

1 **Genetic Determinants of Circulating Estrogen Levels, and**  
2 **Evidence of a Causal Effect of Estradiol on Bone Density in Men**

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78 ***Précis:*** *CYP19A1* was the main regulator of estrogen levels in this GWAS, with additional

79 loci on chromosome X and in *TRIM4* and *CYP11B1/B2*. Findings in the study strengthen the

80 importance of E2 for bone health.

81 ***Short title:*** Genetic Determinants of Estrogen Levels in Men

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100 **Abstract**

101 **Context**

102 Serum estradiol (E2) and estrone (E1) levels exhibit substantial heritability. No genome-wide  
103 association study (GWAS) of estrogen levels has been performed in men of European origin.

104 **Objective**

105 To investigate the genetic regulation of serum E2 and E1 in men.

106 **Design, setting and participants**

107 GWAS in 11,097 men of European origin from nine epidemiological cohorts.

108 **Main Outcome Measures**

109 Genetic determinants of serum E2 and E1 levels.

110 **Results**

111 Variants in/near *CYP19A1* demonstrated the strongest evidence for association with E2,  
112 resolving to three independent signals. Two additional independent signals were found on the  
113 X chromosome; *FAM9B*, rs5934505 (p-value  $3.4 \times 10^{-8}$ ) and *Xq27.3*, rs5951794 (p-value  $3.1$   
114  $\times 10^{-10}$ ). E1 signals were found in *CYP19A1* (rs2899472, p-value  $5.5 \times 10^{-23}$ ), in *TRIM4*  
115 (rs17277546,  $p = 5.8 \times 10^{-14}$ ) and in *CYP11B1/B2* (rs10093796, p-value  $1.2 \times 10^{-8}$ ).

116 E2 signals in *CYP19A1* and *FAM9B* were associated with bone mineral density (BMD).

117 Mendelian Randomization analysis suggested a causal effect of serum E2 on BMD in men. 1  
118 pg/ml genetically increased E2 was associated with a 0.048 SD increase in lumbar spine BMD  
119 (p-value  $2.8 \times 10^{-12}$ ).

120 In men and women combined, *CYP19A1* alleles associated with higher E2 levels were  
121 associated with lower degrees of insulin resistance.

## 122 **Conclusions**

123 Our findings confirm that *CYP19A1* is an important genetic regulator of E2 and E1 levels, and  
124 strengthen the causal importance of E2 for bone health in men. We also report 2 new  
125 independent loci on the X-chromosome for E2, one new locus each in *TRIM4* and  
126 *CYP11B1/B2*, for E1.

127 **Keywords:** estradiol, estrone, GWAS, men, BMD, insulin sensitivity

## 128 **Introduction**

129 17  $\beta$ -estradiol (E2) and estrone (E1) are the major biologically active estrogens in men. E2 is  
130 more potent than E1. Aromatase, encoded by the *CYP19A1* gene, is the key enzyme  
131 responsible for the final step in the synthesis of both E2 and E1. E2 is formed from  
132 aromatization of testosterone, and E1 is formed from aromatization of androstenedione. E2  
133 can also be formed from conversion of E1 by 17 $\beta$ -hydroxysteroid dehydrogenase (1).

134 In men, the circulating levels of E2 and E1 are determined by both genetic and environmental  
135 factors. The heritability for E2 in men has been estimated to be ~30-45% and for E1 ~40% (2,  
136 3). Early studies of the genetic regulation of circulating E2 and E1 levels were hampered by  
137 their small size and the use of immunoassays with poor specificity, precision and accuracy at  
138 lower concentrations. However, in 2010 Orwoll and colleagues performed a large study of  
139 5,000 elderly men of European, Asian and African origin in Sweden, the United States, Hong  
140 Kong and Tobago (4). Serum sex steroid levels were measured using gas chromatography –  
141 mass spectrometry (GC-MS), thereby avoiding the previously mentioned problems with  
142 immunoassays. In addition to geographical differences in E2 and E1 levels, suggestive of  
143 environmental influences, they also found racial differences. Both E2 and E1 levels, as well as  
144 the estradiol to testosterone and estrone to androstenedione ratios, were higher in Black than  
145 in Asian and Caucasian men (4). These data suggested that genetically determined differences  
146 in aromatase activity among Black, Asian and Caucasian men might be responsible for the  
147 observed racial differences in E2 and E1 levels.

148 We made a first attempt to find genetic loci involved in the determination of estrogen levels in  
149 men by analyzing 604 SNPs in 50 candidate sex steroid-related genes (5). In a screening  
150 cohort, the *CYP19A1* SNP rs2470152 showed the most significant association with E2 levels



151 measured by GC-MS. This was confirmed in two replication cohorts. Rs2470152 was also  
152 significantly associated with E1 levels in all three cohorts (n=5531) (5).

153 Meta-analyses of genome-wide association studies (GWAS) enable a comprehensive analysis  
154 of the whole genome in a large number of subjects. Chen and colleagues performed a GWAS  
155 in 3,495 Chinese men, where E2 concentrations were determined using an immunoassay.  
156 They found two independent SNPs in the *CYP19A1* gene to be associated with E2 levels  
157 (rs2414095 and rs2445762) (6). These findings further strengthened the evidence for a major  
158 role of *CYP19A1* in the regulation of serum E2 levels in men, but due to the relatively small  
159 sample size and low power, genetic loci in other regions of the genome could have been  
160 missed. To date no GWAS has been performed in men of European origin. In women, a  
161 smaller GWAS meta-analysis of 1,583 postmenopausal women found no genome-wide  
162 significant SNPs. Among variants that were suggestively associated with E2, several were  
163 located at the *CYP19A1* locus (7).

164 Both E2 and testosterone regulate bone mass (8). Studies of men with non-functional estrogen  
165 receptor alpha (ER $\alpha$ ) (9), and inactivating mutations of the *CYP19A1* gene (10), have  
166 demonstrated that estrogens are important for peak bone mass acquisition in men. Population  
167 based studies have shown that in men, low serum levels of E2 are associated with a lower  
168 bone mineral density (BMD), higher rates of bone loss and an increased risk of fractures (8,  
169 11-14). Some studies also show a smaller contribution of testosterone to BMD in men (8, 11).  
170 The relative contribution of androgens versus estrogens in the regulation of bone mass in men  
171 remains incompletely understood, and studies showing evidence of a causal effect of serum  
172 E2 on BMD in men are still sparse (15).

173 Mendelian randomization is a method used to strengthen or refute the causality of a  
174 biomarker, such as E2, and an outcome measure of interest, such as BMD, when a

175 randomized controlled trial is not possible. Mendelian randomization uses genetic data and  
176 relies on the principle that due to the random assortment of genetic variants at conception,  
177 these genetic variants are independent of many factors that bias observational studies, such as  
178 confounding and reverse causation. Therefore, if a biomarker is etiologically involved in an  
179 outcome measure, the genetic factors that influence the biomarker will also influence the  
180 outcome measure (16). To date, no Mendelian randomization has been performed to  
181 investigate causality between E2 levels and BMD in men.

182 Case reports of men with aromatase deficiency due to an inactivating mutation of the  
183 *CYP19A1* gene, mechanistic animal studies and clinical studies also suggest that estrogen  
184 signaling through ER $\alpha$  is important for insulin sensitivity in men (17-23). Thus, genetic  
185 factors regulating estrogen levels may also be of relevance for the regulation of insulin  
186 sensitivity in men.

187 Here we present the results of the first GWAS of estrogen levels combining several  
188 population-based cohorts of men of European origin. We also present results of our analyses  
189 of the association of resultant genome wide significant associations with two major estrogen  
190 related traits –bone mineral density and insulin sensitivity.

191 **Methods**

192 *Study samples*

193 The discovery stage of the E2 GWAS included 11,097 men of European origin drawn from  
194 nine epidemiological cohorts: the Framingham Heart Study (FHS), the Gothenburg  
195 Osteoporosis and Obesity Determinants (GOOD) study, the Invecchiare in Chianti  
196 (InCHIANTI) study, the LUdwigshafen RIsk and Cardiovascular Health (LURIC) study, the  
197 Multi-Ethnic Study of Atherosclerosis (MESA) study, the Osteoporotic Fractures in Men  
198 (MrOS) Sweden Gothenburg study, the MrOS Sweden Malmö study, the MrOS US Study,  
199 and the Rotterdam 1 (RS1) study. Replication of one SNP displaying considerable  
200 heterogeneity in genome wide significant fixed effect models, but nominal significance only  
201 in random effects models, was performed in the European Male Ageing Study (EMAS,  
202 n=1,641). EMAS is a cohort of men predominantly of European origin with only 0.62 %  
203 (n=21) of the sample used here being of non-European descent.

204 The discovery stage of the E1 GWAS included 7,570 men of European origin drawn from six  
205 of the above-mentioned cohorts: FHS, GOOD, MrOS Sweden Gothenburg, MrOS Sweden  
206 Malmö, MrOS US and RS1.

