels, landscape heterogeneity, and complex evolutionary and geo-climatic histories. To tackle these problems, we summarize some of the main issues associated with the analyses of biogeography and diversification, focusing on how those issues affect the inference of geographic range evolution of lineages in the Neotropics.

Definition and use of areas for analyses. Defining units of study in biogeography is not an easy task, especially when diverse systems are involved such as the Neotropics. Sympatry, or the geographic congruence among the distribution areas of taxa, is often used as a criterion to define units for these studies. The identification of such areas has long been based on expert opinion, with data-driven approaches that use actual species distribution data only becoming available more recently (Holt et al., 2013; Vilhena & Antonelli, 2015; Edler et al., 2016; Antonelli, 2017a, 2017b). These approaches to bioregionalization allow for more objective and reproducible analyses. Areas have also been defined using geologically explicit criteria, including information on the geological history of landmasses or geographic barriers, both of which are not exclusive to the group under study (Antonelli et al., 2009; Albert et al., 2011; Töpel et al., 2016; Bacon et al., 2018). Areas defined based on species distribution patterns and geological history are of particular interest (Perret et al., 2007; Givnish et al., 2014; Tagliacollo et al., 2015b).

The use of areas as discrete entities is useful in parametric biogeographic models where areas are considered as traits that evolve along the phylogeny, and whose ancestral areas are estimated at speciation events (nodes). In these models, the spatial units of analysis are defined by the biogeographic hypothesis under examination. For example, it is possible to determine whether diversification rates have been historically higher in Andean or non-Andean taxa (Chazot et al., 2016). However, defining areas as discrete entities is difficult when there are overlapping boundaries and an excess of widespread taxa. Models have been proposed to objectively define areas of endemism by overlapping taxa with "fuzzy" boundaries (Szumik et al., 2002; Szumik & Goloboff, 2004). Similarly, biotic element analyses have also been proposed to test for non-random distributions of species ranges (Hausdorf & Hennig, 2003). Some of these methods have been applied to Neotropical taxa (Casagrande, Roig-Juñet & Szumik, 2009; Guedes, Sawaya & Nogueira, 2014; Nogueira-Urbano & Escalante, 2015; Azevedo, Valdujo & Nogueira, 2016).

Alternatives to the use of areas. One possibility is to use geographic barriers, rather than areas, as units of analysis, thus focusing on vicariance (Hovenkamp, 1997; Arias, Szumik & Goloboff, 2011; Arias, 2017). This approach explicitly introduces the spatial (landscape) aspect missing from the predefined areas-as-discrete entities used in parametric biogeography. Since this approach is based on taxon-defined ranges, biogeographic reconstructions are not dependent on different area definitions (Arias, 2017). A parametric version of this approach allows geographic (dispersal) barriers to evolve over time within the landscape, something that has been particularly useful for understanding the biogeographic history and evolution of Neotropical freshwater fishes (Albert et al., 2017).

An additional alternative to using discrete areas in biogeographical analyses is the spatial diffusion approach, which conducts spatial-temporal reconstructions under random walk models within likelihood (Lemmon & Lemmon, 2008) or Bayesian (Lemey et al., 2010) frameworks. This approach has been used to study taxa from dry Neotropical biomes (Werneck et al., 2011, 2012a; Nascimento et al., 2013; Camargo et al., 2013), and taxa with broad continental distributions (Gehara et al., 2014). A further development of this approach has been applied to the Neotropical bird genera *Psophia* and *Cinclodes* (Quintero et al., 2015). The method uses georeferenced point-localities to infer ancestral areas and thus does not make assumptions about species ranges and operational units that fit many taxa. On the other hand, this method suffers from the common issue of ancestral lineages occupying average values of the descendant lineages. For instance, analyses with this method have reconstructed the ancestral of *Cinclodes* ovenbirds to a region in-between the western and eastern margins of South America, where no such species occur today (Quintero et al., 2015). Considering the complex and dynamic nature of the Neotropical region, diffusion analyses would certainly benefit from the incorporation of landscape-explicit models that allow the reconstruction of actual paths along branches and fossil-informed diffusion approaches (McRae et al., 2008; Meseguer & Condamine, 2018). These developments would allow the incorporation of spatial heterogeneity via dispersal constraints, derived from estimated ecological niche models or landscape evolution models.

Estimating geographic evolution on single clades. Dispersal-extinction-cladogenesis (DEC) is likely the most popular parametric biogeographic method for estimating the geographic evolution of lineages within a particular clade. This likelihood-based method infers anagenetic evolution (i.e., along branch internodes) as a function of two rate parameters: range expansion (dispersal) and range contraction (local extinction). Cladogenetic evolution (i.e., at speciation nodes) is modeled as the likelihood of alternative range inheritance scenarios that describe the division of ancestral ranges into descendant nodes: sympatric speciation or allopatric (vicariance) speciation, and peripheral isolate speciation in the case of widespread ranges (Ree & Smith, 2008).

The popularity of DEC is based on the fact that, given a time tree and associated terminal distributions, it can provide detailed biogeographic reconstructions of the ancestral origin of a clade and the history of dispersal and extinction events that shaped its spatial evolution (Sanmartín & Meseguer, 2016). A potential drawback of DEC is, however, the number of areas that it can implement. A large number of unit areas rapidly leads to computational and convergence issues. Constraining the number of states based on biological or geological criteria is a way to decrease model complexity (Ree & Sanmartín, 2009).

Bay-area, a data augmentation approach based on stochastic mapping that extends the DEC model to deal with a large number of unit areas, has been proposed to tackle the limited number of areas allowed in DEC (Landis et al., 2013). Another extension of DEC is the DEC+J model, which introduces an extra parameter ("J") to model "jump dispersal" or "founder-event" speciation (Matzke, 2014). The DEC+J model has been used in Neotropical biogeography (Matos-Maraví et al., 2014; Chomicki & Renner, 2015; Espeland et al., 2015), but was recently criticized due to statistical bias (Ree & Sanmartín, 2018). Recently, a new extension was introduced to allow for a time-heterogeneous dispersal process in a Bayesian framework, the "epoch model" (Landis, 2017). This model can be used in the biogeographic dating of speciation events when no fossil or other calibration method exists, and was recently applied to Neotropical cycads (Said Gutiérrez-Ortega et al., 2017).

such as the Akaike information criterion or Bayes factors ([Bozdogan, 1987](#)). Moreover, the rates of these processes may be modified (scaled) to reflect the changing connectivity among the areas of analysis over time ([Ree & Smith, 2008](#)). These advances have contributed to the integration of landscape dynamics and geological history into taxon biogeography in the Neotropics ([Givnish et al., 2014](#); [Palazzesi et al., 2014](#); [Tagliacollo et al., 2015b](#); [Chazot et al., 2016](#)).

Cross-taxonomic (multi-clade) approaches

These approaches (sometimes under the umbrella of “comparative biogeography”; [Antonelli, 2017b](#)) aim to extract generalities on the evolution of a biogeographic region or whole biota, or generalities on the relationships among biogeographic regions or biotas, by reconstructing the history of their individual components. The focus of this approach is on inferring shared biogeographic histories, such as general patterns of colonization and diversification or a common response to climatic and landscape changes. A recent cross-taxonomic analysis for six major clades of terrestrial plants and animals, across all major Neotropical biomes and regions, was presented by [Antonelli et al. \(2018\)](#). That study showed an unexpectedly high number of dispersal events across the entire Neotropics, which took place for tens of millions of years and often involved shifts in major environmental types (in particular from forests to savannas). The high frequency of dispersal events identified in that study reflects patterns reported for tree communities across Neotropical rain forests ([Dexter et al., 2017](#)) and between rainforests and savannas ([Simon et al., 2009](#)).

Multi-clade approaches were traditionally known as “area biogeography” and were the focus of the cladistic biogeographic school for decades ([Nelson & Platnick, 1980](#); [Humphries & Parenti, 1999](#)). The first methods used for cross-taxonomic biogeographic approaches were based on parsimony, which does not allow the formal integration of a temporal dimension ([Crisci et al., 1991](#); [Marshall & Liebherr, 2000](#); [Sanmartín, 2016](#)). The incorporation of time into event-based methods then allowed the identification of dispersal corridors and barriers, such as those underlying the assembly of freshwater fish faunas in South American river basins ([Dagosta & De Pinna, 2017](#)). Recently developed parametric approaches ([Ronquist & Sanmartín, 2011](#); [Sanmartín, 2016](#)) offer now a powerful way to obtain generalities about patterns of dispersal and diversification in biotas, allowing us to test between alternative geological or spatial scenarios ([Sanmartín, Van Der Mark & Ronquist, 2008](#)). An interesting methodology bridging community ecology and cross-taxonomic biogeographic analysis is the phylogeographic concordance factor analysis ([Satler, Zellmer & Carstens, 2016](#)), which uses Bayesian concordance analysis ([Ané et al., 2007](#)) to test for shared evolutionary history among co-distributed species and the existence of strong ecological interactions or dependence ([Satler & Carstens, 2016, 2017](#)).

BRIDGING THE CLASSICAL BIODIVERSITY DISCIPLINES

Previous sections described the complexity of Neotropical biodiversity, outlined major knowns and unknowns, proposed a general integrative framework, and discussed

approaches and applications of methods depending on the scale. Here, we provide a few examples of how to expand beyond the traditional boundaries and scales of the disciplines related to those in [Fig. 4](#).

Assembling biodiversity: from communities to biotas

The Theory of Island Biogeography (TIB; [MacArthur & Wilson, 1967](#)) introduced parameters such as rates of colonization (immigration) and extinction within a mathematical framework, allowing the prediction of the number of species present on an island based solely on its distance from a mainland species source and its area ([Losos & Ricklefs, 2009](#); [Warren et al., 2015](#)). New models inspired by the TIB are now attempting to integrate additional parameters, such as speciation and island age ([Whittaker, Triantis & Ladle, 2008](#)), population abundances ([Rosindell & Harmon, 2013](#)), and trophic interactions ([Gravel et al., 2011](#)).

Community ecology and an expanded TIB are now also adopting a more evolutionary approach by integrating phylogenetic data to the study of community assembly and dynamics, including the role of in situ adaptation or speciation vs. dispersal in community assembly, the temporal sequence of species interactions, and the role of abiotic and biotic factors in diversification of specific lineages ([Webb et al., 2003](#); [Sanmartín, Van Der Mark & Ronquist, 2008](#); [Kursar et al., 2009](#); [Valente, Etienne & Phillimore, 2014](#); [Valente, Phillimore & Etienne, 2015, 2018](#); [Cabral, Valente & Hartig, 2017](#)). Importantly, this requires denser voucher sampling of specimens, which in turn will lead to denser phylogenies, better estimates of species boundaries under multi-species coalescent approaches and tackling the common problem of cryptic species ([Fig. 4](#); [Bravo et al., 2018](#)). By adopting a more historical focus, community ecology methods are explicitly trying to reconstruct the sequence of events leading to modern-day communities, such as the island-like patches of white-sand savannas in Amazonia ([Alonso, Metz & Fine, 2013](#)). These approaches relax the assumption of ecological neutrality, and focus on the distinctive properties of individual lineages, historical contingency, and particularities of present-day outcomes ([Emerson & Kolm, 2005](#); [Sanmartín, Van Der Mark & Ronquist, 2008](#)). In their most recent forms, these models incorporate ecological parameters such as competition and species interactions ([Clarke, Thomas & Freckleton, 2016](#)) or landscape dynamics ([Aguilée, Claessen & Lambert, 2013](#)).

Scaling up community ecology approaches

The original goals of community ecology, as established in the early 20th century, were to predict species distributions and abundances, species richness and equitability, community productivity, food web structure, predator-prey dynamics, succession, and community assembly. However, this discipline has not yet succeeded in meeting most of these goals ([Ricklefs, 2008](#); [Ritchie, 2009](#); [Vellend, 2010](#); [Ricklefs & Jenkins, 2011](#); [Weber & Strauss, 2016](#)). The reasons are many, but may be especially associated to the non-equilibrium condition of most local assemblages, in which the effects of historical contingencies of dispersal, extirpations, and other stochastic processes override the equilibrium expectations generated by local functional processes such as predation

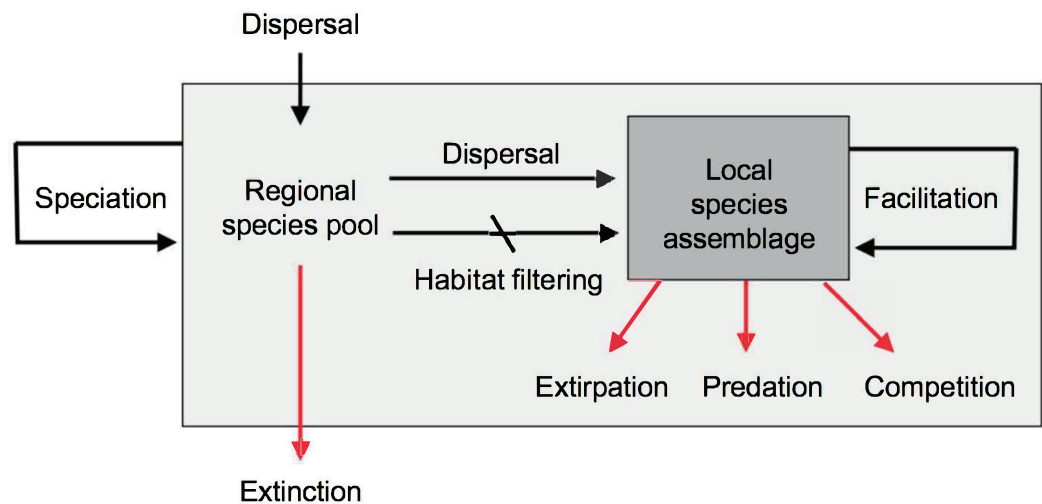


Figure 5 Main evolutionary and ecological processes contributing to the formation of species richness. The regional species pool (light gray box) is defined as the sum of all the local species assemblages (darker gray box). Black arrows indicate processes that increase species richness, red arrows processes that reduce species richness. Note the hierarchical organization of processes resulting in species richness, with evolutionary processes occurring over regional to continental spatiotemporal scales and ecological processes occurring over local scales. Speciation and dispersal contribute new species to the regional pool, while extinction removes species. Dispersal mediated by abiotic habitat filtering and biotic facilitation (Kraft & Ackerly, 2014) increase the richness of local assemblages by enhancing establishment of species pre-adapted to local conditions, or aiding in the establishment of other species. Biotic interactions such as predation and competition may serve to reduce local richness. Diagram modified from Schluter & Ricklefs (1993) and Albert, Val & Hoorn (in press). Full-size [DOI: 10.7717/peerj.5644/fig-5](https://doi.org/10.7717/peerj.5644/fig-5)

and competition (Fig. 5). In other words, the species composition and equitability of most local assemblages are more strongly governed by regional and historical factors than by local ecological interactions (Mittelbach & Schemske, 2015; Manceau, Lambert & Morlon, 2015; Fukami, 2015; Weeks, Claramunt & Cracraft, 2016).

This “crisis” in community ecology has fueled the rise of alternative functionally-neutral theories, like the NTBB (Hubbell, 2001), and the metacommunity theory (Leibold et al., 2004). However, neutral theories have also been criticized for their simplistic assumptions and lack of predictive power under the non-neutral conditions frequently observed in nature (McGill et al., 2006). In general, the field of community ecology appears to be ripe for a paradigm shift (DeAngelis & Grimm, 2017).

While many studies conducted at continental to global scales aim to test broad hypotheses about drivers of biodiversity gradients (Tuomisto, Zuquim & Cárdenas, 2014; Fine, 2015), others rely on analyses of region-wide field data collected over decades (Ter Steege et al., 2013). These surveys set the stage for analyses on the environmental and historical correlates of diversity (Benavides et al., 2005; Stropp, Ter Steege & Malhi, 2009; Ter Steege et al., 2013). Detailed explanations of the heterogeneity found at multiple scales remain a major challenge.

One recent topic of concern is whether Neotropical biodiversity patterns documented today have resulted from purely “natural” processes, or have been largely influenced by human activities (Levis et al., 2017). Evidence from archaeological, remote sensing, biodiversity data, and modeling approaches suggest that humans may have had a much

Table 8 Human impacts on Neotropical biodiversity.