207 Exclusion criteria included chemical or surgical castration and/or medications affecting sex  
208 hormones such as steroid 5-alpha reductase inhibitors, and sex hormone antagonists. All  
209 studies were approved by local ethics committees and all participants provided written  
210 informed consent. Characteristics of the study samples and detailed descriptions of the  
211 participating cohorts, genotyping, quality control and imputation procedures are provided in  
212 the Supplementary Appendix and in Supplemental Tables 1, 2 and 3.

213

214 *Sex hormone measurements*

215 In six discovery cohorts (FHS, GOOD, MrOS Sweden Gothenburg, MrOS Sweden Malmö,  
216 and MrOs US), measurements of E1 and E2 were performed using either the GC-MS or the  
217 liquid chromatography tandem mass spectrometry (LC-MS/MS) technique. In the remaining  
218 discovery cohorts (LURIC, InCHIANTI, MESA and RS-1) measurements were performed  
219 using immunoassays. In the replication cohort (EMAS), E2 was measured using the GC-MS  
220 technique. Methods for all measurements are given in the Supplementary Appendix.

221 *Genotyping and statistical analyses*

222 Nine discovery and one replication study populations were genotyped using a variety of  
223 genotyping platforms including Illumina (HumanHap 550k, 610k, 1M-Duo, Omni1-Quad,  
224 Omni express) and Affymetrix (500K Dual GeneChip + 50K gene-centered MIP set, Array  
225 6.0) (Supplemental Table 2). To increase genomic coverage and allow the evaluation of the  
226 same SNPs across as many study populations as possible, each study imputed genotype data  
227 based on the HapMap CEU Build 36. Algorithms were used to infer unobserved genotypes in  
228 a probabilistic manner using either MACH (<http://www.sph.umich.edu/csg/abecasis/MACH>),  
229 or IMPUTE2 (24). We analyzed only those SNPs (genotyped or imputed) which had a minor  
230 allele frequency of  $>0.01$  and an imputation quality of  $\geq 0.3$ . The X chromosome was available  
231 for analysis in 6 cohorts (FHS, GOOD, LURIC, MrOS Sweden Gothenburg, MrOS Sweden  
232 Malmö and MrOS US), in this study. Imputations of the X-chromosome were performed in all  
233 of these cohorts except MrOS US.

234 Altogether, ~2.5 million SNPs were tested for association with serum E2 and E1 in the  
235 discovery stage. Genome-wide association analyses were performed using an additive genetic  
236 linear regression model adjusted for: 1) age and BMI (E2 and E1) or: 2) age, BMI,  
237 testosterone and SHBG (E2 only), in each of the discovery cohorts. In FHS, a linear mixed

238 effect model with a random effect to account for relationships was used. Imputed genotypes  
239 were analyzed in all cohorts taking the genotype uncertainties into account. The meta-  
240 analyses were performed in the METAL software  
241 (<https://www.sph.umich.edu/csg/abecasis/MACH>), using an inverse-variance weighted fixed  
242 effect model. Random effects models were used when fixed effect models displayed  
243 heterogeneity defined as an  $I^2$ -value  $> 50\%$  (25). These models were calculated using the R-  
244 package (<http://www.r-project.org>). A threshold of  $p < 5 \times 10^{-8}$  was established *a priori* as the  
245 level for genome-wide significance in the discovery analyses (26).

246 Approximate conditional analyses for E2 and E1 were performed using the Genome-wide  
247 Complex Trait Analysis (GCTA) software (27), and the genotypes of the EPIC Norfolk study  
248 cohort used as a reference panel to estimate patterns of Linkage Disequilibrium (28). The GC-  
249 corrected and quality control filtered meta-analysis results and a condition list containing the  
250 lead SNPs of the final loci were used as input for the conditional analysis. An additional  
251 association was declared when the conditional P-value was below the genome-wide  
252 significance threshold. Subsequently, this SNP was added to the list of conditional analysis  
253 SNPs and the conditional analysis was performed again in a stepwise fashion until no  
254 additional significant independent associations were found.

#### 255 *Gene expression analyses*

256 We analyzed associations between identified SNPs associated with serum estrogen levels and  
257 gene expression in the eQTL dataset generated by the GTEx Consortium (version 6p), which  
258 was obtained from <http://www.gtexportal.org/> (29)

#### 259 *Associations with testosterone*

260 Associations with serum testosterone concentrations were retrieved from the discovery dataset  
261 of our previously published GWAS of testosterone levels (30).

262 *Associations with other traits*

263 We hypothesized, based on data in the literature, that our genome wide significant SNPs and  
264 secondary signals from conditional analyses could be associated with BMD and/or insulin  
265 sensitivity. To test these hypotheses we searched publicly available databases for associations  
266 with lumbar spine (LS) and femoral neck (FN) BMD in men ([www.gefos.org](http://www.gefos.org)) (31). Data on  
267 glyceic traits in men and women combined were downloaded from  
268 <http://www.magicinvestigators.org/downloads/> (32, 33). Data on glyceic traits in men and  
269 women separately were contributed by MAGIC investigators (32, 33). HOMA-IR was  
270 calculated as (fasting insulin x fasting glucose)/22.5.

271 *Mendelian Randomization of serum E2 on BMD*

272 To investigate if E2 has a causal effect on BMD we performed a summary statistic two  
273 sample inverse variance weighted Mendelian Randomization (34). We selected the 5 top loci  
274 from our E2 meta-analysis and extracted summary statistics ( $\beta$  and SE) from the  
275 corresponding SNPs in both our E2 study and the GEFOS study on LS and FN BMD. The  
276 variant specific associations were used to create an inverse variance weighted estimate of the  
277 causal effect size and its standard error.

278

## 279 **Results**

280 We performed a GWAS of serum E2 and E1 concentrations, investigating ~2.5 million SNPs  
281 in up to 11,097 men. In analyses of autosomal chromosomes, all 9 discovery cohorts  
282 (n=11,097) were included in the discovery analyses of E2, and six cohorts (n=7,570) were  
283 included in the discovery analyses of E1.

284 In analyses of the X-chromosome, six cohorts (n=8,953) were included in the discovery  
285 analyses of E2, and five cohorts (n=6,917) were included in the discovery analyses of E1.

### 286 *Estradiol*

287 In the model adjusted for age and BMI (Model 1), two loci were associated with E2  
288 concentrations at the genome-wide significance threshold of  $p < 5 \times 10^{-8}$  in the discovery  
289 analyses (Figure S1A). The strongest association was found within the *CYP19A1* locus on  
290 chromosome 15q21.1 (rs727479, effect size 1.39 pg/ml per effect allele, (SE 0.12),  $p = 8.2 \times$   
291  $10^{-30}$ ) (Table 1, Figures 1A, S2A, S3A). This SNP, which is located in the second intron of the  
292 gene, showed heterogeneity of effect size across studies as indicated by an  $I^2$  value of 57%  
293 (25). To take this heterogeneity into account, we additionally calculated a random effects  
294 model, which was also genome-wide significant (effect size = 1.35 pg/ml SE 0.19,  $p = 2.0 \times$   
295  $10^{-12}$ ).

296 The second locus was found on the X-chromosome where one SNP, rs5934505, reached  
297 genome-wide significance ( $p = 3.4 \times 10^{-8}$ ). Rs5934505 is located 79 kb downstream of the  
298 *FAMily with sequence similarity 9, member B (FAM9B)* gene (Xp22.31) (Table 1, Figures 1A,  
299 S2B, S3B). There was heterogeneity of effect size across studies for this SNP ( $I^2 = 72\%$ ). A  
300 random effects model displayed nominal, but not genome-wide, significance in the same  
301 direction as the result from the fixed effect meta-analysis (C-allele associated with higher E2

302 levels, effect size 0.74 pg/ml per effect allele (SE 0.24),  $p = 0.002$ ). Therefore, we attempted  
303 replication for rs5934505 in the EMAS cohort ( $n = 1,641$ ). In this cohort, the C-allele was  
304 also associated with higher E2 levels; effect size of 1.59 pg/ml per effect allele (SE 0.39),  $p$ -  
305 value  $5.2 \times 10^{-5}$ .

306 In the model that was adjusted for testosterone and SHBG levels, in addition to age and BMI  
307 (Model 2 (Figure S1B)), the associations between E2 and the *CYP19A1* locus remained  
308 significant (rs727479:  $p = 3.1 \times 10^{-43}$  (Table 1, Figures 1B, S2C, S3C)). In this analysis, the  $I^2$   
309 value was 69%, but the random effects model was genome wide significant (effect size 1.42  
310 pg/ml per effect allele (SE 0.20),  $p$ -value  $3.5 \times 10^{-13}$ ). A novel genome wide significant locus  
311 on the X-chromosome also appeared in this analysis. Rs5951794 ( $p = 3.1 \times 10^{-10}$ ,  $I^2 = 6\%$ ) is  
312 located in the distal part of the long arm on chromosome X (Xq27.3), approximately 137 Mb  
313 from the *FAM9B* SNP rs5934505 (Table 1, Figures 1B, S2D, S3D).

314 To identify multiple statistically independent SNPs within the same genomic region, we  
315 performed stepwise approximate conditional analyses (GCTA) for each of the genome-wide  
316 significant loci. In the model adjusted for testosterone and SHBG, the analysis revealed two  
317 additional genome-wide significant SNPs in *CYP19A1* locus; rs2899472 in intron 4  
318 (conditional  $p$ -value  $1.1 \times 10^{-8}$ ) and rs16964258 in intron 1 (conditional  $p$ -value  $8.2 \times 10^{-15}$ )  
319 (Table 1, Figures 1B, S2C, S3E-F). In the model adjusted for age and BMI only, no additional  
320 independent associations were found.

321 In Model 1, rs727479 explained 0.9% of the overall variance of E2 levels. When rs5934505  
322 (*FAM9B*) was added, 1.1% of the variance was explained. In Model 2, independent *CYP19A1*  
323 SNPs explained 1.3% of the overall variance in E2 levels. When rs5951794 (*Chr X*) was  
324 added, 1.4% of the variance was explained.



325 In Model 1, rs727479 explained 0.9% of the overall variance of E2 levels. When the other  
326 identified SNP from Model 1, rs5934505 (*FAM9B*) was added, 1.1% of the overall variance in  
327 E2 levels was explained. In Model 2, independent *CYP19A1* SNPs explained 1.3% of the  
328 overall variance in E2 levels. When the other genome wide significant SNP from Model 2,  
329 rs5951794 (*Chr X*), was added, 1.4% of the overall variance in E2 levels was explained.

### 330 *Estrone*

331 Three genome-wide significant loci, located on chromosomes 7, 8 and 15, respectively, were  
332 associated with E1 levels (Figure S1C). The strongest association was found for the *CYP19A1*  
333 locus on chromosome 15. The lead SNP was rs2899472 ( $p = 5.5 \times 10^{-23}$ ) (Table 1, Figures 1C,  
334 S2E, S3G). Because of heterogeneity in effect size at this variant ( $I^2 = 59\%$ ), a random effects  
335 model was run, which was genome wide significant (effect size 2.55 pg/ml per effect allele,  
336 SE 0.41,  $p = 4.6 \times 10^{-10}$ ). In conditional analyses of this locus, the SNP with the most  
337 significant association with E2, rs727479, was also genome wide significantly associated with  
338 E1 (conditional p-value  $3.5 \times 10^{-10}$ ) (Table 1, Figures 1C, S2E, S3H).

339 On chromosome 7, the SNP most significantly associated with E1 levels was rs17277546 ( $p =$   
340  $5.8 \times 10^{-14}$ ), located in the 3' UTR of the *Tripartite motif containing 4 (TRIM4)* gene (Table 1,  
341 Figures 1C, S2F, S3I). On chromosome 8, the SNP most significantly associated with E1  
342 levels was rs10093796 ( $p = 1.2 \times 10^{-8}$ ). This SNP is located between the *CYP11B1* and the  
343 *CYP11B2* genes (Table 1, Figures 1C, S2G, S3J).

344 Estrone is not derived from testosterone and not bound to SHBG in the circulation. Therefore  
345 no analyses of E1 adjusted for these parameters were performed.

346 Independent *CYP19A1* SNPs explained 1.5% of the overall variance in E1 levels.

347 Rs17277546 (*TRIM4*) and rs10093796 (*CYP11B1/B2*) explained 0.5% and 0.1% respectively

348 of the variance. In total, 2.1% of the overall variance in E1 levels was explained by these  
349 genome wide significant SNPs.

### 350 *Gene expression analyses*

351 In the GTEx database, two of the *CYP19A1* SNPs were robustly associated with the  
352 expression level of *CYP19A1*. The alleles associated with higher E2 levels were associated  
353 with higher gene expression levels (rs727479:  $\beta$  0.23,  $p = 1.9 \times 10^{-5}$  (skin), and rs2899472:  $\beta$   
354 0.20,  $p = 9 \times 10^{-8}$  (whole blood)). Rs727479 was also associated with the expression level of  
355 *signal peptide peptidase like 2A (SPPL2A)* ( $\beta$  0.18,  $p = 1.3 \times 10^{-4}$  (transformed fibroblasts)).  
356 *SPPL2A* is located 442 kB upstream of *CYP19A1*. The E1 associated SNP on chromosome 8,  
357 rs10093796, was associated with the expression levels of two adjacent genes in several tissues  
358 (*Lys6/Neurotoxin1 (LYNX1)*) pancreas  $\beta$  0.68,  $p = 5.6 \times 10^{-9}$  and *Lymphocyte Antigen 6*  
359 *Complex, Locus K (LY6K)* skin  $\beta$  0.32,  $p=2.3 \times 10^{-7}$ ). *LYNX1* and *LY6K* are located 95 kB and  
360 168 kB respectively upstream of *CYP11B1*. The other SNPs in our study were not associated  
361 with expression levels in the GTEx database.

### 362 *Associations with estrogen related traits*

363 To further investigate the physiological relevance of our E2 GWAS findings, we performed  
364 look up analyses of other GWAS which had data on phenotypes known or suspected to be  
365 related to E2 levels.

### 366 *Testosterone*

367 To better understand the mechanism underlying the association between our E2-related SNPs  
368 and E2 levels, we studied the association between these SNPs and serum testosterone levels.  
369 If the effect of the SNPs on E2 levels was exerted upstream of the aromatase enzyme, one  
370 would expect that SNPs to be associated with higher testosterone as well as higher E2 levels.  
371 On the other hand, if the effect of the SNPs on E2 levels were exerted through alteration in

372 either the amount or the activity of the aromatase enzyme, only E2 levels would be expected  
373 to be increased, with no increase in testosterone levels. The C-allele of the E2 X chromosome  
374 SNP rs5934505 (*FAM9B*) was positively associated with levels of both testosterone and E2,  
375 suggesting that the effect of rs5934505 is exerted upstream of aromatase (Table 2, Figure 2).  
376 Indeed, we have previously reported that the X chromosome SNP rs5934505 (*FAM9B*) is  
377 associated with circulating testosterone levels in men ( $p = 1.6 \times 10^{-8}$ ) (30). None of the other  
378 E2 SNPs were associated with increased levels of testosterone, suggesting that these SNPs are  
379 affecting either the amount or the activity of aromatase or estradiol clearance. In fact, the G-  
380 allele of the other E2 X-chromosome SNP, rs5951794, was associated with increased E2  
381 levels and slightly decreased testosterone levels (effect size -7.68 ng/dl per effect allele (SE  
382 3.05),  $p = 0.01$ ) (Table 2, Figure 2). Additionally, for *CYP19A1* SNPs, there were indications  
383 of associations with testosterone in the opposite direction compared to E2, but these  
384 associations did not reach statistical significance (rs727479  $p = 0.05$  and rs16964258  $p =$   
385 0.26) (Table 2, Figure 2).

#### 386 *BMD*

387 The primary SNP in *CYP19A1*, rs727479, and the secondary signals rs2899472 and  
388 rs16964258, were all significantly associated with LS BMD in men ( $p \leq 0.01$ ; Table 3).  
389 Rs727479 and rs2899472 were also associated with FN BMD in men ( $p < 0.01$ ). The  
390 direction of the effect was the same for all markers, *i.e.* alleles associated with higher levels of  
391 E2 were associated with a higher BMD. Moreover, rs5934505 (*FAM9B*) was associated with  
392 both FN ( $p = 0.01$ ) and LS ( $p = 7 \times 10^{-6}$ ) BMD. As in the case of *CYP19A1* SNPs, the allele  
393 associated with higher E2 levels was associated with a higher BMD (Table 3).

394

395

396 *Mendelian Randomization E2 and BMD*

397 The data from the GEFOS database show associations between individual SNPs and BMD,  
398 but do not provide information on possible causality between the E2 levels resulting from  
399 these SNPs and BMD. To overcome this we performed a summary statistic Mendelian  
400 Randomization analysis which suggested that there is a causal effect of serum E2 on BMD. A  
401 1 pg/ml genetically increased E2 was associated with a 0.048 SD (SE 0.008),  $p = 2.8 \times 10^{-12}$   
402 increase in LS BMD. For the femoral neck the increase was 0.037 SD (SE 0.007,  $p = 4.4 \times 10^{-8}$   
403 (Figure 3).