Humans have occupied the Neotropics since about the end of the Late Pleistocene (10–20 kya) and were likely instrumental in promoting the extinction of the diverse fauna of large-bodied mammals ([Sandom et al., 2014](#)). The drastic decrease in the density and diversity of large mammals also resulted in major changes to the overall vegetation structure ([Bond, 2005](#)). For example, in South America, the limits between the dry diagonal and the adjacent forests might have shifted significantly compared to where they would have been without any human involvement and its cascading effects ([Doughty, Faurby & Svenning, 2016](#)). In addition to anthropogenic extinctions, humans might also have caused drastic range contractions of many other species, and reduced the abundances of others ([Faurby & Svenning, 2015](#)). The human-linked reduction of the Neotropical megafauna may also have affected the plants that these animals dispersed. This pattern was recently discussed in the context of the impact of over-hunting of primates and tapirs on the total woody biomass of Amazonia ([Peres et al., 2016](#)), and large frugivorous mammals in the Atlantic forest ([Bello et al., 2015](#)). Overall, it seems that the patterns observed could reflect the pressures of overhunting in pre-historic times (before 500 years ago). Humans have restricted the ranges of some species, but actively or passively expanded the ranges of others, such as invasives or domesticated species ([Levis et al., 2017, 2018](#)). Knowledge to date is based on the best-studied groups and it remains unclear whether substantial effects of humans might be frequent among other organisms. We anticipate that this will become an active area of research for the coming years.

Apart from the effects of past human activity on Neotropical biodiversity, current habitat loss, climate change and neglected conservation strategies pose increasingly serious threats to natural landscapes. Indeed, these are widely known to be the primary drivers of the current global biodiversity crisis. Studies that quantify genetic diversity, vulnerability, and extinction risk derived from the impact of habitat loss and climate change are essential to grasp how current human activities are expected to impact the future of Neotropical diversity at multiple levels. Although we now have a fair understanding of several components of Neotropical biodiversity, for many taxonomic groups, well-defined processes remain elusive and biases loom large; refining these issues will constitute an area of active scientific exploration for the next decade and beyond.

deeper impact on Neotropical biodiversity, both in time and space, than traditionally conceived ([Table 8](#)).

Exploring the tripod: ecological interactions-macroevo- biogeography

The multi-layer analytical framework developed by recent eco-genetic research granted the combination of multiple data layers from three different scale dynamics (local mechanisms, macro-evolutionary processes, geographic patterns). The synergy of those layers illustrates a tripod that gathers ecological, evolutionary and biogeographical factors of populations, communities, and meta-communities, respectively ([Ricklefs & Jenkins, 2011](#); [Hanson et al., 2012](#); [Connolly et al., 2017](#)). Detecting ecological signals across multi-layered data, such as the contribution of mutualisms in biogeographic processes (speciation, extinction, migration) remains a major challenge. Tackling this challenge will require linking spatio-temporal data with models that detect common signatures of ecological interactions across layers ([Pilosof et al., 2017](#)). Although recent theoretical advances have unveiled phylogenetic signals from community processes ([Rezende, Jordano & Bascompte, 2007](#); [Minoarivelo et al., 2014](#); [Bastazini, 2017](#)), we urge for new models that can identify ecological signal from multiple layers. The exploration of ecological factors that are associated to positive and negative interactions (i.e., network structure, taxonomic associations) might reveal important insights on the dynamics and complexity of ecological interactions for producing and maintaining Neotropical biodiversity.

Incorporating fossils into biogeography

One important shortcoming of molecular-based biogeographical analyses in general, and parametric models of range evolution in particular, is the fact that it is almost always based on extant data alone. Because of the effects of extinction, the pattern of geographic distribution we observe today may be a poor representation of the actual

biogeographic history, especially if extinction rates have been unequal among areas (Sanmartín & Meseguer, 2016) and taxa (Silvestro *et al.*, 2016). One way to solve this issue is to include extinct lineages in biogeographic analyses (Mao *et al.*, 2012), or to use their past distribution inferred from the fossil record to constrain inferences of ancestral ranges (Meseguer *et al.*, 2015). This approach has often revealed different biogeographic histories for the study group as compared to analyses based on extant data only (Mao *et al.*, 2012; Meseguer *et al.*, 2015). A recent development is the development of the dispersal-extinction-sampling model, to infer rates of dispersal and area extinction exclusively from fossil data (Silvestro *et al.*, 2014, 2015). Under this approach, a separate sampling parameter is used to account for the unevenness of the fossil record both spatially and temporally. When the fossil record is sufficiently abundant, it provides more accurate measures of changes in rates of geographic evolution and less biased extinction rates, than when exclusively extant taxa are used (Silvestro *et al.*, 2015).

Another challenge to understanding current patterns of evolutionary diversity is the absolute dating of phylogenies, which relies heavily on fossils. This shortcoming complicates a detailed understanding of the ages of tropical taxa, especially those from rainforests (Wing *et al.*, 2009). New methodological developments to directly integrate fossil (extinct) lineages into phylogeny reconstruction (Ronquist *et al.*, 2012; Heath, Huelsenbeck & Stadler, 2014; Zhang *et al.*, 2016; Silvestro *et al.*, 2016) offer new hope in the quest to retrieve more accurate depictions of evolutionary patterns.

Finally, estimating the tempo of diversification is difficult without fossil constraints. In simulated phylogenies, the resulting shape of lineage-through-time plots vary significantly when the fossil record is added as compared to phylogenies that incorporate extant taxa exclusively (Matos-Maraví, 2016; Sanmartín & Meseguer, 2016). The inferred macroevolutionary dynamics estimated from molecular phylogenies may thus be misleading if fossil taxa are neglected, or when macroevolutionary tools do not acknowledge the rare sampling of fossil lineages. Clearly, fossils are crucial to not only understand past dynamics, but also for an improved understanding of current patterns (Fritz *et al.*, 2013). We therefore urge for a much tighter integration between the palaeontological and neontological research communities in the Neotropics.

Integrating landscape evolution models into biotic diversification

A potential problem with single clade and cross-taxonomic biogeographic analyses as discussed above is that areas are treated as traits of organisms evolving along phylogenetic trees. Geology is often used to inform the model but does not form its core. For instance, area connectivity is often used in parametric methods to constrain or scale migration rates but not as an actual part of the model. A new generation of methods that use the power of LEMs to study the full panoply of evolutionary processes, at both microevolutionary (population) level (Byrne & Hopper, 2008; McRae *et al.*, 2008) and macroevolutionary (interspecific) scales (Tagliacollo *et al.*, 2015b; Badgley *et al.*, 2017) are now being developed. For example, uplift of a dissected landscape and river capture are two landscape evolution processes with great power to generate high species richness (Albert *et al.*, 2018). Both of these processes simultaneously and continuously merge

and separate portions of adjacent landscape areas, allowing biotic dispersal and larger geographic ranges, vicariant speciation and smaller geographic ranges, and extinction when range sizes are subdivided below a minimum persistence threshold (*Albert et al., 2017*).

CONCLUSIONS

The origins of Neotropical biodiversity

There are often mixed definitions and questions related to the timing and mode of biotic evolution in the Neotropics. The “origin of Neotropical biodiversity” encapsulates at least two contrasting subjects: the timing of origin of hyperdiversity (i.e., “when did the Neotropics reach globally outstanding levels of species richness?”), and the actual age of extant species (i.e., “when did the species that we see today split from their most recent common ancestors, as defined by their stem age?”; *Hoorn et al., 2011*). It is clear that there have been unusual periods of time throughout geological history, both in terms of biotic and abiotic events (*Jaramillo, 2006; Hoorn et al., 2010b; Jaramillo et al., 2010*). However, all periods of time have contributed to the current biodiversity, if seen in the perspective of the “evolutionary continuum” that bridges the core biodiversity disciplines (*Fig. 4*).

Examples of studies that have sought for “special” periods of time often come from time-calibrated molecular phylogenies. For instance, butterfly species-pairs seem to be relatively young in origin (i.e., <2 Ma), suggesting that the Pleistocene and Holocene may have represented “extraordinary times” for Neotropical butterfly speciation (*Garzón-Orduña, Benetti Longhini & Brower, 2014*) and following the refugium theory (*Table 4*). However, time-calibrated phylogenies may not fully address the potential impact of extinction and species duration (*Hoorn et al., 2011*). In other words, if we were able to travel back in time to any period and sequence species around us, the odds are that most species alive might also be around two Ma old or less. In addition, the definition of “species” may vary considerably depending on data source (e.g., based on the fossil record and extant populations) and across taxa. Highly structured populations with considerable genetic divergences may be seen as “incipient species” that have not yet completed the speciation process (*Craig et al., 2018*). Variable species concepts and adequate sampling of extant and extinct taxa represent a serious barrier for our understanding of Neotropical diversification (*Tables 1–3 and 5*).

Advancing Neotropical research

Comparative biology has experienced major advancements in the theory and practice of biogeography and molecular phylogenetics during the past decades. However, we still need to increase sampling of organisms drastically in order to advance our knowledge on the patterns and processes underlying Neotropical biodiversity (*Feeley, 2015*). However, fieldwork in the Neotropics, especially in pristine areas, is time consuming and logistically demanding. Research funding for exploratory inventory projects is also becoming increasingly harder to obtain, despite the fact that highly successful projects (i.e., sequencing the first human genome and creating the Amazon Tree Diversity

Network) were initially discovery-driven, rather than focused on testing specific hypotheses. Furthermore, obtaining permits to collect and export biological samples is also challenging, involving many differences across national legislations. Finally, some authors have seen the need of fieldwork as less relevant in this era of museomics (Buerki & Baker, 2016; Zedane et al., 2016).

Despite all these obstacles, fieldwork remains absolutely essential for biodiversity data generation and monitoring (Albert, 2002). Fieldwork also provides students and researchers with a deeper understanding of their study systems, often providing new ideas and questions, while facilitating the establishment of new collaborations, enabling the exchange of knowledge, fueling the development of new methods, and increasing the possibilities of major discoveries (Fleischner et al., 2017). We should seriously consider new strategies for the generation of new biodiversity data, as well as for the syntheses of the already available data.

Multi-taxon field campaigns could provide unique opportunities for intensive sampling, while optimizing resources, bureaucratic, and logistic efforts. However, this vision requires a radical re-thinking and re-organization. We need to provide young generations with the training, tools and resources needed to carry out research on all aspects of biodiversity. We also need to support taxonomic specialists and institutions in order to adequately study, archive and facilitate free access to biological collections and associated data. In addition, we also need to join forces across nations and disciplines for mutual benefit and joint scientific growth. Clearly, these investments would be worthwhile from a global perspective. The future of Neotropical biodiversity research depends on extensive collaborations and coordinated efforts.

TAKE-HOME MESSAGES

Five main take home messages can be taken from this review, namely:

1. Neotropical biodiversity is exceedingly high, regardless of the axis explored (e.g., taxonomic, phylogenetic, functional, ecosystems);
2. Understanding the origins, evolution, maintenance, and distribution patterns of Neotropical biodiversity is a grand scientific challenge with many remaining unknowns;
3. “Trans-disciplinary biogeography” aims to better integrate the seemingly disparate disciplines required to explore the biotic and abiotic evolution of the Neotropics;
4. Many methodological advances will be required to deal with the increasing wealth of biodiversity data and associated environmental and geological variables;
5. There is an urgent need to fill the many gaps in biodiversity knowledge, including extant and extinct taxa and their interactions. This calls for a “renaissance” for fieldwork.

ACKNOWLEDGEMENTS

This paper is a product of the “Origin of Biodiversity Workshop” organized by Chalmers University of Technology and the University of Gothenburg, under the auspices of the Gothenburg Centre for Advanced Studies (GoCAS). We are particularly grateful to GoCAS organizers Karin Hårding, Gunnar Nyman, and Mattias Marklund for their

continuous support and assistance. We thank Juan E. Guevara, Thomas J. Givnish, Luis F. Aguirre, Tomas Hrbek, Julissa Roncal, and the editor Stephen Johnson for helpful suggestions on the pre-print of this paper. We also thank Tomas Hrbek for handling and editing the peer-reviewed manuscript, and two anonymous reviewers for constructive comments.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

The workshop “Origins of Biodiversity” was funded by Chalmers University of Technology and the University of Gothenburg. The following researchers are supported by scholarship or research grants from the following agencies: Alexandre Antonelli by the Swedish Research Council (B0569601), the European Research Council under the European Union’s Seventh Framework Programme (FP/2007-2013, ERC Grant Agreement 331024), the Swedish Foundation for Strategic Research, a Wallenberg Academy Fellowship, the Faculty of Sciences at the University of Gothenburg, the Wenner-Gren Foundations, and the David Rockefeller Center for Latin American Studies at Harvard University; Camila D. Ritter by the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 249064/2013-8); Daniele Silvestro by the Swedish Research Council (2015-04748); Fernanda P. Werneck by the Conselho Nacional de Desenvolvimento Científico e Tecnológico, the Fundação de Amparo à Pesquisa do Amazonas, the Partnerships for Enhanced Engagement in Research from the U.S. National Academy of Sciences (PEER NAS/USAID, USA), and the L’Oréal-UNESCO for Women in Science Awards (Brazil and France); Isabel Sanmartín by MINECO/FEDER (CGL2015-67849-P); James S. Albert by the National Science Foundation (NSF 0614334, 0741450, and 1354511); Josué A. R. Azevedo by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) (99999.001292/2015-03); Lúcia G. Lohmann by a collaborative Dimensions of Biodiversity-NSF/Biota-FAPESP grant (FAPESP 2012/50260-6) and by the Fundação de Amparo à Pesquisa do Estado de São Paulo (2017/12797-1); Pável Matos-Maraví by a Marie Skłodowska-Curie fellowship (704035); Thaís Guedes by the Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP, 2013/04170-8 and 2014/18837-7). No additional external funding was received for this study. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:
Chalmers University of Technology and the University of Gothenburg.
The Swedish Research Council: B0569601.
The European Research Council under the European Union’s Seventh Framework Programme: FP/2007-2013, ERC Grant Agreement 331024.
The Swedish Foundation for Strategic Research, a Wallenberg Academy Fellowship, the Faculty of Sciences at the University of Gothenburg, the Wenner-Gren Foundations, and

the David Rockefeller Center for Latin American Studies at Harvard University.
 Conselho Nacional de Desenvolvimento Científico e Tecnológico: CNPq 249064/2013-8.
 Swedish Research Council: 2015-04748.
 Conselho Nacional de Desenvolvimento Científico e Tecnológico, the Fundação de Amparo à Pesquisa do Amazonas, the Partnerships for Enhanced Engagement in Research from the U.S. National Academy of Sciences (PEER NAS/USAID, USA).
 The L'Oréal-UNESCO for Women in Science Awards (Brazil and France).
 MINECO/FEDER: CGL2015-67849-P.
 National Science Foundation: NSF 0614334, 0741450, and 1354511.
 Dimensions of Biodiversity-NSF/Biota-FAPESP grant: FAPESP 2012/50260-6.
 The Fundação de Amparo à Pesquisa do Estado de São Paulo: 2017/12797-1.
 Marie Skłodowska-Curie fellowship: 704035.
 Fundação de Amparo à Pesquisa do Estado de São Paulo: FAPESP, 2013/04170-8 and 2014/18837-7.

Competing Interests

Scott V. Edwards is an Academic Editor for PeerJ.

Author Contributions

- Alexandre Antonelli conceived and designed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- María Ariza prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft, formatted references and merged the manuscript revisions.
- James Albert prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Tobias Andermann prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Josué Azevedo prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Christine Bacon prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Søren Faurby analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Thais Guedes prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Carina Hoorn prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Lúcia G. Lohmann prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Pável Matos-Maraví prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

- Camila D. Ritter prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Isabel Sanmartín prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Daniele Silvestro prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Marcelo Tejedor prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Hans ter Steege prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Hanna Tuomisto prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Fernanda P. Werneck prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Alexander Zizka analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Scott V. Edwards conceived and designed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The research in this literature review did not generate any data or code.

REFERENCES

- Aguilée R, Claessen D, Lambert A. 2013.** Adaptive radiation driven by the interplay of eco-evolutionary and landscape dynamics. *Evolution* **67**(5):1291–1306 DOI [10.1111/evo.12008](https://doi.org/10.1111/evo.12008).
- Albert JS. 2002.** Eternal vigilance on an Amazon floodplain. *Current Biology* **12**(13):R442–R443 DOI [10.1016/S0960-9822\(02\)00937-5](https://doi.org/10.1016/S0960-9822(02)00937-5).
- Albert JS, Carvalho TP, Petry P, Holder MA, Maxime EL, Espino J, Corahua I, Quispe R, Rengifo B, Ortega H, Reis RE. 2011.** Aquatic biodiversity in the Amazon: habitat specialization and geographic isolation promote species richness. *Animals* **1**(2):205–241 DOI [10.3390/ani1020205](https://doi.org/10.3390/ani1020205).
- Albert JS, Craig JM, Tagliacollo VA, Petry P. 2018.** Upland and lowland fishes: a test of the river capture hypothesis. In: Hoorn C, Antonelli A, Antonelli A, eds. *Mountains, Climate and Biodiversity*. New York: Wiley-Blackwell, 273–294.
- Albert JS, Petry P, Reis RE. 2011.** Major biogeographic and phylogenetic patterns. In: Albert JS, Reis RE, eds. *Historical Biogeography of Neotropical Freshwater Fishes*. Berkeley: University of California Press, 21–57.
- Albert JS, Schoolmaster DR, Tagliacollo V, Duke-Sylvester SM. 2017.** Barrier displacement on a neutral landscape: toward a theory of continental biogeography. *Systematic Biology* **66**(2):167–182 DOI [10.1093/sysbio/syw080](https://doi.org/10.1093/sysbio/syw080).
- Albert JS, Val P, Hoorn C.** The changing course of the Amazon in the Neogene: Center stage for Neotropical diversification. *Neotropical Ichthyology* (in press).
- Alonso JÁ, Metz MR, Fine PVA. 2013.** Habitat specialization by birds in Western Amazonian white-sand forests. *Biotropica* **45**(3):365–372.

- Arrington DA, Winemiller KO. 2009. Habitat affinity, the seasonal flood pulse, and community assembly in the littoral zone of a Neotropical floodplain river. *Journal of the North American Benthological Society* 25(1):126–141 DOI 10.1899/0887-3593(2006)25[126:HATSFP]2.0.CO;2.
- Andújar C, Arribas P, Ruzicka F, Crampton-Platt A, Timmermans MJTN, Vogler AP. 2015. Phylogenetic community ecology of soil biodiversity using mitochondrial metagenomics. *Molecular Ecology* 24(14):3603–3617 DOI 10.1111/mec.13195.
- Ané C, Larget B, Baum DA, Smith SD, Rokas A. 2007. Bayesian estimation of concordance among gene trees. *Molecular Biology and Evolution* 24(7):1575 DOI 10.1093/molbev/msm107.
- Antonelli A. 2017a. Biogeography: drivers of bioregionalization. *Nature Ecology & Evolution* 1(4):0114 DOI 10.1038/s41559-017-0114.
- Antonelli A. 2017b. Comparative biogeography, big data, and common myths. In: Friis I, Balslev H, eds. *Tropical plant collections: legacies from the past? Essential tools for the future?* Scientia Danica B (Biological). Vol. 6. Copenhagen: Royal Danish Academy of Sciences and Letters.
- Antonelli A, Nylander JAA, Persson C, Sanmartín I. 2009. Tracing the impact of the Andean uplift on Neotropical plant evolution. *Proceedings of the National Academy of Sciences of the United States of America* 106(24):9749–9754 DOI 10.1073/pnas.0811421106.
- Antonelli A, Sanmartín I. 2011. Why are there so many plant species in the Neotropics? *Taxon* 60(2):403–414.
- Antonelli A, Zizka A, Antunes Carvalho F, Scharn R, Bacon CD, Silvestro D, Condamine FL. 2018. Amazonia is the primary source of Neotropical biodiversity. *Proceedings of the National Academy of Sciences of the United States of America* 115(23):6034–6039 DOI 10.1073/pnas.1713819115.
- Arbour JH, López-Fernández HS. 2014. Adaptive landscape and functional diversity of Neotropical cichlids: implications for the ecology and evolution of Cichlinae (Cichlidae; Cichliformes). *Journal of Evolutionary Biology* 27(11):2431–2442 DOI 10.1111/jeb.12486.
- Arellano G, Tello JS, Jørgensen PM, Fuentes AF, Loza MI, Torrez V, Macía MJ. 2016. Disentangling environmental and spatial processes of community assembly in tropical forests from local to regional scales. *Oikos* 125(3):326–335 DOI 10.1111/oik.02426.
- Arias JS. 2017. An event model for phylogenetic biogeography using explicitly geographical ranges. *Journal of Biogeography* 44(10):2225–2235 DOI 10.1111/jbi.13024.
- Arias JS, Szumik CA, Goloboff PA. 2011. Spatial analysis of vicariance: a method for using direct geographical information in historical biogeography. *Cladistics* 27(6):617–628 DOI 10.1111/j.1096-0031.2011.00353.x.
- Armbruster J, Van Der Sleen P, Lujan N. 2018. Family Loricariidae—Suckermouth armoured catfishes. In: Van Der Sleen P, Albert JS, eds. *Field Guide to the Fishes of the Amazon, Orinoco and Guianas*. New Jersey: Princeton University Press, 253–258.
- Azevedo JAR, Valdujo PH, Nogueira C De C. 2016. Biogeography of anurans and squamates in the Cerrado hotspot: coincident endemism patterns in the richest and most impacted savanna on the globe. *Journal of Biogeography* 43(12):2454–2464 DOI 10.1111/jbi.12803.
- Bacon CD. 2013. Biome evolution and biogeographical change through time. *Frontiers of Biogeography* 5(4):227–231.
- Bacon CD, Baker WJ, Simmons MP. 2012. Miocene dispersal drives island radiations in the palm tribe Trachycarpeae (Arecaceae). *Systematic Biology* 61(3):426–442 DOI 10.1093/sysbio/syr123.
- Bacon CD, Silvestro D, Jaramillo CA, Smith BT, Chakrabarty P, Antonelli A. 2015. Biological evidence supports an early and complex emergence of the Isthmus of Panama.

Proceedings of the National Academy of Sciences of the United States of America 112(19):6110–6115 DOI 10.1073/pnas.1423853112.

- Bacon CD, Velásquez-Puentes FJ, Hoorn C, Antonelli A. 2018. Iriarteeae palms tracked the uplift of Andean Cordilleras. *Journal of Biogeography* 45(7):1653–1663 DOI 10.1111/jbi.13350.
- Badgley C, Smiley TM, Terry R, Davis EB, DeSantis LRG, Fox DL, Hopkins SSB, Jezkova T, Matocq MD, Matzke N, McGuire JL, Mulch A, Riddle BR, Louise Roth V, Samuels JX, Strömberg CAE, Yanites BJ. 2017. Biodiversity and topographic complexity: modern and geohistorical perspectives. *Trends in Ecology & Evolution* 32(3):211–226 DOI 10.1016/j.tree.2016.12.010.
- Baker WJ, Couvreur TLP. 2012. Global biogeography and diversification of palms sheds light on the evolution of tropical lineages. I. Historical biogeography. *Journal of Biogeography* 40(2):274–285 DOI 10.1111/j.1365-2699.2012.02795.x.
- Baker PA, Fritz SC, Dick CW, Eckert AJ, Horton BK, Manzoni S, Ribas CC, Garzione CN, Battisti DS. 2014. The emerging field of geogenomics: constraining geological problems with genetic data. *Earth-Science Reviews* 135:38–47 DOI 10.1016/j.earscirev.2014.04.001.
- Basset Y, Cizek L, Cuénoud P, Didham RK, Guilhaumon F, Missa O, Novotny V, Ødegaard F, Roslin T, Schmidl J, Tishechkin AK, Winchester NN, Roubik DW, Aberlenc H-P, Bail J, Barrios H, Bridle JR, Castaño-Meneses G, Corbara B, Curletti G, Da Rocha WD, De Bakker D, Delabie JHC, Dejean A, Fagan LL, Floren A, Kitching RL, Medianero E, Miller SE, De Oliveira EG, Orivel J, Pollet M, Rapp M, Ribeiro SP, Roisin Y, Schmidt JB, Sørensen LL, Leponce M. 2012. Arthropod diversity in a tropical forest. *Science* 338(6113):1481–1484 DOI 10.1126/science.1226727.
- Bastazini VAG. 2017. Untangling the tangled bank: a novel method for partitioning the effects of phylogenies and traits on ecological networks. *Evolutionary Biology* 44(3):312–324 DOI 10.1007/s11692-017-9409-8.
- Beck H. 2006. A review of peccary–palm interactions and their ecological ramifications across the Neotropics. *Journal of Mammalogy* 87(3):519–530 DOI 10.1644/05-MAMM-A-174R1.1.
- Bello C, Galetti M, Pizo MA, Magnago LFS, Rocha MF, Lima RAF, Peres CA, Ovaskainen O, Jordano P. 2015. Defaunation affects carbon storage in tropical forests. *Science Advances* 1(11):e1501105 DOI 10.1126/sciadv.1501105.
- Bemmels JB, Wright SJ, Garwood NC, Queenborough SA, Valencia R, Dick CW. 2018. Filter-dispersal assembly of lowland Neotropical rainforests across the Andes. *Ecography* 6:1 DOI 10.1111/ecog.03473.
- Benavides AM, Duque A, Duivenvoorden JF, Vasco A, Callejas R. 2005. A first quantitative census of vascular epiphytes in rain forests of Colombian Amazonia. *Biodiversity and Conservation* 14(3):739–758 DOI 10.1007/s10531-004-3920-9.
- Bertrand Y, Pleijel F, Rouse GW. 2006. Taxonomic surrogacy in biodiversity assessments, and the meaning of Linnaean ranks. *Systematics and Biodiversity* 4(2):149–159 DOI 10.1017/S1477200005001908.
- Bickford D, Lohman DJ, Sodhi NS, Ng PKL, Meier R, Winker K, Ingram KK, Das I. 2007. Cryptic species as a window on diversity and conservation. *Trends in Ecology & Evolution* 22(3):148–155 DOI 10.1016/j.tree.2006.11.004.
- Blisniuk PM, Stern LA, Page Chamberlain C, Idleman B, Zeitler PK. 2005. Climatic and ecologic changes during Miocene surface uplift in the Southern Patagonian Andes. *Earth and Planetary Science Letters* 230(1–2):125–142 DOI 10.1016/j.epsl.2004.11.015.

- Bond WJ. 2005. Large parts of the world are brown or black: a different view on the “Green World” hypothesis. *Journal of Vegetation Science* 16(3):261 DOI 10.1658/1100-9233(2005)016[0261:lpotwa]2.0.co;2.
- Boucher DH. 1988. The idea of mutualism, past and future. *The Biology of Mutualism: Ecology and Evolution*. Oxford: Oxford University Press, 388.
- Bozdogan H. 1987. Model selection and Akaike’s Information Criterion (AIC): the general theory and its analytical extensions. *Psychometrika* 52(3):345–370 DOI 10.1007/bf02294361.
- Bravo GA, Antonelli A, Bacon CD, Bartoszek K, Blom M, Huynh S, Jones G, Knowles LL, Lamichhaney S, Marcussen T, Morlon H, Nakhleh L, Oxelman B, Pfeil B, Schliep A, Wahlberg N, Werneck F, Wiedenhoeft J, Willows-Munro S, Edwards SV. 2018. Embracing heterogeneity: building the tree of life and the future of phylogenomics. *PeerJ* 6:e26449v3.
- Brehm G, Colwell RK, Kluge J. 2007. The role of environment and mid-domain effect on moth species richness along a tropical elevational gradient. *Global Ecology and Biogeography* 16(2):205–219 DOI 10.1111/j.1466-8238.2006.00281.x.
- Brown JH, Svenning J-C. 2013. Why are there so many species in the tropics? *Journal of Biogeography* 41(1):8–22 DOI 10.1111/jbi.12228.
- Bruno JF, Stachowicz JJ, Bertness MD. 2003. Inclusion of facilitation into ecological theory. *Trends in Ecology & Evolution* 18(3):119–125 DOI 10.1016/S0169-5347(02)00045-9.
- Buerki S, Baker WJ. 2016. Collections-based research in the genomic era. *Biological Journal of the Linnean Society* 117(1):5–10 DOI 10.1111/bij.12721.
- Burns KJ, Unitt P, Mason NA. 2016. A genus-level classification of the family Thraupidae (Class Aves: Order Passeriformes). *Zootaxa* 4088(3):329 DOI 10.11646/zootaxa.4088.3.2.
- Byrne M, Hopper SD. 2008. Granite outcrops as ancient islands in old landscapes: evidence from the phylogeography and population genetics of *Eucalyptus caesia* (Myrtaceae) in Western Australia. *Biological Journal of the Linnean Society* 93(1):177–188 DOI 10.1111/j.1095-8312.2007.00946.x.
- Cabral JS, Valente L, Hartig F. 2017. Mechanistic simulation models in macroecology and biogeography: state-of-art and prospects. *Ecography* 40(2):267–280 DOI 10.1111/ecog.02480.
- Cámara-Leret R, Tuomisto H, Ruokolainen K, Balslev H, Munch Kristiansen S. 2017. Modelling responses of western Amazonian palms to soil nutrients. *Journal of Ecology* 105(2):367–381 DOI 10.1111/1365-2745.12708.
- Camargo A, Werneck FP, Morando M, Sites JW, Avila LJ. 2013. Quaternary range and demographic expansion of *Liolaemus darwini* (Squamata: Liolaemidae) in the Monte Desert of Central Argentina using Bayesian phylogeography and ecological niche modelling. *Molecular Ecology* 22(15):4038–4054 DOI 10.1111/mec.12369.
- Cardoso D, Särkinen T, Alexander S, Amorim AM, Bittrich V, Celis M, Daly DC, Fiaschi P, Funk VA, Giacomini LL, Goldenberg R, Heiden G, Iganci J, Kelloff CL, Knapp S, Cavalcante De Lima H, Machado AFP, Dos Santos RM, Mello-Silva R, Michelangeli FA, Mitchell J, Moonlight P, De Moraes PLR, Mori SA, Nunes TS, Pennington TD, Pirani JR, Prance GT, De Queiroz LP, Rapini A, Riina R, Rincon CAV, Roque N, Shimizu G, Sobral M, Stehmann JR, Stevens WD, Taylor CM, Trovó M, Van Den Berg C, Van Der Werff H, Viana PL, Zartman CE, Forzza RC. 2017. Amazon plant diversity revealed by a taxonomically verified species list. *Proceedings of the National Academy of Sciences of the United States of America* 114(40):10695–10700 DOI 10.1073/pnas.1706756114.

- Carnaval AC, Hickerson MJ, Haddad CFB, Rodrigues MT, Moritz C. 2009. Stability predicts genetic diversity in the Brazilian Atlantic forest hotspot. *Science* 323(5915):785–789 DOI 10.1126/science.1166955.
- Carnaval AC, Moritz C. 2008. Historical climate modelling predicts patterns of current biodiversity in the Brazilian Atlantic forest. *Journal of Biogeography* 35(7):1187–1201 DOI 10.1111/j.1365-2699.2007.01870.x.
- Carnaval AC, Waltari E, Rodrigues MT, Rosauer D, VanDerWal J, Damasceno R, Prates I, Strangas M, Spanos Z, Rivera D, Pie MR, Firkowski CR, Bornschein MR, Ribeiro LF, Moritz C. 2014. Prediction of phylogeographic endemism in an environmentally complex biome. *Proceedings of the Royal Society B: Biological Sciences* 281(1792):20141461 DOI 10.1098/rspb.2014.1461.
- Casagrande MD, Roig-Juñet S, Szumik C. 2009. Endemismo a diferentes escalas espaciales: un ejemplo con Carabidae (Coleóptera: Insecta) de América del Sur austral. *Revista chilena de historia natural* 82:17–42 DOI 10.4067/S0716-078X2009000100002.
- Chazot N, Willmott KR, Condamine FL, De-Silva DL, Freitas AVL, Lamas G, Morlon H, Giraldo CE, Jiggins CD, Joron M, Mallet J, Uribe S, Elias M. 2016. Into the Andes: multiple independent colonizations drive montane diversity in the Neotropical clearwing butterflies Godyridina. *Molecular Ecology* 25(22):5765–5784 DOI 10.1111/mec.13773.
- Chao A, Jost L. 2012. Coverage-based rarefaction and extrapolation: standardizing samples by completeness rather than size. *Ecology* 93(12):2533–2547 DOI 10.1890/11-1952.1.
- Cheng H, Sinha A, Cruz FW, Wang X, Edwards RL, d’Horta FM, Ribas CC, Vuille M, Stott LD, Auler AS. 2013. Climate change patterns in Amazonia and biodiversity. *Nature Communications* 4(1):1411–1416 DOI 10.1038/ncomms2415.
- Chomicki G, Renner SS. 2015. Phylogenetics and molecular clocks reveal the repeated evolution of ant-plants after the late Miocene in Africa and the early Miocene in Australasia and the Neotropics. *New Phytologist* 207(2):411–424 DOI 10.1111/nph.13271.
- Chomicki G, Ward PS, Renner SS. 2015. Macroevolutionary assembly of ant/plant symbioses: *Pseudomyrmex* ants and their ant-housing plants in the Neotropics. *Proceedings of the Royal Society B: Biological Sciences* 282(1819):20152200–20152209 DOI 10.1098/rspb.2015.2200.
- Cione AL, De Las Mercedes Azpelicueta M, Casciotta JR, Dozo MT. 2005. Tropical freshwater teleosts from Miocene beds of eastern Patagonia, southern Argentina. *Geobios* 38(1):29–42 DOI 10.1016/j.geobios.2003.08.005.
- Clarke M, Thomas GH, Freckleton RP. 2016. Trait evolution in adaptive radiations: modeling and measuring interspecific competition on phylogenies. *The American Naturalist* 189(2):121–137 DOI 10.1086/689819.
- Clinebell RR, Phillips OL, Gentry AH, Stark N, Zuuring H. 1995. Prediction of neotropical tree and liana species richness from soil and climatic data. *Biodiversity and Conservation* 4(1):56–90 DOI 10.1007/BF00115314.
- Coddington JA, Agnarsson I, Miller JA, Kuntner M, Hormiga G. 2009. Undersampling bias: the null hypothesis for singleton species in tropical arthropod surveys. *Journal of Animal Ecology* 78(3):573–584 DOI 10.1111/j.1365-2656.2009.01525.x.
- Colli GR, Fenker J, Tedeschi LG, Barreto-Lima AF, Mott T, Ribeiro SLB. 2016. In the depths of obscurity: knowledge gaps and extinction risk of Brazilian worm lizards (Squamata, Amphisbaenidae). *Biological Conservation* 204:51–62 DOI 10.1016/j.biocon.2016.07.033.
- Condamine FL, Antonelli A, Lagomarsino LP, Hoorn C, Liow LH. 2018. Teasing apart mountain uplift, climate change and biotic drivers of species diversification. In: Hoorn C, Antonelli A, Perrigo A, eds. *Mountains, Climate and Biodiversity*. New York: Wiley-Blackwell, 257–272.