404 *Insulin sensitivity*

405 The publicly available GWAS results for measures of insulin sensitivity included only  
406 autosomal chromosomes, and did not include results for men and women separately. Thus the  
407 following results apply for men and women combined. Insulin resistance expressed as  
408 HOMA-IR was negatively associated with the E2 increasing A-alleles of rs727479 ( $p =$   
409 0.004) and rs2899472 ( $p = 0.003$ ) in *CYP19A1*. This was due to a negative association of  
410 these alleles with fasting insulin ( $p = 0.003$  for rs727479 and  $p = 0.017$  for rs2899472)  
411 (Supplemental Table 4). Adjustments for BMI had no effect on the results (BMI-adjusted  
412 fasting insulin  $p = 0.002$  for rs727479 and  $p = 0.031$  for rs2899472). There were no  
413 associations with fasting glucose for these SNPs. The MAGIC investigators also provided us  
414 with data not publicly available on fasting insulin and fasting glucose for men and women  
415 separately (fasting insulin: men  $n \approx 26,000$ , women  $n \approx 32,000$ , fasting glucose: men  $n \approx$   
416 36,000, women  $n \approx 43,000$ ). In this dataset, the association between rs727479 and fasting  
417 insulin was significant in women ( $\beta -0.014$  (SE 0.004),  $p = 0.002$ ). In men the direction of the  
418 association was the same as in women, but was not statistically significant (rs727479:  $\beta -$   
419 0.006 (SE 0.005),  $p = 0.19$ ).

## 420 **Discussion**

421 In this GWAS, SNPs in the *CYP19A1* gene showed the strongest associations with both E1  
422 and E2 levels. This confirms data from previous studies (5, 6, 35) and establishes *CYP19A1* as  
423 an important genetic regulator of estrogen levels in men. We found three independent signals  
424 in *CYP19A1*, which extends the results from previous studies. We also identified two  
425 additional signals for E2 on chromosome X and two additional signals for E1, on  
426 chromosomes 7 and 8, respectively. Moreover, SNPs found to be associated with E2 levels in  
427 this study were also associated with known or suspected estrogen-related traits including  
428 BMD and insulin sensitivity. Mendelian randomization analysis using the independent E2  
429 SNPs suggests a causal effect of E2 on BMD in men.

430 The finding of several independent signals for both E1 and E2 in *CYP19A1* is consistent with  
431 the findings in the previously reported GWAS in Chinese men, where two independent SNPs  
432 were found. This strengthens the conception that the regulation of estrogen levels is governed  
433 by more than one signal in the gene. The organization of *CYP19A1* is rather complex. The  
434 gene consists of a 30-kb coding region and a 93-kb regulatory region including 10 tissue-  
435 specific promoters (36). There are four blocks of linkage disequilibrium (LD) in the gene.  
436 Rs727479, which displayed the most significant association with E2 levels in our study, is  
437 located in intron 2 in LD block 4, which covers 50 kB including the entire coding region,  
438 exons/promoters I.6, I.3 and PII, through 5.8 kb downstream of exon 10 (37). Rs727479 has  
439 been associated with E2 levels in previous candidate gene studies investigating haplotype-  
440 tagging SNPs in *CYP19A1*, as well as in more comprehensive studies investigating larger  
441 numbers of SNPs in many genes, in both men (35, 38, 39) and postmenopausal women (40).  
442 Moreover, rs727479 was the second most significant SNP in the GWAS of E2 levels in  
443 postmenopausal women performed by Prescott and colleagues, although it did not reach

444 genome-wide significance ( $p = 5 \times 10^{-7}$ ), perhaps due to the relatively low number of study  
445 participants (7). In all of these studies, the direction of the effect was the same as in our study:  
446 the A-allele was associated with higher E2 levels. The most significant SNP in the male  
447 Chinese GWAS performed by Chen and colleagues, rs2414095, is in very strong linkage ( $r^2 =$   
448 0.96) with rs727479 (6), and it is also located in intron 3 in LD block 4. The findings from our  
449 gene expression analyses that rs727479 is associated with the expression of *CYP19A1* in two  
450 tissues further support the relevance of this SNP in the regulation of E2 levels.

451 To our knowledge, the *CYP19A1* loci rs2899472 and rs16964258 have not been linked to E2  
452 levels in previous studies. Rs2899472 is located in intron 4, in LD-block 4. Rs16964258 is  
453 located in a different region of the gene; intron 1, between LD blocks 1 and 2. Interestingly,  
454 the SNP most significantly associated with estrogen levels in our previous extended candidate  
455 gene study, rs2470152 (5), is also located in this region, 10 kb downstream of rs16964258.  
456 The  $D'$  for rs2470152 and rs16964258 is 1.0 but the  $r^2$  is 0.062, indicating that the SNPs are  
457 probably linked but, due to different allele frequencies, they are not proxy SNPs of one  
458 another.

459 The signal in the *FAM9B* region on the X-chromosome, rs5934505, has not been associated  
460 with E2 levels before, but associations of this locus with testosterone levels are known from  
461 our earlier testosterone GWAS (30), a finding which was later replicated by Jin and  
462 colleagues in a smaller GWAS in men ( $n = 3,225$ ) (41). Because testosterone is the precursor  
463 of estradiol, it is likely that the association of rs5934505 in the *FAM9B* region with E2 levels  
464 is mediated through the regulation of testosterone production and not through the conversion  
465 of testosterone to E2 per se. Rs5934505 is located in a CNV-insertion area (Xp22), 145 kb  
466 upstream of the *family with sequence similarity 9, member A* (*FAM9A*), and 79 kb  
467 downstream of *family with sequence similarity 9, member B* (*FAM9B*) genes. Both genes are  
468 expressed exclusively in the testes, and share 46% amino acid identity. Very little is known

469 about their functions (42). The *Kallman syndrome 1 (KALI)* gene is located 214 kb  
470 downstream of rs5934505. *KALI* encodes the extracellular matrix glycoprotein anosmin-1  
471 implicated in the embryonic migration of gonadotropin releasing hormone and olfactory  
472 neurons. Deleterious mutations in *KALI* cause X-linked Kallmann syndrome, characterized  
473 by hypogonadotropic hypogonadism and anosmia (43), but there are no previous data  
474 supporting that minor alterations in the function of KAL1 are associated with sex steroid  
475 levels. Moreover, rs5934505 is correlated ( $r^2 = 0.35$ ) with another SNP, rs5978985, in this  
476 region, which was associated with male puberty in a recent GWAS (44).

477 The other signal on the X chromosome, rs5951794, has not previously been associated with  
478 sex steroid levels, and the mechanism underlying the association in our study is not known. In  
479 contrast to rs5934505 (*FAM9B*), rs5951794 was not associated with higher testosterone  
480 levels. Therefore, the effect of this SNP would be expected to be exerted through alteration in  
481 the amount or activity of the aromatase enzyme or through regulation of estradiol clearance.  
482 In fact, rs5951794 was associated with slightly lower levels of testosterone. This might be the  
483 result of E2 mediated suppression of LH, which in turn would result in decreased testosterone  
484 levels. Rs5951794 is located approximately 65 kb downstream of a region rich in micro-  
485 RNAs (*MIRs 506-510, 513-514*), expressed mainly in the testes (45). Aside from the micro-  
486 RNA cluster, *Fragile X mental retardation 1 (FMRI)* is the closest gene located  
487 approximately 700 kb downstream of rs5951794. Keeping the distance in mind, one could  
488 speculate that rs5951794 could affect the regulation of *FMRI*, a gene which in addition to its  
489 crucial role in the pathogenesis of Fragile X Syndrome associated mental retardation, is also  
490 the leading molecular cause of premature ovarian failure (46).

491 The E1 signal rs17277546 in the *TRIM4* gene has also been shown to be associated in our  
492 previous GWAS of dehydroepiandrosterone sulphate (DHEAS) concentrations (47). Serum  
493 levels of DHEAS and DHEA are highly collinear (48). Serum levels of DHEAS could

494 therefore be a marker of serum levels of DHEA. In our earlier GWAS, the G allele was  
495 associated with higher levels of DHEAS, and in the present study, the G allele was associated  
496 with higher levels of estrone. Thus, an increased amount of adrenal derived precursors for  
497 estrogen synthesis is a possible explanation for the present findings. *TRIM4* is a member of  
498 the *tripartite motif (TRIM) family*. Members of this family have been implicated in many  
499 biological processes including cell differentiation, apoptosis and transcriptional regulation  
500 (49). The mechanism relating rs17277546 to DHEAS levels is not known, but in our previous  
501 GWAS, we found that rs17277546 is strongly associated with expression levels of *TRIM4* in  
502 cell lines from liver and adipose tissue in publically available databases. This indicates that  
503 rs17277546 is a functional SNP, or linked to such a SNP (47).

504 The chromosome 8 signal, rs10093618 is located 1.5 kb upstream of the *CYP11B1* gene. The  
505 product of *CYP11B1*, the steroid 11 $\beta$ -hydroxylase enzyme, catalyzes the conversion of 11-  
506 deoxycortisol to cortisol, representing the final step in cortisol biosynthesis, and 11-  
507 deoxycorticosterone to corticosterone. Deficiency of this enzyme leads to congenital adrenal  
508 hyperplasia. Hyperandrogenism is a hallmark of this condition since accumulated precursors  
509 are shunted into the androgen synthesis pathway (50). One could thus speculate that  
510 rs10093618, or an unknown variant linked with it, affects the production or efficiency of the  
511 steroid 11 $\beta$ -hydroxylase enzyme and thereby regulates the level of adrenal precursors for the  
512 sex steroid synthesis pathway, notably androstenedione, which is a direct precursor in estrone  
513 biosynthesis.

514 Because serum E2 levels in men are positively associated with BMD, the SNPs associated  
515 with higher E2 levels would be expected to be associated with higher BMD. In fact, in our  
516 previous extended candidate gene study there was such an association between the lead  
517 *CYP19A1* SNP, rs2470152, and BMD (5). Thus, the association in the present study between  
518 E2 associated SNPs in *CYP19A1* as well as *FAM9B*, and BMD is a plausible finding. In fact,



519 rs5934505 is in complete linkage,  $r^2$  1.0, with rs5934507, which was identified as the only  
520 male specific signal in our previous GWAS of BMD (31). Because of the known association  
521 of rs5934505 with testosterone, the BMD signal was thought to be mediated via testosterone  
522 levels in the BMD GWAS. Given the findings in the present study of an association between  
523 rs5934505 and E2, it seems more likely that the association with BMD is mediated at least in  
524 part via E2 levels rather than solely via a direct effect of testosterone (Figure 3).

525 Although an association between serum E2 levels and BMD in men has been shown in earlier  
526 association studies, a causal relation has not been demonstrated. In this study, using  
527 Mendelian Randomization analysis, we provide evidence that there is a causal effect of E2 on  
528 BMD. For instance, in the RS-1 cohort, where the E2 levels were 12.7 pg/ml (SD 6.6), 1 SD  
529 of genetically instrumented decrease in E2 would result in a  $6.6 \times 0.048 = 0.32$  SD decrease in  
530 LS BMD and  $6.6 \times 0.037 = 0.24$  SD decrease in FN BMD.

531 According to Johnell and coworkers, the relative risk for hip fracture in men aged 65 was 2.94  
532 (95 % CI 2.02-4.27) for each SD decrease in FN BMD (51). Using this information of the  
533 association between FN-BMD and hip fracture risk together with the causal effect of serum  
534 estradiol on FN-BMD as estimated in the present MR analysis, 1 SD (using the SD of serum  
535 estradiol from the RS-1 cohort) decrease in genetically instrumented E2 level could increase  
536 the relative risk for hip fracture by 47 %.

537 In this study, SNPs in *CYP19A1* that were associated with higher E2 levels, were also  
538 associated with improved insulin sensitivity and lower fasting insulin in men and women  
539 combined. In men, the role of estrogens in the regulation of insulin sensitivity is not fully  
540 understood. However, mechanistic studies and clinical trials suggest that estrogen signaling is  
541 important in the regulation of insulin sensitivity in men (18, 20, 22, 23). Furthermore, men  
542 with aromatase deficiency due to an inactivating mutation of the *CYP19A1* gene are

543 overweight or obese, and display and insulin resistance, which often improves with estrogen  
544 replacement therapy (17).

545 The strengths of our study include the large sample size, with 11,097 men in the discovery  
546 analysis of E2 levels, and the large proportion of serum samples analyzed using the MS-  
547 technique. This enabled us to find multiple signals in the *CYP19A1* locus, and new signals on  
548 other chromosomes, for both E1 and E2. A potential weakness of our study is that not all  
549 samples were analyzed by MS. As a result of the lower specificity of the immunoassays,  
550 weaker genetic signals might have been missed. It is likely that future studies with even larger  
551 numbers of samples analyzed by MS could uncover signals not found in this study.  
552 Nevertheless we believe that due to the large proportion of samples analyzed by MS our  
553 findings are robust and the risk for false positive signals is low. We also found SNP  
554 associations with BMD and measures of insulin sensitivity. Additionally, the Mendelian  
555 Randomization analysis provides evidence of a causal effect of E2 on BMD in men. The  
556 mechanisms underlying some of the associations in our study should be further investigated to  
557 expand our understanding of the regulation of sex steroid levels.

558

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562 **References**

- 563 1. Payne AH, Hales DB. Overview of steroidogenic enzymes in the pathway from cholesterol  
564 to active steroid hormones. *Endocr Rev.* 2004;25(6):947-70.
- 565 2. Travison TG, Zhuang WV, Lunetta KL, Karasik D, Bhasin S, Kiel DP, Coviello AD,  
566 Murabito JM. The heritability of circulating testosterone, oestradiol, oestrone and sex  
567 hormone binding globulin concentrations in men: the Framingham Heart Study. *Clin*  
568 *Endocrinol (Oxf).* 2014;80(2):277-82.
- 569 3. Bogaert V, Taes Y, Konings P, Van Steen K, De Bacquer D, Goemaere S, Zmierzak H,  
570 Crabbe P, Kaufman JM. Heritability of blood concentrations of sex-steroids in relation to  
571 body composition in young adult male siblings. *Clin Endocrinol (Oxf).* 2008;69(1):129-35.
- 572 4. Orwoll ES, Nielson CM, Labrie F, Barrett-Connor E, Cauley JA, Cummings SR, Ensrud K,  
573 Karlsson M, Lau E, Leung PC, Lunggren O, Mellstrom D, Patrick AL, Stefanick ML,  
574 Nakamura K, Yoshimura N, Zmuda J, Vandenput L, Ohlsson C. Evidence for geographical  
575 and racial variation in serum sex steroid levels in older men. *J Clin Endocrinol Metab.*  
576 2010;95(10):E151-60.
- 577 5. Eriksson AL, Lorentzon M, Vandenput L, Labrie F, Lindersson M, Syvanen AC, Orwoll  
578 ES, Cummings SR, Zmuda JM, Ljunggren O, Karlsson MK, Mellstrom D, Ohlsson C.  
579 Genetic variations in sex steroid-related genes as predictors of serum estrogen levels in men. *J*  
580 *Clin Endocrinol Metab.* 2009;94(3):1033-41.
- 581 6. Chen Z, Tao S, Gao Y, Zhang J, Hu Y, Mo L, Kim ST, Yang X, Tan A, Zhang H, Qin X,  
582 Li L, Wu Y, Zhang S, Zheng SL, Xu J, Mo Z, Sun J. Genome-wide association study of sex  
583 hormones, gonadotropins and sex hormone-binding protein in Chinese men. *J Med Genet.*  
584 2013;50(12):794-801.
- 585 7. Prescott J, Thompson DJ, Kraft P, Chanock SJ, Audley T, Brown J, Leyland J, Folkard E,  
586 Doody D, Hankinson SE, Hunter DJ, Jacobs KB, Dowsett M, Cox DG, Easton DF, De Vivo I.

587 Genome-wide association study of circulating estradiol, testosterone, and sex hormone-  
588 binding globulin in postmenopausal women. *PLoS One*. 2012;7(6):e37815.

589 8. Ohlsson C, Borjesson AE, Vandenput L. Sex steroids and bone health in men. *Bonekey*  
590 *Rep*. 2012;1:2.

591 9. Smith EP, Boyd J, Frank GR, Takahashi H, Cohen RM, Specker B, Williams TC, Lubahn  
592 DB, Korach KS. Estrogen resistance caused by a mutation in the estrogen-receptor gene in a  
593 man. *N Engl J Med*. 1994;331(16):1056-61.

594 10. Morishima A, Grumbach MM, Simpson ER, Fisher C, Qin K. Aromatase deficiency in  
595 male and female siblings caused by a novel mutation and the physiological role of estrogens.  
596 *J Clin Endocrinol Metab*. 1995;80(12):3689-98.

597 11. Khosla S, Melton LJ, 3rd, Robb RA, Camp JJ, Atkinson EJ, Oberg AL, Rouleau PA,  
598 Riggs BL. Relationship of volumetric BMD and structural parameters at different skeletal  
599 sites to sex steroid levels in men. *J Bone Miner Res*. 2005;20(5):730-40.

600 12. Mellstrom D, Vandenput L, Mallmin H, Holmberg AH, Lorentzon M, Oden A,  
601 Johansson H, Orwoll ES, Labrie F, Karlsson MK, Ljunggren O, Ohlsson C. Older men with  
602 low serum estradiol and high serum SHBG have an increased risk of fractures. *J Bone Miner*  
603 *Res*. 2008;23(10):1552-60.

604 13. Amin S, Zhang Y, Felson DT, Sawin CT, Hannan MT, Wilson PW, Kiel DP. Estradiol,  
605 testosterone, and the risk for hip fractures in elderly men from the Framingham Study. *Am J*  
606 *Med*. 2006;119(5):426-33.

607 14. Amin S, Zhang Y, Sawin CT, Evans SR, Hannan MT, Kiel DP, Wilson PW, Felson DT.  
608 Association of hypogonadism and estradiol levels with bone mineral density in elderly men  
609 from the Framingham study. *Ann Intern Med*. 2000;133(12):951-63.

610 15. Finkelstein JS, Lee H, Leder BZ, Burnett-Bowie SA, Goldstein DW, Hahn CW, Hirsch  
611 SC, Linker A, Perros N, Servais AB, Taylor AP, Webb ML, Youngner JM, Yu EW. Gonadal

612 steroid-dependent effects on bone turnover and bone mineral density in men. *J Clin Invest.*  
613 2016;126(3):1114-25.