- Connolly SR, Keith SA, Colwell RK, Rahbek C. 2017. Process, Mechanism, and Modeling in Macroecology. *Trends in Ecology & Evolution* 32(11):835–844 DOI 10.1016/j.tree.2017.08.011.
- Correa Restrepo Z, Núñez Avellaneda LA, González-Caro S, Velásquez-Puentes FJ, Bacon CD. 2016. Exploring palm-insect interactions across geographical and environmental gradients. *Botanical Journal of the Linnean Society* 182(2):389–397 DOI 10.1111/boj.12443.
- Costa FRC, Guillaumet J-L, Lima AP, Pereira OS. 2009a. Gradients within gradients: The mesoscale distribution patterns of palms in a central Amazonian forest. *Journal of Vegetation Science* 20(1):69–78 DOI 10.1111/j.1654-1103.2009.05314.x.
- Costa GC, Hampe A, Ledru M-P, Martinez PA, Mazzochini GG, Shepard DB, Werneck FP, Moritz C, Carnaval AC. 2017. Biome stability in South America over the last 30 kyr: Inferences from long-term vegetation dynamics and habitat modelling. *Global Ecology and Biogeography* 27(3):285–297 DOI 10.1111/geb.12694.
- Costa FRC, Magnusson WE, Luizao RC. 2005. Mesoscale distribution patterns of Amazonian understorey herbs in relation to topography, soil and watersheds. *Journal of Ecology* 93(5):863–878 DOI 10.1111/j.1365-2745.2005.01020.x.
- Costa GC, Nogueira C, Machado RB, Colli GR. 2007. Squamate richness in the Brazilian Cerrado and its environmental-climatic associations. *Diversity and Distributions* 13(6):714–724 DOI 10.1111/j.1472-4642.2007.00369.x.
- Costa GC, Nogueira C, Machado RB, Colli GR. 2009b. Sampling bias and the use of ecological niche modeling in conservation planning: a field evaluation in a biodiversity hotspot. *Biodiversity and Conservation* 19(3):883–899 DOI 10.1007/s10531-009-9746-8.
- Cowie RH, Holland BS. 2006. Dispersal is fundamental to biogeography and the evolution of biodiversity on oceanic islands. *Journal of Biogeography* 33(2):193–198 DOI 10.1111/j.1365-2699.2005.01383.x.
- Craig JM, Malabarba LR, Crampton WGR, Albert JS. 2018. Revision of banded knifefishes of the *Gymnotus carapo* and *G. tigre* clades (Gymnotidae Gymnotiformes) from the Southern Neotropics. *Zootaxa* 4379(1):47–73 DOI 10.11646/zootaxa.4379.1.3.
- Craw D, Upton P, Burridge CP, Wallis GP, Waters JM. 2016. Rapid biological speciation driven by tectonic evolution in New Zealand. *Nature Geoscience* 9(2):140–144 DOI 10.1038/ngeo2618.
- Crisci JV, Cigliano MM, Morrone JJ, Roig-Juñet S. 1991. Historical biogeography of southern South America. *Systematic Biology* 40(2):152–171 DOI 10.1093/sysbio/40.2.152.
- Cruz-Neto AP, Garland T, Abe AS. 2001. Diet, phylogeny, and basal metabolic rate in phyllostomid bats. *Zoology* 104(1):49–58 DOI 10.1078/0944-2006-00006.
- Ribeiro JEL Da S. 1999. *Flora da Reserva Ducke: guia de identificação das plantas vasculares de uma floresta de terra-firme na Amazônia Central*. Manaus: INPA.
- Dagosta FCP, De Pinna M. 2017. Biogeography of Amazonian fishes: deconstructing river basins as biogeographic units. *Neotropical Ichthyology* 15(3):1–24 DOI 10.1590/1982-0224-20170034.
- De Baets K, Antonelli A, Donoghue PCJ. 2016. Tectonic blocks and molecular clocks. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 371(1699):20160098 DOI 10.1098/rstb.2016.0098.
- De Queiroz K. 2005. A unified concept of species and its consequences for the future of taxonomy. *Proceedings of the California Academy of Sciences* 56(18):196–215.
- DeAngelis DL, Grimm V. 2017. Individual-based models in ecology after four decades. *F1000Prime Reports* 6:39 DOI 10.12703/P6-39.

- Dexter KG, Lavin M, Torke BM, Twyford AD, Kursar TA, Coley PD, Drake C, Hollands R, Pennington RT. 2017. Dispersal assembly of rain forest tree communities across the Amazon basin. *Proceedings of the National Academy of Sciences of the United States of America* 114(10):2645–2650 DOI 10.1073/pnas.1613655114.
- Diazgranados M, Barber JC. 2017. Geography shapes the phylogeny of frailejones (Espeletiinae Cuatrec., Asteraceae): a remarkable example of recent rapid radiation in sky islands. *PeerJ* 5:e2968 DOI 10.7717/peerj.2968.
- Diniz-Filho JAF, Siqueira T, Padial AA, Rangel TF, Landeiro VL, Bini LM. 2011. Spatial autocorrelation analysis allows disentangling the balance between neutral and niche processes in metacommunities. *Oikos* 121(2):201–210 DOI 10.1111/j.1600-0706.2011.19563.x.
- Doughty CE, Faurby S, Svenning J-C. 2016. The impact of the megafauna extinctions on savanna woody cover in South America. *Ecography* 39(2):213–222 DOI 10.1111/ecog.01593.
- Dransfield J, Uhl NW, Connie AB, Baker WJ, Harley MM, Lewis CE. 2008. *Genera Palmarum: The Evolution and Classification of Palms*. Kew: Royal Botanic Gardens.
- Duellman WE. 1989. Tropical herpetofaunal communities: patterns of community structure in neotropical rainforests. In: Harmelin-Vivien ML, Bourlière F, eds. *Vertebrates in Complex Tropical Systems. Ecological Studies*. New York: Springer, 61–88.
- Edler D, Guedes T, Zizka A, Rosvall M, Antonelli A. 2016. Infomap bioregions: interactive mapping of biogeographical regions from species distributions. *Systematic Biology* 66(2):197–204 DOI 10.1093/sysbio/syw087.
- Eldredge N. 1989. *Time frames: the evolution of punctuated equilibria*. New Jersey: Princeton University Press.
- Emerson BC, Kolm N. 2005. Species diversity can drive speciation. *Nature* 434(7036):1015–1017 DOI 10.1038/nature03450.
- Espeland M, Hall JPW, DeVries PJ, Lees DC, Cornwall M, Hsu Y-F, Wu L-W, Campbell DL, Talavera G, Vila R, Salzman S, Ruehr S, Lohman DJ, Pierce NE. 2015. Ancient Neotropical origin and recent recolonisation: phylogeny, biogeography and diversification of the Riodinidae (Lepidoptera: Papilionoidea). *Molecular Phylogenetics and Evolution* 93:296–306 DOI 10.1016/j.ympev.2015.08.006.
- Esquivel-Muelbert A, Baker TR, Dexter KG, Lewis SL, Ter Steege H, Lopez-Gonzalez G, Monteagudo Mendoza A, Brien R, Feldpausch TR, Pitman N, Alonso A, Van Der Heijden G, Peña-Claros M, Ahuite M, Alexiades M, Álvarez Dávila E, Murakami AA, Arroyo L, Aulestia M, Balslev H, Barroso J, Boot R, Cano A, Chama Moscoso V, Comiskey JA, Cornejo F, Dallmeier F, Daly DC, Dávila N, Duivenvoorden JF, Duque Montoya AJ, Erwin T, Di Fiore A, Frederickson T, Fuentes A, García-Villacorta R, Gonzales T, Guevara Andino JE, Honorio Coronado EN, Huamantupa-Chuquimaco I, Maria Jiménez RE, Killeen TJ, Malhi Y, Mendoza C, Mogollón H, Jørgensen PM, Montero JC, Mostacedo B, Nauray W, Neill D, Vargas PN, Palacios S, Palacios Cuenca W, Pallqui Camacho NC, Peacock J, Phillips JF, Pickavance G, Quesada CA, Ramirez-Angulo H, Restrepo Z, Reynel Rodriguez C, Paredes MR, Peñuela-Mora MC, Sierra R, Silveira M, Stevenson P, Stropp J, Terborgh J, Tirado M, Toledo M, Torres-Lezama A, Umaña MN, Urrego LE, Vasquez Martinez R, Gamarra LV, Vela CIA, Vilanova Torre E, Vos V, Von Hildebrand P, Vriesendorp C, Wang O, Young KR, Zartman CE, Phillips OL. 2017. Seasonal drought limits tree species across the Neotropics. *Ecography* 40(5):618–629 DOI 10.1111/ecog.01904.
- Faith DP. 1992. Conservation evaluation and phylogenetic diversity. *Biological Conservation* 61(1):1–10 DOI 10.1016/0006-3207(92)91201-3.

- Faurby S, Eiserhardt WL, Svenning J-C. 2016. Strong effects of variation in taxonomic opinion on diversification analyses. *Methods in Ecology and Evolution* 7(1):4–13 DOI 10.1111/2041-210X.12449.
- Faurby S, Svenning J-C. 2015. Historic and prehistoric human-driven extinctions have reshaped global mammal diversity patterns. *Diversity and Distributions* 21(10):1155–1166 DOI 10.1111/ddi.12369.
- Feeley KJ. 2015. Are we filling the data void? An assessment of the amount and extent of plant collection records and census data available for tropical South America. *PLOS ONE* 10(4):e0125629 DOI 10.1371/journal.pone.0125629.
- Fenker J, Tedeschi LG, Pyron RA, Nogueira C de C. 2014. Phylogenetic diversity, habitat loss and conservation in South American pitvipers (Crotalinae: Bothrops and Bothrocophias). *Diversity and Distributions* 20(10):1108–1119 DOI 10.1111/ddi.12217.
- Field R, Hawkins BA, Cornell HV, Currie DJ, Filho JAFD, Guégan JF, Kaufman DM, Kerr JT, Mittelbach GG, Oberdorff T, O'Brien EM, Turner JRG. 2009. Spatial species-richness gradients across scales: a meta-analysis. *Journal of Biogeography* 36(1):132–147 DOI 10.1111/j.1365-2699.2008.01963.x.
- Figueiredo J, Hoorn C, Van Der Ven P, Soares E. 2009. Late Miocene onset of the Amazon river and the Amazon deep-sea fan: evidence from the Foz do Amazonas basin. *Geology* 37(7):619–622 DOI 10.1130/G25567A.1.
- Figueiredo FOG, Zuquim G, Tuomisto H, Moulatlet GM, Balslev H, Costa FRC. 2017. Beyond climate control on species range: the importance of soil data to predict distribution of Amazonian plant species. *Journal of Biogeography* 45(1):190–200 DOI 10.1111/jbi.13104.
- Fine PVA. 2015. Ecological and evolutionary drivers of geographic variation in species diversity. *Annual Review of Ecology, Evolution, and Systematics* 46(1):369–392 DOI 10.1146/annurev-ecolsys-112414-054102.
- Fine PVA, Ree RH. 2006. Evidence for a time-integrated species-area effect on the latitudinal gradient in tree diversity. *American Naturalist* 168(6):796–804 DOI 10.1086/508635.
- Flantua SGA, Hooghiemstra H, Van Boxel JH, Cabrera M, Gonzalez-Carranza Z, Gonzalez-Arrango C. 2015. Connectivity dynamics since the last glacial maximum in the northern Andes: a pollen-driven framework to assess potential migration. In: Stevens WD, Montiel OMR, Raven PH, eds. *Paleobotany and Biogeography; A Festschrift for Alan Graham in His 80 Year*. St. Louis: Missouri Botanical Garden Press, 98–123.
- Fleischner TL, Espinoza RE, Gerrish GA, Greene HW, Kimmerer RW, Lacey EA, Pace S, Parrish JK, Swain HM, Trombulak SC, Weisberg S, Winkler DW, Zander L. 2017. Teaching biology in the field: importance, challenges, and solutions. *BioScience* 67(6):558–567 DOI 10.1093/biosci/bix036.
- Flynn JJ, Wyss AR, Charrier R, Swisher CC. 1995. An early Miocene anthropoid skull from the Chilean Andes. *Nature* 373(6515):603–607 DOI 10.1038/373603a0.
- Forest F, Grenyer R, Rouget M, Davies TJ, Cowling RM, Faith DP, Balmford A, Manning JC, Procheş Ş, van der Bank M, Reeves G, Hedderson TAJ, Savolainen V. 2007. Preserving the evolutionary potential of floras in biodiversity hotspots. *Nature Communications* 445:757–760 DOI 10.1038/nature05587.
- Fouquet A, Loebmann D, Castroviejo-Fisher S, Padial JM, Orrico VGD, Lyra ML, Roberto IJ, Kok PJR, Haddad CLFB, Rodrigues MT. 2012. From Amazonia to the Atlantic forest: molecular phylogeny of Phyzelaphryninae frogs reveals unexpected diversity and a striking biogeographic pattern emphasizing conservation challenges. *Molecular Phylogenetics and Evolution* 65(2):547–561 DOI 10.1016/j.ympev.2012.07.012.