614 16. Lawlor DA, Harbord RM, Sterne JA, Timpson N, Davey Smith G. Mendelian  
615 randomization: using genes as instruments for making causal inferences in epidemiology. *Stat*  
616 *Med.* 2008;27(8):1133-63.

617 17. Zirilli L, Rochira V, Diazzi C, Caffagni G, Carani C. Human models of aromatase  
618 deficiency. *J Steroid Biochem Mol Biol.* 2008;109(3-5):212-8.

619 18. Van Sinderen ML, Steinberg GR, Jorgensen SB, To SQ, Knowler KC, Clyne CD,  
620 Honeyman J, Chow JD, Herridge KA, Jones ME, Simpson ER, Boon WC. Hepatic glucose  
621 intolerance precedes hepatic steatosis in the male aromatase knockout (ArKO) mouse. *PLoS*  
622 *One.* 2014;9(2):e87230.

623 19. Zhu L, Martinez MN, Emfinger CH, Palmisano BT, Stafford JM. Estrogen signaling  
624 prevents diet-induced hepatic insulin resistance in male mice with obesity. *Am J Physiol*  
625 *Endocrinol Metab.* 2014;306(10):E1188-97.

626 20. Davis KE, M DN, Sun K, W MS, J DB, J AZ, Zeve D, L DH, D WC, L MG, Xu Y, Z  
627 VW, S AK, Clegg DJ. The sexually dimorphic role of adipose and adipocyte estrogen  
628 receptors in modulating adipose tissue expansion, inflammation, and fibrosis. *Mol Metab.*  
629 2013;2(3):227-42.

630 21. Cooke PS, Heine PA, Taylor JA, Lubahn DB. The role of estrogen and estrogen receptor-  
631 alpha in male adipose tissue. *Mol Cell Endocrinol.* 2001;178(1-2):147-54.

632 22. Juang PS, Peng S, Allehmazdeh K, Shah A, Coviello AD, Herbst KL. Testosterone with  
633 dutasteride, but not anastrozole, improves insulin sensitivity in young obese men: a  
634 randomized controlled trial. *J Sex Med.* 2014;11(2):563-73.

635 23. Finkelstein JS, Lee H, Burnett-Bowie SA, Pallais JC, Yu EW, Borges LF, Jones BF,  
636 Barry CV, Wulczyn KE, Thomas BJ, Leder BZ. Gonadal steroids and body composition,  
637 strength, and sexual function in men. *N Engl J Med.* 2013;369(11):1011-22.

638 24. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method  
639 for the next generation of genome-wide association studies. *PLoS Genet.* 2009;5(6):e1000529.

640 25. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-  
641 analyses. *BMJ.* 2003;327(7414):557-60.

642 26. Pe'er I, Yelensky R, Altshuler D, Daly MJ. Estimation of the multiple testing burden for  
643 genomewide association studies of nearly all common variants. *Genet Epidemiol.*  
644 2008;32(4):381-5.

645 27. Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, Steinthorsdottir V, Thorleifsson G,  
646 Zillikens MC, Speliotes EK, Magi R, Workalemahu T, White CC, Bouatia-Naji N, Harris TB,  
647 Berndt SI, Ingelsson E, Willer CJ, Weedon MN, Luan J, Vedantam S, Esko T, Kilpelainen  
648 TO, Kutalik Z, Li S, Monda KL, Dixon AL, Holmes CC, Kaplan LM, Liang L, Min JL,  
649 Moffatt MF, Molony C, Nicholson G, Schadt EE, Zondervan KT, Feitosa MF, Ferreira T,  
650 Lango Allen H, Weyant RJ, Wheeler E, Wood AR, Estrada K, Goddard ME, Lettre G,  
651 Mangino M, Nyholt DR, Purcell S, Smith AV, Visscher PM, Yang J, McCarroll SA, Nemesh  
652 J, Voight BF, Absher D, Amin N, Aspelund T, Coin L, Glazer NL, Hayward C, Heard-Costa  
653 NL, Hottenga JJ, Johansson A, Johnson T, Kaakinen M, Kapur K, Ketkar S, Knowles JW,  
654 Kraft P, Kraja AT, Lamina C, Leitzmann MF, McKnight B, Morris AP, Ong KK, Perry JR,  
655 Peters MJ, Polasek O, Prokopenko I, Rayner NW, Ripatti S, Rivadeneira F, Robertson NR,  
656 Sanna S, Sovio U, Surakka I, Teumer A, van Wingerden S, Vitart V, Zhao JH, Cavalcanti-  
657 Proenca C, Chines PS, Fisher E, Kulzer JR, Lecoeur C, Narisu N, Sandholt C, Scott LJ,  
658 Silander K, Stark K, Tammesoo ML, Teslovich TM, Timpson NJ, Watanabe RM, Welch R,  
659 Chasman DI, Cooper MN, Jansson JO, Kettunen J, Lawrence RW, Pellikka N, Perola M,

660 Vandenput L, Alavere H, Almgren P, Atwood LD, Bennett AJ, Biffar R, Bonnycastle LL,  
661 Bornstein SR, Buchanan TA, Campbell H, Day IN, Dei M, Dorr M, Elliott P, Erdos MR,  
662 Eriksson JG, Freimer NB, Fu M, Gaget S, Geus EJ, Gjesing AP, Grallert H, Grassler J,  
663 Groves CJ, Guiducci C, Hartikainen AL, Hassanali N, Havulinna AS, Herzig KH, Hicks AA,  
664 Hui J, Igl W, Jousilahti P, Jula A, Kajantie E, Kinnunen L, Kolcic I, Koskinen S, Kovacs P,  
665 Kroemer HK, Krzelj V, Kuusisto J, Kvaloy K, Laitinen J, Lantieri O, Lathrop GM, Lokki  
666 ML, Luben RN, Ludwig B, McArdle WL, McCarthy A, Morken MA, Nelis M, Neville MJ,  
667 Pare G, Parker AN, Peden JF, Pichler I, Pietilainen KH, Platou CG, Pouta A, Ridderstrale M,  
668 Samani NJ, Saramies J, Sinisalo J, Smit JH, Strawbridge RJ, Stringham HM, Swift AJ, Teder-  
669 Laving M, Thomson B, Usala G, van Meurs JB, van Ommen GJ, Vatin V, Volpato CB,  
670 Wallaschofski H, Walters GB, Widen E, Wild SH, Willemsen G, Witte DR, Zgaga L, Zitting  
671 P, Beilby JP, James AL, Kahonen M, Lehtimaki T, Nieminen MS, Ohlsson C, Palmer LJ,  
672 Raitakari O, Ridker PM, Stumvoll M, Tonjes A, Viikari J, Balkau B, Ben-Shlomo Y,  
673 Bergman RN, Boeing H, Smith GD, Ebrahim S, Froguel P, Hansen T, Hengstenberg C,  
674 Hveem K, Isomaa B, Jorgensen T, Karpe F, Khaw KT, Laakso M, Lawlor DA, Marre M,  
675 Meitinger T, Metspalu A, Midthjell K, Pedersen O, Salomaa V, Schwarz PE, Tuomi T,  
676 Tuomilehto J, Valle TT, Wareham NJ, Arnold AM, Beckmann JS, Bergmann S, Boerwinkle  
677 E, Boomsma DI, Caulfield MJ, Collins FS, Eiriksdottir G, Gudnason V, Gyllensten U,  
678 Hamsten A, Hattersley AT, Hofman A, Hu FB, Illig T, Iribarren C, Jarvelin MR, Kao WH,  
679 Kaprio J, Launer LJ, Munroe PB, Oostra B, Penninx BW, Pramstaller PP, Psaty BM,  
680 Quertermous T, Rissanen A, Rudan I, Shuldiner AR, Soranzo N, Spector TD, Syvanen AC,  
681 Uda M, Uitterlinden A, Volzke H, Vollenweider P, Wilson JF, Wittman JC, Wright AF,  
682 Abecasis GR, Boehnke M, Borecki IB, Deloukas P, Frayling TM, Groop LC, Haritunians T,  
683 Hunter DJ, Kaplan RC, North KE, O'Connell JR, Peltonen L, Schlessinger D, Strachan DP,  
684 Hirschhorn JN, Assimes TL, Wichmann HE, Thorsteinsdottir U, van Duijn CM, Stefansson

685 K, Cupples LA, Loos RJ, Barroso I, McCarthy MI, Fox CS, Mohlke KL, Lindgren CM. Meta-  
686 analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism  
687 in the genetic basis of fat distribution. *Nat Genet.* 2010;42(11):949-60.

688 28. Day N, Oakes S, Luben R, Khaw KT, Bingham S, Welch A, Wareham N. EPIC-Norfolk:  
689 study design and characteristics of the cohort. *European Prospective Investigation of Cancer.*  
690 *Br J Cancer.* 1999;80 Suppl 1:95-103.