- Fraser LH, Pither J, Jentsch A, Sternberg M, Zobel M, Askarizadeh D, Bartha S, Beierkuhnlein C, Bennett JA, Bittel A, Boldgiv B, Boldrini II, Bork E, Brown L, Cabido M, Cahill J, Carlyle CN, Campetella G, Chelli S, Cohen O, Csergo AM, Diaz S, Enrico L, Ensing D, Fidelis A, Fridley JD, Foster B, Garriss H, Goheen JR, Henry HAL, Hohn M, Jouri MH, Klironomos J, Koorem K, Lawrence-Lodge R, Long R, Manning P, Mitchell R, Moora M, Muller SC, Nabinger C, Naseri K, Overbeck GE, Palmer TM, Parsons S, Pesek M, Pillar VD, Pringle RM, Roccaforte K, Schmidt A, Shang Z, Stahlmann R, Stotz GC, Sugiyama SI, Szentes S, Thompson D, Tungalag R, Undrakhbold S, Van Rooyen M, Wellstein C, Wilson JB, Zupo T. 2015. Worldwide evidence of a unimodal relationship between productivity and plant species richness. *Science* 349(6275):302–305 DOI 10.1126/science.aab3916.
- Fritz SA, Schnitzler J, Eronen JT, Hof C, Böhning-Gaese K, Graham CH. 2013. Diversity in time and space: wanted dead and alive. *Trends in Ecology & Evolution* 28(9):509–516 DOI 10.1016/j.tree.2013.05.004.
- Fukami T. 2015. Historical contingency in community assembly: integrating niches, species pools, and priority effects. *Annual Review of Ecology, Evolution, and Systematics* 46(1):1–23 DOI 10.1146/annurev-ecolsys-110411-160340.
- Funk WC, Caminer M, Ron SR. 2011. High levels of cryptic species diversity uncovered in Amazonian frogs. *Proceedings of the Royal Society B: Biological Sciences* 279(1734):1806–1814 DOI 10.1098/rspb.2011.1653.
- Garnier E, Cortez J, Billès G, Navas M-L, Roumet C, Debussche M, Laurent G, Blanchard A, Aubry D, Bellmann A, Neill C, Toussaint J-P. 2004. Plant functional markers capture ecosystem properties during secondary succession. *Ecology* 85(9):2630–2637 DOI 10.1890/03-0799.
- Garreaud RD, Muñoz RC. 2005. The low-level jet off the West Coast of Subtropical South America: structure and variability. *Monthly Weather Review* 133(8):2246–2261 DOI 10.1175/MWR2972.1.
- Garzón-Orduña IJ, Benetti Longhini JE, Brower AVZ. 2014. Timing the diversification of the Amazonian biota: butterfly divergences are consistent with Pleistocene refugia. *Journal of Biogeography* 41(9):1631–1638 DOI 10.1111/jbi.12330.
- Gaston KJ, Spicer JI. 2004. *Biodiversity: An Introduction*. Oxford: Blackwell Publishing.
- Gehara M, Crawford AJ, Orrico VGD, Rodríguez A, Lötters S, Fouquet A, Barrientos LS, Brusquetti F, De La Riva I, Ernst R, Urrutia GG, Glaw F, Guayasamin JM, Hölting M, Jansen M, Kok PJR, Kwet A, Lingnau R, Lyra M, Moravec J, Pombal JP, Rojas-Runjaic FJM, Schulze A, Señaris JC, Solé M, Rodrigues MT, Twomey E, Haddad CFB, Vences M, Köhler J. 2014. High levels of diversity uncovered in a widespread nominal taxon: continental phylogeography of the neotropical tree frog *Dendropsophus minutus*. *PLOS ONE* 9(10):e103958 DOI 10.1371/journal.pone.0103958.
- Gehara M, Garda AA, Werneck FP, Oliveira EF, Da Fonseca EM, Camurugi F, Magalhães F De M, Lanna FM, Sites JW Jr, Marques R, Silveira-Filho R, São Pedro VA, Colli GR, Costa GC, Burbrink FT. 2017. Estimating synchronous demographic changes across populations using hABC and its application for a herpetological community from northeastern Brazil. *Molecular Ecology* 26(18):4756–4771 DOI 10.1111/mec.14239.
- Gentry AH. 1982. Neotropical floristic diversity: phytogeographical connections between central and South America, Pleistocene climatic fluctuations, or an accident of the andean orogeny? *Annals of the Missouri Botanical Garden* 69(3):557–593 DOI 10.2307/2399084.
- Gentry AH. 1988. Changes in plant community diversity and floristic composition on environmental and geographical gradients. *Annals of the Missouri Botanical Garden* 75(1):1–34 DOI 10.2307/2399464.

- Givnish TJ, Barfuss MHJ, Van Ee B, Riina R, Schulte K, Horres R, Gonsiska PA, Jabaily RS, Crayn DM, Smith JAC, Winter K, Brown GK, Evans TM, Holst BK, Luther H, Till W, Zizka G, Berry PE, Sytsma KJ. 2014. Adaptive radiation, correlated and contingent evolution, and net species diversification in Bromeliaceae. *Molecular Phylogenetics and Evolution* 71:55–78 DOI 10.1016/j.ympev.2013.10.010.
- Gillman LN, Wright SD, Cusens J, McBride PD, Malhi Y, Whittaker RJ. 2014. Latitude, productivity and species richness. *Global Ecology and Biogeography* 24(1):107–117 DOI 10.1111/geb.12245.
- Glassman SF. 1999. *A taxonomic treatment of the palm subtribe Attaleinae (Tribe Cocoeae)*. Urbana: University of Illinois Press.
- Gómez-Acevedo S, Rico-Arce L, Delgado-Salinas A, Magallón S, Eguiarte LE. 2010. Neotropical mutualism between Acacia and Pseudomyrmex: phylogeny and divergence times. *Molecular Phylogenetics and Evolution* 56(1):393–408 DOI 10.1016/j.ympev.2010.03.018.
- Gotelli NJ, Colwell RK. 2001. Quantifying biodiversity: procedures and pitfalls in the measurement and comparison of species richness. *Ecology Letters* 4(4):379–391 DOI 10.1046/j.1461-0248.2001.00230.x.
- Gravel D, Massol F, Canard E, Mouillot D, Mouquet N. 2011. Trophic theory of island biogeography. *Ecology Letters* 14(10):1010–1016 DOI 10.1111/j.1461-0248.2011.01667.x.
- Gray M. 2004. *Geodiversity*. Chichester: John Wiley & Sons.
- Guedes TB, Sawaya RJ, Nogueira C De C. 2014. Biogeography, vicariance and conservation of snakes of the neglected and endangered Caatinga region, north-eastern Brazil. *Journal of Biogeography* 41(5):919–931 DOI 10.1111/jbi.12272.
- Gómez IC, Sääksjärvi IE, Mayhew PJ, Pollet M, Del Castillo CR, Nieves-Aldrey J-L, Broad GR, Roininen H, Tuomisto H. 2018. Variation in the species richness of parasitoid wasps (Ichneumonidae: Pimplinae and Rhyssinae) across sites on different continents. *Insect Conservation and Diversity* 11:305–316.
- Haffer J. 1969. Speciation in Amazonian forest birds. *Science* 165(3889):131–137 DOI 10.1126/science.165.3889.131.
- Hanson CA, Fuhrman JA, Horner-Devine MC, Martiny JBH. 2012. Beyond biogeographic patterns: processes shaping the microbial landscape. *Nature Reviews Microbiology* 10(7):497–506 DOI 10.1038/nrmicro2795.
- Hausdorf B, Hennig C. 2003. Biotic element analysis in biogeography. *Systematic Biology* 52(5):717–723 DOI 10.1080/10635150390235584.
- Hawiltschek O, Ramírez Garrido S, Glaw F. 2017. How marine currents influenced the widespread natural overseas dispersal of reptiles in the Western Indian Ocean region. *Journal of Biogeography* 44(6):1435–1440 DOI 10.1111/jbi.12940.
- Hazen WE. 1964. Readings in population and community ecology. *The American Midland Naturalist* 72(1):250–253 DOI 10.2307/2423228.
- Heath TA, Huelsenbeck JP, Stadler T. 2014. The fossilized birth-death process for coherent calibration of divergence-time estimates. *Proceedings of the National Academy of Sciences of the United States of America* 111(29):E2957–E2966 DOI 10.1073/pnas.1319091111.
- Heithaus ER. 1979. Community structure of neotropical flower visiting bees and wasps: diversity and phenology. *Ecology* 60(1):190–202 DOI 10.2307/1936480.
- Henderson A, Galeano A, Bernal R. 1997. *Field guide to the palms of the Americas*. Princeton: Princeton Press University.
- Henderson A, Galeano G, Bernal R. 1995. *Field guide to the palms of the Americas*. Princeton: Princeton University Press, 352.

- Hickerson MJ, Stahl EA, Lessios HA. 2006. Test for simultaneous divergence using approximate Bayesian computation. *Evolution* 60(12):2435–2453 DOI 10.1554/05-578.1.
- Hill M. 1973. Diversity and evenness: a unifying notation and its consequences. *Ecology* 54(2):427–432 DOI 10.2307/1934352.
- Higgins MA, Ruokolainen K, Tuomisto H, Llerena N, Cardenas G, Phillips OL, Vásquez R, Räsänen M. 2011. Geological control of floristic composition in Amazonian forests. *Journal of Biogeography* 38(11):2136–2149 DOI 10.1111/j.1365-2699.2011.02585.x.
- Holt BG, Lessard J-P, Borregaard MK, Fritz SA, Araújo MB, Dimitrov D, Fabre P-H, Graham CH, Graves GR, Jönsson KA, Nogués-Bravo D, Wang Z, Whittaker RJ, Fjeldså J, Rahbek C. 2013. An update of Wallace's zoogeographic regions of the world. *Science* 339(6115):74–78 DOI 10.1126/science.1228282.
- Hoorn C, Bogota-A GR, Romero-Baez M, Lammertsma EI, Flantua SGA, Dantas EL, Dino R, Do Carmo DA, Chemale F Jr. 2017. The Amazon at sea: onset and stages of the Amazon River from a marine record, with special reference to Neogene plant turnover in the drainage basin. *Global and Planetary Change* 153:51–65 DOI 10.1016/j.gloplacha.2017.02.005.
- Hoorn C, Guerrero J, Sarmiento GA, Lorente MA. 1995. Andean tectonics as a cause for changing drainage patterns in Miocene northern South America. *Geology* 23(3):237 DOI 10.1130/0091-7613(1995)023<0237:ATAACF>2.3.CO;2.
- Hoorn C, Perrigo A, Antonelli A. 2018. *Mountains, climate and biodiversity*. New York: Wiley-Blackwell.
- Hoorn C, Wesselingh FP, Hovikoski J, Guerrero J. 2010a. The development of the Amazonian mega-wetland (Miocene; Brazil, Colombia, Peru, Bolivia). In: Hoorn C, Wesselingh FP, eds. *Amazonia: Landscape and Species Evolution*. Oxford: Wiley-Blackwell Publishing Ltd., 123–142.
- Hoorn C, Wesselingh FP, Ter Steege H, Bermudez MA, Mora A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo JP, Jaramillo CA, Riff D, Negri FR, Hooghiemstra H, Lundberg JG, Stadler T, Sarkinen T, Antonelli A. 2010b. Amazonia through time: Andean uplift, climate change, landscape evolution, and biodiversity. *Science* 330(6006):927–931 DOI 10.1126/science.1194585.
- Hoorn C, Wesselingh FP, Ter Steege H, Bermudez MA, Mora A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo JP, Jaramillo CA, Riff D, Negri FR, Hooghiemstra H, Lundberg JG, Stadler T, Sarkinen T, Antonelli A. 2011. Origins of biodiversity—response. *Science* 331:399–400 DOI 10.1126/science.331.6016.399.
- Hopkins MJG. 2005. Flora da Reserva Ducke, Amazonas, Brasil. *Rodriguésia* 56(86):9–25 DOI 10.1590/2175-78602005568602.
- Horton BK. 2018. Sedimentary record of Andean mountain building. *Earth-Science Reviews* 178:279–309 DOI 10.1016/j.earscirev.2017.11.025.
- Hovenkamp P. 1997. Vicariance events, not areas, should be used in biogeographical analysis. *Cladistics* 13(1–2):67–79 DOI 10.1111/j.1096-0031.1997.tb00241.x.
- Hrbek T, Da Silva VMF, Dutra N, Gravena W, Martin AR, Farias IP. 2014. A new species of river Dolphin from Brazil or: how little do we know our biodiversity. *PLOS ONE* 9(1):e83623 DOI 10.1371/journal.pone.0083623.
- Hrbek T, Seckinger J, Meyer A. 2007. A phylogenetic and biogeographic perspective on the evolution of poeciliid fishes. *Molecular Phylogenetics and Evolution* 43(3):986–998 DOI 10.1016/j.ympev.2006.06.009.
- Hubbell SP. 2001. *The unified neutral theory of biodiversity and biogeography* (MPB-32). Princeton: Princeton University Press.

- Hughes CE, Eastwood R. 2006. Island radiation on a continental scale: exceptional rates of plant diversification after uplift of the Andes. *Proceedings of the National Academy of Sciences of the United States of America* 103(27):10334–10339 DOI 10.1073/pnas.0601928103.
- Hughes CE, Pennington RT, Antonelli A. 2013. Neotropical plant evolution: assembling the big picture. *Botanical Journal of the Linnean Society* 171(1):1–18 DOI 10.1111/boj.12006.
- Humboldt AV, Bonpland A. 1805. *Essai sur la géographie des plantes: accompagné d'un tableau physique des régions équinoxiales, fondé sur des mesures exécutées, depuis le dixième degré de latitude boréale jusqu'au dixième degré de latitude australe, pendant les années 1799, 1800, 1801, 1802 et 1803*. Paris: Chez Levrault, Schoell et compagnie, libraires.
- Humphreys AM, Barraclough TG. 2014. The evolutionary reality of higher taxa in mammals. *Proceedings of the Royal Society B: Biological Sciences* 281(1783):20132750 DOI 10.1098/rspb.2013.2750.
- Humphries CJ, Parenti LR. 1999. *Cladistic biogeography*. New York: Oxford University Press.
- Insel N, Poulsen CJ, Ehlers TA. 2010. Influence of the Andes Mountains on South American moisture transport, convection, and precipitation. *Climate Dynamics* 35(7–8):1477–1492 DOI 10.1007/s00382-009-0637-1.
- Jablonski D, Roy K, Valentine JW. 2006. Out of the tropics: evolutionary dynamics of the latitudinal diversity gradient. *Science* 314(5796):102–106 DOI 10.1126/science.1130880.
- Jaramillo CA. 2006. Cenozoic plant diversity in the neotropics. *Science* 311(5769):1893–1896 DOI 10.1126/science.1121380.
- Jaramillo CA, Ochoa D, Contreras L, Pagani M, Carvajal-Ortiz H, Pratt LM, Krishnan S, Cardona A, Romero M, Quiroz L, Rodriguez G, Rueda MJ, La Parra De F, Morón S, Green W, Bayona G, Montes C, Quintero O, Ramirez R, Mora G, Schouten S, Bermudez H, Navarrete R, Parra F, Alvarán M, Osorno J, Crowley JL, Valencia V, Vervoort J. 2010. Effects of rapid global warming at the paleocene-eocene boundary on neotropical vegetation. *Science* 330(6006):957–961 DOI 10.1126/science.1193833.
- Jaramillo CA, Romero I, D'Apolito C, Bayona G, Duarte E, Louwe S, Escobar J, Luque J, Carrillo-Briceño JD, Zapata V, Mora A, Schouten S, Zavada M, Harrington G, Ortiz J, Wesselingh FP. 2017. Miocene flooding events of western Amazonia. *Science Advances* 3(5):e1601693 DOI 10.1126/sciadv.1601693.
- Jenkins CN, Alves MAS, Uezu A, Vale MM. 2015. Patterns of vertebrate diversity and protection in Brazil. *PLOS ONE* 10(12):e0145064 DOI 10.1371/journal.pone.0145064.
- Kalko EKV, Handley CO. 2001. Neotropical bats in the canopy: diversity, community structure, and implications for conservation. *Plant Ecology* 153(1–2):319–333 DOI 10.1023/A:1017590007861.
- Jin L, Chakraborty R. 1994. Estimation of genetic distance and coefficient of gene diversity from single-probe multilocus DNA fingerprinting data. *Molecular Biology and Evolution* 11(1):120–127 DOI 10.1093/oxfordjournals.molbev.a040086.
- Jost L. 2006. Entropy and diversity. *Oikos* 113:363–375 DOI 10.1111/j.2006.0030-1299.14714.x.
- Kier G, Mutke J, Dinerstein E, Ricketts TH, Küper W, Kreft H, Barthlott W. 2005. Global patterns of plant diversity and floristic knowledge. *Journal of Biogeography* 32(7):1107–1116 DOI 10.1111/j.1365-2699.2005.01272.x.
- Kluge J, Bach K, Kessler M. 2008. Elevational distribution and zonation of tropical pteridophyte assemblages in Costa Rica. *Basic and Applied Ecology* 9(1):35–43 DOI 10.1016/j.baae.2006.11.004.