691 29. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue  
692 gene regulation in humans. *Science.* 2015;348(6235):648-60.

693 30. Ohlsson C, Wallaschofski H, Lunetta KL, Stolk L, Perry JR, Koster A, Petersen AK,  
694 Eriksson J, Lehtimäki T, Huhtaniemi IT, Hammond GL, Maggio M, Coviello AD, Ferrucci L,  
695 Heier M, Hofman A, Holliday KL, Jansson JO, Kahonen M, Karasik D, Karlsson MK, Kiel  
696 DP, Liu Y, Ljunggren O, Lorentzon M, Lyytikäinen LP, Meitinger T, Mellström D, Melzer D,  
697 Miljkovic I, Nauck M, Nilsson M, Penninx B, Pye SR, Vasani RS, Reincke M, Rivadeneira F,  
698 Tajar A, Teumer A, Uitterlinden AG, Ulloor J, Viikari J, Volker U, Volzke H, Wichmann HE,  
699 Wu TS, Zhuang WV, Ziv E, Wu FC, Raitakari O, Eriksson A, Bidlingmaier M, Harris TB,  
700 Murray A, de Jong FH, Murabito JM, Bhasin S, Vandenput L, Haring R. Genetic  
701 determinants of serum testosterone concentrations in men. *PLoS Genet.* 2011;7(10):e1002313.

702 31. Estrada K, Styrkarsdóttir U, Evangelou E, Hsu YH, Duncan EL, Ntzani EE, Oei L,  
703 Albagha OM, Amin N, Kemp JP, Koller DL, Li G, Liu CT, Minster RL, Moayyeri A,  
704 Vandenput L, Willner D, Xiao SM, Yerges-Armstrong LM, Zheng HF, Alonso N, Eriksson J,  
705 Kammerer CM, Kaptoge SK, Leo PJ, Thorleifsson G, Wilson SG, Wilson JF, Aalto V, Alen  
706 M, Aragaki AK, Aspelund T, Center JR, Dailiana Z, Duggan DJ, Garcia M, Garcia-Giralt N,  
707 Giroux S, Hallmans G, Hocking LJ, Husted LB, Jameson KA, Khusainova R, Kim GS,  
708 Kooperberg C, Koromila T, Kruk M, Laaksonen M, Lacroix AZ, Lee SH, Leung PC, Lewis  
709 JR, Masi L, Mencej-Bedrac S, Nguyen TV, Nogueix X, Patel MS, Prezelj J, Rose LM, Scollen



710 S, Siggeirsdottir K, Smith AV, Svensson O, Trompet S, Trummer O, van Schoor NM, Woo J,  
711 Zhu K, Balcells S, Brandi ML, Buckley BM, Cheng S, Christiansen C, Cooper C, Dedoussis  
712 G, Ford I, Frost M, Goltzman D, Gonzalez-Macias J, Kahonen M, Karlsson M,  
713 Khusnutdinova E, Koh JM, Kollia P, Langdahl BL, Leslie WD, Lips P, Ljunggren O, Lorenc  
714 RS, Marc J, Mellstrom D, Obermayer-Pietsch B, Olmos JM, Pettersson-Kymmer U, Reid  
715 DM, Riancho JA, Ridker PM, Rousseau F, Slagboom PE, Tang NL, Urreizti R, Van Hul W,  
716 Viikari J, Zarrabeitia MT, Aulchenko YS, Castano-Betancourt M, Grundberg E, Herrera L,  
717 Ingvarsson T, Johannsdottir H, Kwan T, Li R, Luben R, Medina-Gomez C, Palsson ST,  
718 Reppe S, Rotter JI, Sigurdsson G, van Meurs JB, Verlaan D, Williams FM, Wood AR, Zhou  
719 Y, Gautvik KM, Pastinen T, Raychaudhuri S, Cauley JA, Chasman DI, Clark GR, Cummings  
720 SR, Danoy P, Dennison EM, Eastell R, Eisman JA, Gudnason V, Hofman A, Jackson RD,  
721 Jones G, Jukema JW, Khaw KT, Lehtimaki T, Liu Y, Lorentzon M, McCloskey E, Mitchell  
722 BD, Nandakumar K, Nicholson GC, Oostra BA, Peacock M, Pols HA, Prince RL, Raitakari  
723 O, Reid IR, Robbins J, Sambrook PN, Sham PC, Shuldiner AR, Tylavsky FA, van Duijn CM,  
724 Wareham NJ, Cupples LA, Econs MJ, Evans DM, Harris TB, Kung AW, Psaty BM, Reeve J,  
725 Spector TD, Streeten EA, Zillikens MC, Thorsteinsdottir U, Ohlsson C, Karasik D, Richards  
726 JB, Brown MA, Stefansson K, Uitterlinden AG, Ralston SH, Ioannidis JP, Kiel DP,  
727 Rivadeneira F. Genome-wide meta-analysis identifies 56 bone mineral density loci and  
728 reveals 14 loci associated with risk of fracture. *Nat Genet.* 2012;44(5):491-501.

729 32. Manning AK, Hivert MF, Scott RA, Grimsby JL, Bouatia-Naji N, Chen H, Rybin D, Liu  
730 CT, Bielak LF, Prokopenko I, Amin N, Barnes D, Cadby G, Hottenga JJ, Ingelsson E,  
731 Jackson AU, Johnson T, Kanoni S, Ladenvall C, Lagou V, Lahti J, Lecoeur C, Liu Y,  
732 Martinez-Larrad MT, Montasser ME, Navarro P, Perry JR, Rasmussen-Torvik LJ, Salo P,  
733 Sattar N, Shungin D, Strawbridge RJ, Tanaka T, van Duijn CM, An P, de Andrade M,  
734 Andrews JS, Aspelund T, Atalay M, Aulchenko Y, Balkau B, Bandinelli S, Beckmann JS,

735 Beilby JP, Bellis C, Bergman RN, Blangero J, Boban M, Boehnke M, Boerwinkle E,  
736 Bonnycastle LL, Boomsma DI, Borecki IB, Bottcher Y, Bouchard C, Brunner E, Budimir D,  
737 Campbell H, Carlson O, Chines PS, Clarke R, Collins FS, Corbaton-Anchuelo A, Couper D,  
738 de Faire U, Dedoussis GV, Deloukas P, Dimitriou M, Egan JM, Eiriksdottir G, Erdos MR,  
739 Eriksson JG, Eury E, Ferrucci L, Ford I, Forouhi NG, Fox CS, Franzosi MG, Franks PW,  
740 Frayling TM, Froguel P, Galan P, de Geus E, Gigante B, Glazer NL, Goel A, Groop L,  
741 Gudnason V, Hallmans G, Hamsten A, Hansson O, Harris TB, Hayward C, Heath S, Hercberg  
742 S, Hicks AA, Hingorani A, Hofman A, Hui J, Hung J, Jarvelin MR, Jhun MA, Johnson PC,  
743 Jukema JW, Jula A, Kao WH, Kaprio J, Kardia SL, Keinanen-Kiukaanniemi S, Kivimaki M,  
744 Kolcic I, Kovacs P, Kumari M, Kuusisto J, Kyvik KO, Laakso M, Lakka T, Lannfelt L,  
745 Lathrop GM, Launer LJ, Leander K, Li G, Lind L, Lindstrom J, Lobbens S, Loos RJ, Luan J,  
746 Lyssenko V, Magi R, Magnusson PK, Marmot M, Meneton P, Mohlke KL, Mooser V,  
747 Morken MA, Miljkovic I, Narisu N, O'Connell J, Ong KK, Oostra BA, Palmer LJ, Palotie A,  
748 Pankow JS, Peden JF, Pedersen NL, Pehlic M, Peltonen L, Penninx B, Pericic M, Perola M,  
749 Perusse L, Peyser PA, Polasek O, Pramstaller PP, Province MA, Raikonen K, Rauramaa R,  
750 Rehnberg E, Rice K, Rotter JJ, Rudan I, Ruukonen A, Saaristo T, Sabater-Lleal M, Salomaa  
751 V, Savage DB, Saxena R, Schwarz P, Seedorf U, Sennblad B, Serrano-Rios M, Shuldiner AR,  
752 Sijbrands EJ, Siscovick DS, Smit JH, Small KS, Smith NL, Smith AV, Stancakova A,  
753 Stirrups K, Stumvoll M, Sun YV, Swift AJ, Tonjes A, Tuomilehto J, Trompet S, Uitterlinden  
754 AG, Uusitupa M, Vikstrom M, Vitart V, Vohl MC, Voight BF, Vollenweider P, Waeber G,  
755 Waterworth DM, Watkins H, Wheeler E, Widen E, Wild SH, Willems SM, Willemsen G,  
756 Wilson JF, Witteman JC, Wright AF, Yaghoobkar H, Zelenika D, Zemunik T, Zgaga L,  
757 Wareham NJ, McCarthy MI, Barroso I, Watanabe RM, Florez JC, Dupuis J, Meigs JB,  
758 Langenberg C. A genome-wide approach accounting for body mass index identifies genetic

759 variants influencing fasting glycemic traits and insulin resistance. *Nat Genet.* 2012;44(6):659-  
760 69.