- Kluge J, Kessler M, Dunn RR. 2006. What drives elevational patterns of diversity? A test of geometric constraints, climate and species pool effects for pteridophytes on an elevational gradient in Costa Rica. *Global Ecology and Biogeography* 15(4):358–371 DOI 10.1111/j.1466-822X.2006.00223.x.
- Kraft NJB, Ackerly DD. 2014. Assembly of plant communities. In: Monson RK, ed. *Ecology and the Environment*. New York: Springer, 67–88.
- Kristiansen T, Svenning JC, Eiserhardt WL, Pedersen D, Brix H, Munch Kristiansen S, Knadel M, Grández C, Balslev H. 2012. Environment versus dispersal in the assembly of western Amazonian palm communities. *Journal of Biogeography* 39(7):1318–1332 DOI 10.1111/j.1365-2699.2012.02689.x.
- Kursar TA, Dexter KG, Lokvam J, Pennington RT, Richardson JE, Weber MG, Murakami ET, Drake C, McGregor R, Coley PD. 2009. The evolution of antiherbivore defenses and their contribution to species coexistence in the tropical tree genus *Inga*. *Proceedings of the National Academy of Sciences of the United States of America* 106(43):18073–18078 DOI 10.1073/pnas.0904786106.
- Lagomarsino LP, Condamine FL, Antonelli A, Mulch A, Davis CA. 2016. The abiotic and biotic drivers of rapid diversification in Andean bellflowers (Campanulaceae). *New Phytologist* 210(4):1430–1442 DOI 10.1111/nph.13920.
- Landis MJ. 2017. Biogeographic dating of speciation times using paleogeographically informed processes. *Systematic Biology* 66(2):128–144 DOI 10.1093/sysbio/syw040.
- Landis MJ, Matzke NJ, Moore BR, Huelsenbeck JP. 2013. Bayesian analysis of biogeography when the number of areas is large. *Systematic Biology* 62(6):789–804 DOI 10.1093/sysbio/syt040.
- Ledo RMD, Colli GR. 2017. The historical connections between the Amazon and the Atlantic Forest revisited. *Journal of Biogeography* 44(11):2551–2563 DOI 10.1111/jbi.13049.
- Leibold MA, Holyoak M, Mouquet N, Amarasekare P, Chase JM, Hoopes MF, Holt RD, Shurin JB, Law R, Tilman D, Loreau M, Gonzalez A. 2004. The metacommunity concept: a framework for multi-scale community ecology. *Ecology Letters* 7(7):601–613 DOI 10.1111/j.1461-0248.2004.00608.x.
- Leite RN, Rogers DS. 2013. Revisiting Amazonian phylogeography: insights into diversification hypotheses and novel perspectives. *Organisms Diversity & Evolution* 13(4):639–664 DOI 10.1007/s13127-013-0140-8.
- Lehtonen S, Jones MM, Zuquim G, Prado J, Tuomisto H. 2015. Phylogenetic relatedness within Neotropical fern communities increases with soil fertility. *Global Ecology and Biogeography* 24:695–705.
- Lemey P, Rambaut A, Drummond AJ, Suchard MA. 2009. Bayesian phylogeography finds its roots. *PLOS Computational Biology* 5(9):e1000520 DOI 10.1371/journal.pcbi.1000520.
- Lemey P, Rambaut A, Welch JJ, Suchard MA. 2010. Phylogeography takes a relaxed random walk in continuous space and time. *Molecular Biology and Evolution* 27(8):1877–1885 DOI 10.1093/molbev/msq067.
- Lemmon A, Lemmon EM. 2008. A likelihood framework for estimating phylogeographic history on a continuous landscape. *Systematic Biology* 57(4):544–561 DOI 10.1080/10635150802304761.
- Leprieur F, Tedesco PA, Hugueny B, Beauchard O, Dürr HH, Brosse S, Oberdorff T. 2011. Partitioning global patterns of freshwater fish beta diversity reveals contrasting signatures of past climate changes. *Ecology Letters* 14(4):325–334 DOI 10.1111/j.1461-0248.2011.01589.x.
- Levis C, Costa FRC, Bongers F, Peña-Claros M, Clement CR, Junqueira AB, Neves EG, Tamanaha EK, Figueiredo FOG, Salomão RP, Castilho CV, Magnusson WE, Phillips OL,

- Guevara JE, Sabatier D, Molino JF, López DC, Mendoza AM, Pitman NCA, Duque A, Vargas PN, Zartman CE, Vasquez R, Andrade A, Camargo JL, Feldpausch TR, Laurance SGW, Laurance WF, Killeen TJ, Nascimento HEM, Montero JC, Mostacedo B, Amaral IL, Vieira ICG, Brien R, Castellanos H, Terborgh J, De Jesus Veiga Carim M, Da Silva Guimarães JR, De Souza Coelho L, De Almeida Matos FD, Wittmann F, Mogollón HF, Damasco G, Dávila N, García-Villacorta R, Coronado ENH, Emilio T, De Andrade Lima Filho D, Schietti J, Souza P, Targhetta N, Comiskey JA, Marimon BS, Marimon BH, Neill D, Alonso A, Arroyo L, Carvalho FA, De Souza FC, Dallmeier F, Pansonato MP, Duivenvoorden JF, Fine PVA, Stevenson PR, Araujo-Murakami A, C GAA, Baraloto C, Do Amaral DD, Engel J, Henkel TW, Maas P, Petronelli P, Revilla JDC, Stropp J, Daly D, Gribel R, Paredes MR, Silveira M, Thomas-Caesar R, Baker TR, Da Silva NF, Ferreira LV, Peres CA, Silman MR, Cerón C, Valverde FC, Di Fiore A, Jimenez EM, Mora MCP, Toledo M, Barbosa EM, De Matos Bonates LC, Arboleda NC, De Sousa Farias E, Fuentes A, Guillaumet JL, Jørgensen PM, Malhi Y, De Andrade Miranda IP, Phillips JF, Prieto A, Rudas A, Ruschel AR, Silva N, Hildebrand Von P, Vos VA, Zent EL, Zent S, Cintra BBL, Nascimento MT, Oliveira AA, Ramirez-Angulo H, Ramos JF, Rivas G, Schöngart J, Sierra R, Tirado M, Van der Heijden G, Torre EV, Wang O, Young KR, Baider C, Cano A, Farfan-Rios W, Ferreira C, Hoffman B, Mendoza C, Mesones I, Torres-Lezama A, Medina MNU, Van Andel TR, Villarroel D, Zagt R, Alexiades MN, Balslev H, Garcia-Cabrera K, Gonzales T, Hernandez L, Huamantupa-Chuquimaco I, Manzatto AG, Milliken W, Cuenca WP, Pansini S, Pauletto D, Arevalo FR, Reis NFC, Sampaio AF, Giraldo LEU, Sandoval EHV, Gamarra LV, Vela CIA, Ter Steege H. 2017. Persistent effects of pre-Columbian plant domestication on Amazonian forest composition. *Science* 355(6328):925–931 DOI 10.1126/science.aal0157.
- Levis C, Flores BM, Moreira PA, Luize BG, Alves RP, Franco-Moraes J, Lins J, Konings E, Peña Claros M, Bongers F, Costa FRC, Clement CR. 2018. How people domesticated Amazonian forests. *Frontiers in Ecology and Evolution* 5:15 DOI 10.3389/fevo.2017.00171.
- Lohmann LG, Taylor CM. 2014. A new generic classification of tribe Bignoniaceae (Bignoniaceae)1. *Annals of the Missouri Botanical Garden* 99(3):348–489 DOI 10.3417/2003187.
- Lomolino MV, Riddle B, Whitakker RJ. 2017. *Biogeography*. Fourth Edition. Sunderland: Sinauer Associates.
- Losos JB, Ricklefs RE. 2009. *The theory of island biogeography revisited*. Princeton: Princeton University Press.
- Lovejoy NR, Lester K, Crampton WGR, Marques FPL, Albert JS. 2010. Phylogeny, biogeography, and electric signal evolution of Neotropical knifefishes of the genus *Gymnotus* (Osteichthyes: Gymnotidae). *Molecular Phylogenetics and Evolution* 54(1):278–290 DOI 10.1016/j.ympev.2009.09.017.
- Lundberg JG, Kottelat M, Smith GR, Stiassny MLJ, Gill AC. 2000. So many fishes, so little time: an overview of recent ichthyological discovery in continental waters. *Annals of the Missouri Botanical Garden* 87(1):26–62 DOI 10.2307/2666207.
- MacArthur RH, Wilson EO. 1967. *The Theory of Island Biogeography*. Princeton: Princeton University Press.
- Magurran AE. 2013. *Measuring Biological Diversity*. Oxford: John Wiley & Sons.
- Mahé F, De Vargas C, Bass D, Czech L, Stamatakis A, Lara E, Singer D, Mayor J, Bunge J, Sernaker S, Siemensmeyer T, Trautmann I, Romac S, Berney C, Kozlov A, Mitchell EAD, Seppey CVW, Egge E, Lentendu G, Wirth R, Trueba G, Dunthorn M. 2017. Parasites dominate hyperdiverse soil protist communities in Neotropical rainforests. *Nature Ecology & Evolution* 1:0091 DOI 10.1038/s41559-017-0091.

- Maldonado C, Molina CI, Zizka A, Persson C, Taylor CM, Albán J, Chilquillo E, Ronsted N, Antonelli A. 2015. Estimating species diversity and distribution in the era of Big Data: to what extent can we trust public databases? *Global Ecology and Biogeography* **24**(8):973–984 DOI [10.1111/geb.12326](https://doi.org/10.1111/geb.12326).
- Manceau M, Lambert A, Morlon H. 2015. Phylogenies support out-of-equilibrium models of biodiversity. *Ecology Letters* **18**(4):347–356 DOI [10.1111/ele.12415](https://doi.org/10.1111/ele.12415).
- Mao K, Milne RI, Zhang L, Peng Y, Liu J, Thomas P, Mill RR, Renner SS. 2012. Distribution of living Cupressaceae reflects the breakup of Pangea. *Proceedings of the National Academy of Sciences of the United States of America* **109**(20):7793–7798 DOI [10.1073/pnas.1114319109](https://doi.org/10.1073/pnas.1114319109).
- Margalef R. 1963. On certain unifying principles in ecology. *American Naturalist* **97**(897):357–374 DOI [10.1086/282286](https://doi.org/10.1086/282286).
- Marshall CJ, Liebherr JK. 2000. Cladistic biogeography of the Mexican transition zone. *Journal of Biogeography* **27**(1):203–216 DOI [10.1046/j.1365-2699.2000.00388.x](https://doi.org/10.1046/j.1365-2699.2000.00388.x).
- Matos-Maraví P, Núñez Aguila R, Peña C, Miller JY, Sourakov A, Wahlberg N. 2014. Causes of endemic radiation in the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini). *BMC Evolutionary Biology* **14**:199 DOI [10.1186/s12862-014-0199-7](https://doi.org/10.1186/s12862-014-0199-7).
- Matos-Maraví P. 2016. Investigating the timing of origin and evolutionary processes shaping regional species diversity: insights from simulated data and neotropical butterfly diversification rates. *Evolution* **70**(7):1638–1650 DOI [10.1111/evo.12960](https://doi.org/10.1111/evo.12960).
- Matzke NJ. 2014. Model selection in historical biogeography reveals that founder-event speciation is a crucial process in Island clades. *Systematic Biology* **63**(6):951–970 DOI [10.1093/sysbio/syu056](https://doi.org/10.1093/sysbio/syu056).
- McGill BJ, Enquist BJ, Weiher E, Westoby M. 2006. Rebuilding community ecology from functional traits. *Trends in Ecology & Evolution* **21**(4):178–185 DOI [10.1016/j.tree.2006.02.002](https://doi.org/10.1016/j.tree.2006.02.002).
- McKenna DD, Farrell BD. 2006. Tropical forests are both evolutionary cradles and museums of leaf beetle diversity. *Proceedings of the National Academy of Sciences of the United States of America* **103**(29):10947–10951 DOI [10.1073/pnas.0602712103](https://doi.org/10.1073/pnas.0602712103).
- McRae BH, Dickson BG, Keitt TH, Shah VB. 2008. Using circuit theory to model connectivity in ecology, evolution, and conservation. *Ecology* **89**(10):2712–2724 DOI [10.1890/07-1861.1](https://doi.org/10.1890/07-1861.1).
- Melo WA, Freitas CG, Bacon CD, Collevatti RG. 2018. The road to evolutionary success: insights from the demographic history of an Amazonian palm. *Heredity* **121**(2):183–195 DOI [10.1038/s41437-018-0074-1](https://doi.org/10.1038/s41437-018-0074-1).
- Mendes LW, Tsai SM, Navarrete AA, De Hollander M, Van Veen JA, Kuramae EE. 2015. Soil-Borne Microbiome: linking diversity to function. *Microbial Ecology* **70**(1):255–265 DOI [10.1007/s00248-014-0559-2](https://doi.org/10.1007/s00248-014-0559-2).
- Meseguer AS, Condamine FL. 2018. Ancient tropical extinctions contributed to the latitudinal diversity gradient. *bioRxiv* 236646 DOI [10.1101/236646](https://doi.org/10.1101/236646).
- Meseguer AS, Lobo JM, Ree R, Beerling DJ, Sanmartín I. 2015. Integrating fossils, phylogenies, and niche models into biogeography to reveal ancient evolutionary history: the case of *Hypericum* (Hypericaceae). *Systematic Biology* **64**(2):215–232 DOI [10.1093/sysbio/syu088](https://doi.org/10.1093/sysbio/syu088).
- Meyer C, Kreft H, Guralnick R, Jetz W. 2015. Global priorities for an effective information basis of biodiversity distributions. *Nature Communications* **6**(1):8221 DOI [10.1038/ncomms9221](https://doi.org/10.1038/ncomms9221).
- Minoarivelo HO, Hui C, Terblanche JS, Kosakovsky Pond SL, Scheffler K. 2014. Detecting phylogenetic signal in mutualistic interaction networks using a Markov process model. *Oikos* **123**(10):1250–1260 DOI [10.1111/oik.00857](https://doi.org/10.1111/oik.00857).

- Mittelbach GG, Schemske DW. 2015. Ecological and evolutionary perspectives on community assembly. *Trends in Ecology and Evolution* 30(5):241–247 DOI 10.1016/j.tree.2015.02.008.
- Moritz C, Patton JL, Schneider CJ, Smith TB. 2000. Diversification of rainforest faunas: an integrated molecular approach. *Annual Review of Ecology and Systematics* 31(1):533–563 DOI 10.1146/annurev.ecolsys.31.1.533.
- Morrone JJ. 2013. Cladistic biogeography of the neotropical region: identifying the main events in the diversification of the terrestrial biota. *Cladistics* 30(2):202–214 DOI 10.1111/cla.12039.
- Moura MR, Villalobos F, Costa GC, Garcia PCA. 2016. Disentangling the role of climate, topography and vegetation in species richness gradients. *PLOS ONE* 11(3):e0152468 DOI 10.1371/journal.pone.0152468.
- Nascimento FF, Lazar A, Menezes AN, Da Matta Durans A, Moreira JC, Salazar-Bravo J, Andrea PSD, Bonvicino CR. 2013. The role of historical barriers in the diversification processes in open vegetation formations during the Miocene/Pliocene using an ancient rodent lineage as a model. *PLOS ONE* 8(4):e61924 DOI 10.1371/journal.pone.0061924.
- Nelson G, Platnick NI. 1980. A vicariance approach to historical biogeography. *BioScience* 30(5):339–343 DOI 10.2307/1307856.
- Noguera-Urbano EA, Escalante T. 2015. Areas de endemismo de los mamíferos (mammalia) neotropicales. *Acta Biológica Colombiana* 20(3):47–65 DOI 10.15446/abc.v20n3.46179.
- Nores M. 2002. An alternative hypothesis for the origin of Amazonian bird diversity. *Journal of Biogeography* 26(3):475–485 DOI 10.1046/j.1365-2699.1999.t01-1-00311.x.
- Novo NM, Tejedor MF, Pérez ME, Krause JM. 2017. New primate locality from the early Miocene of Patagonia, Argentina. *American Journal of Physical Anthropology* 164(4):861–867 DOI 10.1002/ajpa.23309.
- Onstein RE, Baker WJ, Couvreur TLP, Faurby S, Svenning JC, Kissling WD. 2017. Frugivory-related traits promote speciation of tropical palms. *Nature Ecology & Evolution* 1(12):1903–1911 DOI 10.1038/s41559-017-0348-7.
- Palazzesi L, Barreda VD, Cuitiño JI, Guler MV, Tellería MC, Santos RV. 2014. Fossil pollen records indicate that Patagonian desertification was not solely a consequence of Andean uplift. *Nature Communications* 5(1):3558–3579 DOI 10.1038/ncomms4558.
- Papadopoulou A, Knowles LL. 2016. Toward a paradigm shift in comparative phylogeography driven by trait-based hypotheses. *Proceedings of the National Academy of Sciences of the United States of America* 113(29):8018–8024 DOI 10.1073/pnas.1601069113.
- Peres CA, Emilio T, Schietti J, Desmoulière SJM, Levi T. 2016. Dispersal limitation induces long-term biomass collapse in overhunted Amazonian forests. *Proceedings of the National Academy of Sciences of the United States of America* 113(4):892–897 DOI 10.1073/pnas.1516525113.
- Pérez-Escobar OA, Chomicki G, Condamine FL, Karremans AP, Bogarín D, Matzke NJ, Silvestro D, Antonelli A. 2017. Recent origin and rapid speciation of Neotropical orchids in the world’s richest plant biodiversity hotspot. *New Phytologist* 215(2):891–905 DOI 10.1111/nph.14629.
- Perret M, Chautems A, Spichiger R, Barraclough TG, Savolainen V. 2007. The geographical pattern of speciation and floral diversification in the neotropics: the tribe Sinningieae (Gesneriaceae) as a case study. *Evolution* 61(7):1641–1660 DOI 10.1111/j.1558-5646.2007.00136.x.
- Phillips OL, Vargas PN, Monteagudo AL, Cruz AP, Zans MEC, Sánchez WG, Yli Halla M, Rose S. 2003. Habitat association among Amazonian tree species: a landscape-scale approach. *Journal of Ecology* 91(5):757–775 DOI 10.1046/j.1365-2745.2003.00815.x.

- Pilosof S, Porter MA, Pascual M, Kéfi S. 2017. The multilayer nature of ecological networks. *Nature Ecology & Evolution* 1(4):0101 DOI 10.1038/s41559-017-0101.
- Prance GT. 1990. The floristic composition of the forest of Central Amazonian Brazil. In: Gentry AH, ed. *Four Neotropical Rain Forests*. New Haven: Yale University Press, 112–140.
- Prates I, Rivera D, Rodrigues MT, Carnaval AC. 2016a. A mid-Pleistocene rainforest corridor enabled synchronous invasions of the Atlantic Forest by Amazonian anole lizards. *Molecular Ecology* 25(20):5174–5186 DOI 10.1111/mec.13821.
- Prates I, Xue AT, Brown JL, Alvarado-Serrano DF, Rodrigues MT, Hickerson MJ, Carnaval AC. 2016b. Inferring responses to climate dynamics from historical demography in neotropical forest lizards. *Proceedings of the National Academy of Sciences of the United States of America* 113(29):7978–7985 DOI 10.1073/pnas.1601063113.
- Purvis A, Hector A. 2000. Getting the measure of biodiversity. *Nature* 405(6783):212–219 DOI 10.1038/35012221.
- Quintero I, Keil P, Jetz W, Crawford FW. 2015. Historical biogeography using species geographical ranges. *Systematic Biology* 64(6):1059–1073 DOI 10.1093/sysbio/syv057.
- Ramírez SR, Eltz T, Fujiwara MK, Gerlach G, Goldman-Huertas B, Tsutsui ND, Pierce NE. 2011. Asynchronous diversification in a specialized plant-pollinator mutualism. *Science* 333(6050):1742–1746 DOI 10.1126/science.1209175.
- Räsänen ME, Linna AM, Santos JCR, Negri FR. 1995. Late Miocene tidal deposits in the Amazonian foreland basin. *Science* 269:386–390.
- Räsänen ME, Salo JS, Kalliola RJ. 1987. Fluvial perturbation in the western Amazon basin: regulation by long-term Sub-Andean tectonics. *Science* 238(4832):1398–1401 DOI 10.1126/science.238.4832.1398.
- Raven PH, Axelrod DI. 1974. Angiosperm biogeography and past continental movements. *Annals of the Missouri Botanical Garden* 61(3):539 DOI 10.2307/2395021.
- Ree RH, Sanmartín I. 2009. Prospects and challenges for parametric models in historical biogeographical inference. *Journal of Biogeography* 36(7):1211–1220 DOI 10.1111/j.1365-2699.2008.02068.x.
- Ree RH, Sanmartín I. 2018. Conceptual and statistical problems with the DEC+J model of founder-event speciation and its comparison with DEC via model selection. *Journal of Biogeography* 45(4):741–749 DOI 10.1111/jbi.13173.
- Ree RH, Smith SA. 2008. Maximum likelihood inference of geographic range evolution by dispersal, local extinction, and cladogenesis. *Systematic Biology* 57(1):4–14 DOI 10.1080/10635150701883881.
- Reis RE, Albert JS, Di Dario F, Mincarone MM, Petry P, Rocha LA. 2016. Fish biodiversity and conservation in South America. *Journal of Fish Biology* 89(1):12–47 DOI 10.1111/jfb.13016.
- Rezende EL, Jordano P, Bascompte J. 2007. Effects of phenotypic complementarity and phylogeny on the nested structure of mutualistic networks. *Oikos* 116(11):1919–1929 DOI 10.1111/j.2007.0030-1299.16029.x.
- Ribas CC, Aleixo A, Nogueira ACR, Miyaki CY, Cracraft J. 2012. A palaeobiogeographic model for biotic diversification within Amazonia over the past three million years. *Proceedings of the Royal Society B: Biological Sciences* 279(1729):681–689 DOI 10.1098/rspb.2011.1120.
- Ribeiro LF, Bornschein MR, Belmonte-Lopes R, Firkowski CR, Morato SAA, Pie MR. 2015. Seven new microendemic species of *Brachycephalus* (Anura: Brachycephalidae) from southern Brazil. *PeerJ* 3:e1011 DOI 10.7717/peerj.1011.
- Ricklefs RE. 2008. Disintegration of the ecological community. *American Naturalist* 172(6):741–750 DOI 10.1086/593002.

- Ricklefs RE, Jenkins DG. 2011. Biogeography and ecology: towards the integration of two disciplines. *Philosophical Transactions of the Royal Society B: Biological Sciences* 366(1576):2438–2448 DOI 10.1098/rstb.2011.0066.
- Ritchie ME. 2009. *Scale, heterogeneity, and the structure and diversity of ecological communities*. Princeton: Princeton University Press.
- Rohrman A, Sachse D, Mulch A, Pingel H, Tofelde S, Alonso RN, Strecker MR. 2016. Miocene orographic uplift forces rapid hydrological change in the southern central Andes. *Scientific Reports* 6(1):35678 DOI 10.1038/srep35678.
- Rojas D, Vale Á, Ferrero V, Navarro L. 2012. The role of frugivory in the diversification of bats in the Neotropics. *Journal of Biogeography* 39(11):1948–1960 DOI 10.1111/j.1365-2699.2012.02709.x.
- Roncal J. 2006. Habitat differentiation of sympatric *Geonoma macrostachys* (Arecaceae) varieties in Peruvian lowland forests. *Journal of Tropical Ecology* 22(4):483–486 DOI 10.1017/S0266467406003270.
- Ronquist F, Klopstein S, Vilhelmsen L, Schulmeister S, Murray DL, Rasnitsyn AP. 2012. A total-evidence approach to dating with fossils, applied to the early radiation of the hymenoptera. *Systematic Biology* 61(6):973–999 DOI 10.1093/sysbio/sys058.
- Ronquist F, Sanmartín I. 2011. Phylogenetic methods in biogeography. *Annual Review of Ecology, Evolution, and Systematics* 42(1):441–464 DOI 10.1146/annurev-ecolsys-102209-144710.
- Rosauer DF, Jetz W. 2014. Phylogenetic endemism in terrestrial mammals. *Global Ecology and Biogeography* 24(2):168–179 DOI 10.1111/geb.12237.
- Rosen DE. 1975. A vicariance model of caribbean biogeography. *Systematic Zoology* 24(4):431–464 DOI 10.2307/2412905.
- Rosen DE. 1978. Vicariant patterns and historical explanation in biogeography. *Systematic Biology* 27(2):159–188 DOI 10.2307/2412970.
- Rosenberger AL, Tejedor MF, Cooke SB, Pekar S. 2009. Platyrrhine ecophylogenetics in space and time. In: Garber P, Estrada A, Bicca-Marquez J, Strier K, eds. *South American Primates*. New York: Springer, 69–113.
- Rosindell J, Harmon LJ. 2013. A unified model of species immigration, extinction and abundance on islands. *Journal of Biogeography* 40(6):1107–1118 DOI 10.1111/jbi.12064.
- Rossatto DR. 2014. Spatial patterns of species richness and phylogenetic diversity of woody plants in the neotropical savannas of Brazil. *Brazilian Journal of Botany* 37(3):283–292 DOI 10.1007/s40415-014-0070-5.
- Sacek V. 2014. Drainage reversal of the Amazon River due to the coupling of surface and lithospheric processes. *Earth and Planetary Science Letters* 401:301–312 DOI 10.1016/j.epsl.2014.06.022.
- Safi K, Cianciaruso MV, Loyola RD, Brito D, Armour-Marshall K, Diniz-Filho JAF. 2011. Understanding global patterns of mammalian functional and phylogenetic diversity. *Philosophical Transactions of the Royal Society B: Biological Sciences* 366(1577):2536–2544 DOI 10.1098/rstb.2011.0024.
- Said Gutiérrez-Ortega J, Yamamoto T, Vovides AP, Angel Pérez-Farrera M, Martínez JF, Molina-Freaner F, Watano Y, Kajita T. 2017. Aridification as a driver of biodiversity: a case study for the cycad genus *Dioon* (Zamiaceae). *Annals of Botany* 121(1):47–60 DOI 10.1093/aob/mcx123.
- Salo J, Kalliola R, Häkkinen I, Mäkinen Y, Niemelä P, Puhakka M, Coley PD. 1986. River dynamics and the diversity of Amazon lowland forest. *Nature* 322(6076):254–258 DOI 10.1038/322254a0.

- Sandom C, Faurby S, Sandel B, Svenning JC. 2014. Global late quaternary megafauna extinctions linked to humans, not climate change. *Proceedings of the Royal Society B: Biological Sciences* 281(1787):20133254 DOI 10.1098/rspb.2013.3254.
- Sanín MJ, Kissling WD, Bacon CD, Borchsenius F, Galeano G, Svenning JC, Olivera J, Ramírez R, Trénel P, Pintaud JC. 2016. The Neogene rise of the tropical Andes facilitated diversification of wax palms (Ceroxylon: Arecaceae) through geographical colonization and climatic niche separation. *Botanical Journal of the Linnean Society* 182(2):303–317 DOI 10.1111/boj.12419.
- Sanmartín I. 2016. Breaking the chains of parsimony: the development of parametric methods in historical biogeography. In: Barry Cox C, Moore PD, Ladle R, eds. *Biogeography: An Ecological and Evolutionary Approach*. Chichester: Wiley-Blackwell, 239–243.
- Sanmartín I, Meseguer AS. 2016. Extinction in Phylogenetics and biogeography: from timetrees to patterns of biotic assemblage. *Frontiers in Genetics* 7:35 DOI 10.3389/fgene.2016.00035.
- Sanmartín I, Ronquist F. 2004. Southern hemisphere biogeography inferred by event-based models: plant versus animal patterns. *Systematic Biology* 53(2):216–243 DOI 10.1080/10635150490423430.
- Sanmartín I, Van Der Mark P, Ronquist F. 2008. Inferring dispersal: a Bayesian approach to phylogeny-based island biogeography, with special reference to the Canary Islands. *Journal of Biogeography* 35(3):428–449 DOI 10.1111/j.1365-2699.2008.01885.x.
- Santos JC, Coloma LA, Summers K, Caldwell JP, Ree R, Cannatella DC. 2009. Amazonian amphibian diversity is primarily derived from late Miocene Andean Lineages. *PLOS Biology* 7(3):e1000056 DOI 10.1371/journal.pbio.1000056.
- Satler JD, Carstens BC. 2016. Phylogeographic concordance factors quantify phylogeographic congruence among co-distributed species in the *Sarracenia alata* pitcher plant system. *Evolution* 70(5):1105–1119 DOI 10.1111/evo.12924.
- Satler JD, Carstens BC. 2017. Do ecological communities disperse across biogeographic barriers as a unit? *Molecular Ecology* 26(13):3533–3545 DOI 10.1111/mec.14137.
- Satler JD, Zellmer AJ, Carstens BC. 2016. Biogeographic barriers drive co-diversification within associated eukaryotes of the *Sarracenia alata* pitcher plant system. *PeerJ* 4:e1576 DOI 10.7717/peerj.1576.
- Schluter D, Ricklefs RE. 1993. Species diversity: an introduction to the problem. In: Ricklefs RE, Schluter D, eds. *Species Diversity in Ecological Communities: Historical and Geographical Perspectives*. Chicago: University of Chicago Press, 1–10.
- Sexton JP, McIntyre PJ, Angert AL, Rice KJ. 2009. Evolution and ecology of species range limits. *Annual Review of Ecology, Evolution, and Systematics* 40(1):415–436 DOI 10.1146/annurev.ecolsys.110308.120317.
- Shephard GE, Müller RD, Liu L, Gurnis M. 2010. Miocene drainage reversal of the Amazon River driven by plate–mantle interaction. *Nature Geoscience* 3(12):870–875 DOI 10.1038/ngeo1017.
- Silvestro D, Kostikova A, Litsios G, Pearman PB, Salamin N. 2015. Measurement errors should always be incorporated in phylogenetic comparative analysis. *Methods in Ecology and Evolution* 6(3):340–346 DOI 10.1111/2041-210X.12337.
- Silvestro D, Schnitzler J, Liow LH, Antonelli A, Salamin N. 2014. Bayesian estimation of speciation and extinction from incomplete fossil occurrence data. *Systematic Biology* 63(3):349–367 DOI 10.1093/sysbio/syu006.
- Silvestro D, Tejedor MF, Serrano-Serrano ML, Loiseau O, Rossier V, Rolland J, Zizka A, Höhna A, Antonelli A, Salamin N. Early arrival and climatically-linked geographic expansion

- of New World monkeys from tiny African ancestors. *Systematic Biology* (in press)
DOI [10.1093/sysbio/syy046](https://doi.org/10.1093/sysbio/syy046).
- Silvestro D, Zizka A, Bacon CD, Cascales-Miñana B, Salamin N, Antonelli A. 2016.** Fossil biogeography: a new model to infer dispersal, extinction and sampling from palaeontological data. *Philosophical Transactions of the Royal Society B: Biological Sciences* **371(1691)**:20150225
DOI [10.1098/rstb.2015.0225](https://doi.org/10.1098/rstb.2015.0225).
- Simões PI, Stow A, Hödl W, Amézquita A, Farias IP, Lima AP. 2014.** The value of including intraspecific measures of biodiversity in environmental impact surveys is highlighted by the Amazonian Brilliant-Thighed Frog (*Allobates Femoralis*). *Tropical Conservation Science* **7(4)**:811–828
DOI [10.1177/194008291400700416](https://doi.org/10.1177/194008291400700416).
- Simon MF, Grether R, De Queiroz LP, Skema C, Pennington RT, Hughes CE. 2009.** Recent assembly of the Cerrado, a neotropical plant diversity hotspot, by in situ evolution of adaptations to fire. *Proceedings of the National Academy of Sciences of the United States of America* **106(48)**:20359–20364
DOI [10.1073/pnas.0903410106](https://doi.org/10.1073/pnas.0903410106).
- Smith BT, McCormack JE, Cuervo AM, Hickerson MJ, Aleixo A, Cadena CD, Pérez-Emán J, Burney CW, Xie X, Harvey MG, Faircloth BC, Glenn TC, Derryberry EP, Prejean J, Fields S, Brumfield RT. 2014.** The drivers of tropical speciation. *Nature* **515(7527)**:406–409
DOI [10.1038/nature13687](https://doi.org/10.1038/nature13687).
- Smith BT, Seeholzer GF, Harvey MG, Cuervo AM, Brumfield RT. 2017.** A latitudinal phylogeographic diversity gradient in birds. *PLOS Biology* **15(4)**:e2001073
DOI [10.1371/journal.pbio.2001073](https://doi.org/10.1371/journal.pbio.2001073).
- Soria-Auza RW, Kessler M. 2008.** The influence of sampling intensity on the perception of the spatial distribution of tropical diversity and endemism: a case study of ferns from Bolivia. *Diversity and Distributions* **14(1)**:123–130
DOI [10.1111/j.1472-4642.2007.00433.x](https://doi.org/10.1111/j.1472-4642.2007.00433.x).
- Stebbins GL. 1974.** *Flowering plants: evolution above the species level*. Cambridge: The Belknap Press of Harvard University Press.
- Strecker AL, Olden JD, Whittier JB, Paukert CP. 2011.** Defining conservation priorities for freshwater fishes according to taxonomic, functional, and phylogenetic diversity. *Ecological Applications* **21(8)**:3002–3013
DOI [10.1890/11-0599.1](https://doi.org/10.1890/11-0599.1).
- Stropp J, Ter Steege H, Malhi Y, ATDN, RAINFOR. 2009.** Disentangling regional and local tree diversity in the Amazon. *Ecography* **32(1)**:46–54
DOI [10.1111/j.1600-0587.2009.05811.x](https://doi.org/10.1111/j.1600-0587.2009.05811.x).
- Swenson NG. 2011.** The role of evolutionary processes in producing biodiversity patterns, and the interrelationships between taxonomic, functional and phylogenetic biodiversity. *American Journal of Botany* **98(3)**:472–480
DOI [10.3732/ajb.1000289](https://doi.org/10.3732/ajb.1000289).
- Szumik CA, Cuezco F, Goloboff PA, Chalup AE. 2002.** An optimality criterion to determine areas of endemism. *Systematic Biology* **51(5)**:806–816
DOI [10.1080/10635150290102483](https://doi.org/10.1080/10635150290102483).
- Szumik CA, Goloboff PA. 2004.** Areas of endemism: an improved optimality criterion. *Systematic Biology* **53(6)**:968–977
DOI [10.1080/10635150490888859](https://doi.org/10.1080/10635150490888859).
- Tagliacollo VA, Duke-Sylvester SM, Matamoros WA, Chakrabarty P, Albert JS. 2015a.** Coordinated dispersal and pre-isthmian assembly of the Central American Ichthyofauna. *Systematic Biology* **66(2)**:183–196
DOI [10.1093/sysbio/syv064](https://doi.org/10.1093/sysbio/syv064).
- Tagliacollo VA, Roxo FF, Duke-Sylvester SM, Oliveira C, Albert JS. 2015b.** Biogeographical signature of river capture events in Amazonian lowlands. *Journal of Biogeography* **42(12)**:2349–2362
DOI [10.1111/jbi.12594](https://doi.org/10.1111/jbi.12594).
- Tedesco PA, Beauchard O, Bigorne R, Blanchet S, Buisson L, Conti L, Cornu J-F, Dias MS, Grenouillet G, Hugueny B, Jézéquel C, Leprieur F, Brosse S, Oberdorff T. 2017.** A global

- database on freshwater fish species occurrence in drainage basins. *Scientific Data* 4:170141 DOI 10.1038/sdata.2017.141.
- Tejedor MF, Muñoz-Saba Y. 2013. La sistemática de los platirrininos y el registro fósil. In: Deftter TR, Stenvenson PR, Bueno ML, Guzmán Caro DC, eds. *Primates colombianos en peligro de extinción*. Bogotá: researchgate.net, 68–86.
- Tejedor MF, Tauber AA, Rosenberger AL, Swisher CC, Palacios ME. 2006. New primate genus from the Miocene of Argentina. *Proceedings of the National Academy of Sciences of the United States of America* 103(14):5437–5441 DOI 10.1073/pnas.0506126103.
- Ter Steege H, Haripersaud PP, Bánki OS, Schieving F. 2011. A model of botanical collectors' behavior in the field: never the same species twice. *American Journal of Botany* 98(1):31–37 DOI 10.3732/ajb.1000215.
- Ter Steege H, Pitman NCA, Phillips OL, Chave J, Sabatier D, Duque A, Molino J-F, Prévost M-F, Spichiger R, Castellanos H, Von Hildebrand P, Vasquez R. 2006. Continental-scale patterns of canopy tree composition and function across Amazonia. *Nature* 443(7110):444–447 DOI 10.1038/nature05134.
- Ter Steege H, Pitman NCA, Sabatier D, Baraloto C, Salomão RP, Guevara JE, Phillips OL, Castilho CV, Magnusson WE, Molino J-F, Monteagudo A, Núñez Vargas P, Montero JC, Feldpausch TR, Coronado ENH, Killeen TJ, Mostacedo B, Vasquez R, Assis RL, Terborgh J, Wittmann F, Andrade A, Laurance WF, Laurance SGW, Marimon BS, Marimon B-H, Guimarães Vieira IC, Amaral IL, Brien R, Castellanos H, Cárdenas López D, Duivenvoorden JF, Mogollón HF, Matos FDDA, Dávila N, García-Villacorta R, Stevenson Diaz PR, Costa F, Emilio T, Levis C, Schiatti J, Souza P, Alonso A, Dallmeier F, Montoya AJD, Fernandez Piedade MT, Araujo-Murakami A, Arroyo L, Gribel R, Fine PVA, Peres CA, Toledo M, Aymard C GA, Baker TR, Cerón C, Engel J, Henkel TW, Maas P, Petronelli P, Stropp J, Zartman CE, Daly D, Neill D, Silveira M, Paredes MR, Chave J, Lima Filho DDA, Jørgensen PM, Fuentes A, Schöngart J, Cornejo Valverde F, Di Fiore A, Jimenez EM, Peñuela Mora MC, Phillips JF, Rivas G, Van Andel TR, Von Hildebrand P, Hoffman B, Zent EL, Malhi Y, Prieto A, Rudas A, Ruschell AR, Silva N, Vos V, Zent S, Oliveira AA, Schutz AC, Gonzales T, Trindade Nascimento M, Ramirez-Angulo H, Sierra R, Tirado M, Umaña Medina MN, Van Der Heijden G, Vela CIA, Vilanova Torre E, Vriesendorp C, Wang O, Young KR, Baider C, Balslev H, Ferreira C, Mesones I, Torres-Lezama A, Urrego Giraldo LE, Zagt R, Alexiades MN, Hernandez L, Huamantupa-Chuquimaco I, Milliken W, Palacios Cuenca W, Pauletto D, Valderrama Sandoval E, Valenzuela Gamarra L, Dexter KG, Feeley K, Lopez-Gonzalez G, Silman MR. 2013. Hyperdominance in the Amazonian tree flora. *Science* 342(6156):1243092 DOI 10.1126/science.1243092.
- Ter Steege H, Vaessen RW, Cárdenas López D, Sabatier D, Antonelli A, De Oliveira SM, Pitman NCA, Jørgensen PM, Salomão RP. 2016. The discovery of the Amazonian tree flora with an updated checklist of all known tree taxa. *Scientific Reports* 6(1):29549 DOI 10.1038/srep29549.
- Thomaz AT, Malabarba LR, Knowles LL. 2017. Genomic signatures of paleodrainages in a freshwater fish along the southeastern coast of Brazil: genetic structure reflects past riverine properties. *Heredity* 119(4):287–294 DOI 10.1038/hdy.2017.46.
- Thomé MTC, Sequeira F, Brusquetti F, Carstens B, Haddad CFB, Rodrigues MT, Alexandrino J. 2016. Recurrent connections between Amazon and Atlantic forests shaped diversity in Caatinga four-eyed frogs. *Journal of Biogeography* 43(5):1045–1056 DOI 10.1111/jbi.12685.
- Tilman D, Downing JA. 1994. Biodiversity and stability in grasslands. *Nature Communications* 367:363–365 DOI 10.1038/367363a0.

- Toju H, Yamamichi M, Guimarães PR, Olesen JM, Mougi A, Yoshida T, Thompson JN. 2017. Species-rich networks and eco-evolutionary synthesis at the metacommunity level. *Nature Ecology & Evolution* 1(2):1–11 DOI 10.1038/s41559-016-0024.
- Töpel M, Zizka A, Calió MF, Scharn R, Silvestro D, Antonelli A. 2016. SpeciesGeoCoder: fast categorization of species occurrences for analyses of biodiversity, biogeography, ecology, and evolution. *Systematic Biology* 66(2):145–151 DOI 10.1093/sysbio/syw064.
- Toussaint A, Charpin N, Brosse S, Villéger S. 2016. Global functional diversity of freshwater fish is concentrated in the Neotropics while functional vulnerability is widespread. *Scientific Reports* 6(1):51 DOI 10.1038/srep22125.
- Tucker CM, Cadotte MW, Carvalho SB, Davies TJ, Ferrier S, Fritz SA, Grenyer R, Helmus MR, Jin LS, Mooers AO, Pavoine S, Purschke O, Redding DW, Rosauer DF, Winter M, Mazel F. 2016. A guide to phylogenetic metrics for conservation, community ecology and macroecology. *Biological Reviews* 92(2):698–715 DOI 10.1111/brv.12252.
- Tuomisto H. 2010. A diversity of beta diversities: straightening up a concept gone awry. Part 1. Defining beta diversity as a function of alpha and gamma diversity. *Ecography* 33(1):2–22 DOI 10.1111/j.1600-0587.2009.05880.x.
- Tuomisto H. 2018. Different ways of defining diversity, and how to apply them in montane systems. In: Hoorn C, Perrigo A, Antonelli A, eds. *Mountains, Climate, and Biodiversity*. First Edition. Hoboken: John Wiley & Sons Ltd., 295–308.
- Tuomisto H, Moulatlet GM, Balslev H, Emilio T, Figueiredo FOG, Pedersen D, Ruokolainen K. 2016. A compositional turnover zone of biogeographical magnitude within lowland Amazonia. *Journal of Biogeography* 43(12):2400–2411 DOI 10.1111/jbi.12864.
- Tuomisto H, Poulsen AD. 1996. Influence of edaphic specialization on pteridophyte distribution in neotropical rain forests. *Journal of Biogeography* 23(3):283–293 DOI 10.1046/j.1365-2699.1996.00044.x.
- Tuomisto H, Ruokolainen K, Yli-Halla M. 2003. Dispersal, Environment, and Floristic Variation of Western Amazonian Forests. *Science* 299:241–244.
- Tuomisto H, Poulsen AD, Ruokolainen K, Moran RC, Quintana C, Celi J, Cañas G. 2003. Linking floristic patterns with soil heterogeneity and satellite imagery in ecuadorian amazonia. *Ecological Applications* 13(2):352–371 DOI 10.1890/1051-0761(2003)013[0352:LFPWSH]2.0.CO;2.
- Tuomisto H, Ruokolainen K, Kalliola R, Linna A, Danjoy W, Rodriguez Z. 1995. Dissecting Amazonian Biodiversity. *Science* 269(5220):63–66 DOI 10.1126/science.269.5220.63.
- Tuomisto H, Zuquim G, Cárdenas G. 2014. Species richness and diversity along edaphic and climatic gradients in Amazonia. *Ecography* 37(11):1034–1046 DOI 10.1111/ecog.00770.
- Valdujo PH, Carnaval ACOQ, Graham CH. 2013. Environmental correlates of anuran beta diversity in the Brazilian Cerrado. *Ecography* 36(6):708–717 DOI 10.1111/j.1600-0587.2012.07374.x.
- Valente LM, Etienne RS, Phillimore AB. 2014. The effects of island ontogeny on species diversity and phylogeny. *Proceedings of the Royal Society B: Biological Sciences* 281(1784):20133227 DOI 10.1098/rspb.2013.3227.
- Valente LM, Phillimore AB, Etienne RS. 2015. Equilibrium and non-equilibrium dynamics simultaneously operate in the Galápagos islands. *Ecology Letters* 18(8):844–852 DOI 10.1111/ele.12461.
- Valente L, Phillimore AB, Etienne RS. 2018. Using molecular phylogenies in island biogeography: it's about time. *Ecography* 182:820 DOI 10.1111/ecog.03503.
- Van Der Sleen P, Albert JS. 2017. *Field Guide to the Fishes of the Amazon, Orinoco, and Guianas*. Princeton: Princeton University Press.

- Vellend M. 2010. Conceptual Synthesis in Community Ecology. *The Quarterly Review of Biology* 85(2):183–206 DOI 10.1086/652373.
- Vilhena DA, Antonelli A. 2015. A network approach for identifying and delimiting biogeographical regions. *Nature Communications* 6(1):1–9 DOI 10.1038/ncomms7848.
- Vine FJ, Matthews DH. 1963. Magnetic anomalies over oceanic ridges. *Nature* 199(4897):947–949 DOI 10.1038/199947a0.
- Wallace AR. 1889. *A narrative of travels on the amazon and rio negro: with an account of the native tribes, and observations on the climate, geology, and natural history of the amazon valley*. London: Ward, Lock.
- Wang X, Edwards RL, Auler AS, Cheng H, Kong X, Wang Y, Cruz FW, Dorale JA, Chiang H-W. 2017. Hydroclimate changes across the Amazon lowlands over the past 45,000 years. *Nature* 541(7636):204–207 DOI 10.1038/nature20787.
- Warren BH, Simberloff D, Ricklefs RE, Aguilée R, Condamine FL, Gravel D, Morlon H, Mouquet N, Rosindell J, Casquet J, Conti E, Cornuault J, Fernández-Palacios JM, Hengl T, Norder SJ, Rijsdijk KF, Sanmartín I, Strasberg D, Triantis KA, Valente LM, Whittaker RJ, Gillespie RG, Emerson BC, Thébaud C. 2015. Islands as model systems in ecology and evolution: prospects fifty years after MacArthur-Wilson. *Ecology Letters* 18(2):200–217 DOI 10.1111/ele.12398.
- Webb CO, Ackerly DD, McPeck MA, Donoghue MJ. 2003. Phylogenies and community ecology. *Annual Review of Ecology and Systematics* 33(1):475–505 DOI 10.1146/annurev.ecolsys.33.010802.150448.
- Weber MG, Strauss SY. 2016. Coexistence in close relatives: beyond competition and reproductive isolation in sister taxa. *Annual Review of Ecology, Evolution, and Systematics* 47(1):359–381 DOI 10.1146/annurev-ecolsys-112414-054048.
- Weeks BC, Claramunt S, Cracraft J. 2016. Integrating systematics and biogeography to disentangle the roles of history and ecology in biotic assembly. *Journal of Biogeography* 43(8):1546–1559 DOI 10.1111/jbi.12747.
- Wegener A. 1912. Die Entstehung der Kontinente. *Geologische Rundschau* 3(4):276–292 DOI 10.1007/BF02202896.
- Werneck FP, Costa GC, Colli GR, Prado DE, Sites JW Jr. 2011. Revisiting the historical distribution of Seasonally Dry Tropical Forests: new insights based on palaeodistribution modelling and palynological evidence. *Global Ecology and Biogeography* 20(2):272–288 DOI 10.1111/j.1466-8238.2010.00596.x.
- Werneck FP, Gamble T, Colli GR, Rodrigues MT, Sites JW Jr. 2012a. Deep diversification and long-term persistence in the South American “dry diagonal”: integrating continent-wide phylogeography and distribution modeling of geckos. *Evolution* 66(10):3014–3034 DOI 10.1111/j.1558-5646.2012.01682.x.
- Werneck FP, Nogueira C, Colli GR, Sites JW, Costa GC. 2012b. Climatic stability in the Brazilian Cerrado: implications for biogeographical connections of South American savannas, species richness and conservation in a biodiversity hotspot. *Journal of Biogeography* 39(9):1695–1706 DOI 10.1111/j.1365-2699.2012.02715.x.
- Whittaker RJ, Triantis KA, Ladle RJ. 2008. A general dynamic theory of oceanic island biogeography. *Journal of Biogeography* 35(6):977–994 DOI 10.1111/j.1365-2699.2008.01892.x.
- Wiens JJ, Donoghue MJ. 2004. Historical biogeography, ecology and species richness. *Trends in Ecology & Evolution* 19(12):639–644 DOI 10.1016/j.tree.2004.09.011.

- Willig MR, Kaufman DM, Stevens RD. 2003.** Latitudinal gradients of biodiversity: pattern, process, scale, and synthesis. *Annual Review of Ecology, Evolution, and Systematics* **34**(1):273–309 DOI [10.1146/annurev.ecolsys.34.012103.144032](https://doi.org/10.1146/annurev.ecolsys.34.012103.144032).
- Wilson JB, Peet RK, Dengler J, Pärtel M. 2012.** Plant species richness: the world records. *Journal of Vegetation Science* **23**(4):796–802 DOI [10.1111/j.1654-1103.2012.01400.x](https://doi.org/10.1111/j.1654-1103.2012.01400.x).
- Wing SL, Herrera F, Jaramillo CA, Gómez-Navarro C, Wilf P, Labandeira CC. 2009.** Late Paleocene fossils from the Cerrejon Formation, Colombia, are the earliest record of Neotropical rainforest. *Proceedings of the National Academy of Sciences of the United States of America* **106**(44):18627–18632 DOI [10.1073/pnas.0905130106](https://doi.org/10.1073/pnas.0905130106).
- Worm B, Barbier EB, Beaumont N, Duffy JE, Folke C, Halpern BS, Jackson JBC, Lotze HK, Micheli F, Palumbi SR, Sala E, Selkoe KA, Stachowicz JJ, Watson R. 2006.** Impacts of Biodiversity Loss on Ocean Ecosystem Services. *Science* **314**(5800):787–790 DOI [10.1126/science.1132294](https://doi.org/10.1126/science.1132294).
- Zedane L, Hong-Wa C, Murienne J, Jezierski C, Baldwin BG, Besnard G. 2016.** Museomics illuminate the history of an extinct, paleoendemic plant lineage (Hesperelaea, Oleaceae) known from an 1875 collection from Guadalupe Island, Mexico. *Biological Journal of the Linnean Society* **117**(1):44–57 DOI [10.1111/bij.12509](https://doi.org/10.1111/bij.12509).
- Zhang C, Stadler T, Klopstein S, Heath TA, Ronquist F. 2016.** Total-evidence dating under the fossilized birth–death process. *Systematic Biology* **65**(2):228–249 DOI [10.1093/sysbio/syv080](https://doi.org/10.1093/sysbio/syv080).
- Zizka A, Ter Steege H, Pessoa MDCR, Antonelli A. 2017.** Finding needles in the haystack: where to look for rare species in the American tropics. *Ecography* **41**(2):321–330 DOI [10.1111/ecog.02192](https://doi.org/10.1111/ecog.02192).
- Zuquim G, Costa FRC, Prado J, Braga-Neto R. 2009.** Distribution of pteridophyte communities along environmental gradients in Central Amazonia, Brazil. *Biodiversity and Conservation* **18**(1):151–166 DOI [10.1007/s10531-008-9464-7](https://doi.org/10.1007/s10531-008-9464-7).