761 33. Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, Jackson AU, Wheeler E,  
762 Glazer NL, Bouatia-Naji N, Gloyn AL, Lindgren CM, Magi R, Morris AP, Randall J, Johnson  
763 T, Elliott P, Rybin D, Thorleifsson G, Steinthorsdottir V, Henneman P, Grallert H, Dehghan  
764 A, Hottenga JJ, Franklin CS, Navarro P, Song K, Goel A, Perry JR, Egan JM, Lajunen T,  
765 Grarup N, Sparso T, Doney A, Voight BF, Stringham HM, Li M, Kanoni S, Shrader P,  
766 Cavalcanti-Proenca C, Kumari M, Qi L, Timpson NJ, Gieger C, Zabena C, Rocheleau G,  
767 Ingelsson E, An P, O'Connell J, Luan J, Elliott A, McCarroll SA, Payne F, Roccascocca RM,  
768 Pattou F, Sethupathy P, Ardlie K, Ariyurek Y, Balkau B, Barter P, Beilby JP, Ben-Shlomo Y,  
769 Benediktsson R, Bennett AJ, Bergmann S, Bochud M, Boerwinkle E, Bonnefond A,  
770 Bonnycastle LL, Borch-Johnsen K, Bottcher Y, Brunner E, Bumpstead SJ, Charpentier G,  
771 Chen YD, Chines P, Clarke R, Coin LJ, Cooper MN, Cornelis M, Crawford G, Crisponi L,  
772 Day IN, de Geus EJ, Delplanque J, Dina C, Erdos MR, Fedson AC, Fischer-Rosinsky A,  
773 Forouhi NG, Fox CS, Frants R, Franzosi MG, Galan P, Goodarzi MO, Graessler J, Groves CJ,  
774 Grundy S, Gwilliam R, Gyllensten U, Hadjadj S, Hallmans G, Hammond N, Han X,  
775 Hartikainen AL, Hassanali N, Hayward C, Heath SC, Herberg S, Herder C, Hicks AA,  
776 Hillman DR, Hingorani AD, Hofman A, Hui J, Hung J, Isomaa B, Johnson PR, Jorgensen T,  
777 Jula A, Kaakinen M, Kaprio J, Kesaniemi YA, Kivimaki M, Knight B, Koskinen S, Kovacs P,  
778 Kyvik KO, Lathrop GM, Lawlor DA, Le Bacquer O, Lecoeur C, Li Y, Lyssenko V, Mahley  
779 R, Mangino M, Manning AK, Martinez-Larrad MT, McAteer JB, McCulloch LJ, McPherson  
780 R, Meisinger C, Melzer D, Meyre D, Mitchell BD, Morken MA, Mukherjee S, Naitza S,  
781 Narisu N, Neville MJ, Oostra BA, Orru M, Pakyz R, Palmer CN, Paolisso G, Pattaro C,  
782 Pearson D, Peden JF, Pedersen NL, Perola M, Pfeiffer AF, Pichler I, Polasek O, Posthuma D,  
783 Potter SC, Pouta A, Province MA, Psaty BM, Rathmann W, Rayner NW, Rice K, Ripatti S,

784 Rivadeneira F, Roden M, Rolandsson O, Sandbaek A, Sandhu M, Sanna S, Sayer AA, Scheet  
785 P, Scott LJ, Seedorf U, Sharp SJ, Shields B, Sigurethsson G, Sijbrands EJ, Silveira A,  
786 Simpson L, Singleton A, Smith NL, Sovio U, Swift A, Syddall H, Syvanen AC, Tanaka T,  
787 Thorand B, Tichet J, Tonjes A, Tuomi T, Uitterlinden AG, van Dijk KW, van Hoek M,  
788 Varma D, Visvikis-Siest S, Vitart V, Vogelzangs N, Waeber G, Wagner PJ, Walley A,  
789 Walters GB, Ward KL, Watkins H, Weedon MN, Wild SH, Willemsen G, Witteman JC,  
790 Yarnell JW, Zeggini E, Zelenika D, Zethelius B, Zhai G, Zhao JH, Zillikens MC, Borecki IB,  
791 Loos RJ, Meneton P, Magnusson PK, Nathan DM, Williams GH, Hattersley AT, Silander K,  
792 Salomaa V, Smith GD, Bornstein SR, Schwarz P, Spranger J, Karpe F, Shuldiner AR, Cooper  
793 C, Dedoussis GV, Serrano-Rios M, Morris AD, Lind L, Palmer LJ, Hu FB, Franks PW,  
794 Ebrahim S, Marmot M, Kao WH, Pankow JS, Sampson MJ, Kuusisto J, Laakso M, Hansen T,  
795 Pedersen O, Pramstaller PP, Wichmann HE, Illig T, Rudan I, Wright AF, Stumvoll M,  
796 Campbell H, Wilson JF, Bergman RN, Buchanan TA, Collins FS, Mohlke KL, Tuomilehto J,  
797 Valle TT, Altshuler D, Rotter JI, Siscovick DS, Penninx BW, Boomsma DI, Deloukas P,  
798 Spector TD, Frayling TM, Ferrucci L, Kong A, Thorsteinsdottir U, Stefansson K, van Duijn  
799 CM, Aulchenko YS, Cao A, Scuteri A, Schlessinger D, Uda M, Ruukonen A, Jarvelin MR,  
800 Waterworth DM, Vollenweider P, Peltonen L, Mooser V, Abecasis GR, Wareham NJ, Sladek  
801 R, Froguel P, Watanabe RM, Meigs JB, Groop L, Boehnke M, McCarthy MI, Florez JC,  
802 Barroso I. New genetic loci implicated in fasting glucose homeostasis and their impact on  
803 type 2 diabetes risk. *Nat Genet.* 2010;42(2):105-16.

804 34. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with  
805 multiple genetic variants using summarized data. *Genet Epidemiol.* 2013;37(7):658-65.

806 35. Ahn J, Schumacher FR, Berndt SI, Pfeiffer R, Albanes D, Andriole GL, Ardanaz E,  
807 Boeing H, Bueno-de-Mesquita B, Chanock SJ, Clavel-Chapelon F, Diver WR, Feigelson HS,  
808 Gaziano JM, Giovannucci E, Haiman CA, Henderson BE, Hoover RN, Kolonel LN, Kraft P,

809 Ma J, Le Marchand L, Overvad K, Palli D, Stattin P, Stampfer M, Stram DO, Thomas G,  
810 Thun MJ, Travis RC, Trichopoulos D, Virtamo J, Weinstein SJ, Yeager M, Kaaks R, Hunter  
811 DJ, Hayes RB. Quantitative trait loci predicting circulating sex steroid hormones in men from  
812 the NCI-Breast and Prostate Cancer Cohort Consortium (BPC3). *Hum Mol Genet.*  
813 2009;18(19):3749-57.

814 36. Bulun SE, Sebastian S, Takayama K, Suzuki T, Sasano H, Shozu M. The human CYP19  
815 (aromatase P450) gene: update on physiologic roles and genomic organization of promoters. *J*  
816 *Steroid Biochem Mol Biol.* 2003;86(3-5):219-24.

817 37. Haiman CA, Stram DO, Pike MC, Kolonel LN, Burt NP, Altshuler D, Hirschhorn J,  
818 Henderson BE. A comprehensive haplotype analysis of CYP19 and breast cancer risk: the  
819 Multiethnic Cohort. *Hum Mol Genet.* 2003;12(20):2679-92.

820 38. Travis RC, Schumacher F, Hirschhorn JN, Kraft P, Allen NE, Albanes D, Berglund G,  
821 Berndt SI, Boeing H, Bueno-de-Mesquita HB, Calle EE, Chanock S, Dunning AM, Hayes R,  
822 Feigelson HS, Gaziano JM, Giovannucci E, Haiman CA, Henderson BE, Kaaks R, Kolonel  
823 LN, Ma J, Rodriguez L, Riboli E, Stampfer M, Stram DO, Thun MJ, Tjonneland A,  
824 Trichopoulos D, Vineis P, Virtamo J, Le Marchand L, Hunter DJ. CYP19A1 genetic variation  
825 in relation to prostate cancer risk and circulating sex hormone concentrations in men from the  
826 Breast and Prostate Cancer Cohort Consortium. *Cancer Epidemiol Biomarkers Prev.*  
827 2009;18(10):2734-44.

828 39. Huhtaniemi IT, Pye SR, Holliday KL, Thomson W, O'Neill TW, Platt H, Payne D, John  
829 SL, Jiang M, Bartfai G, Boonen S, Casanueva FF, Finn JD, Forti G, Giwercman A, Han TS,  
830 Kula K, Lean ME, Pendleton N, Punab M, Silman AJ, Vanderschueren D, Labrie F, Wu FC.  
831 Effect of polymorphisms in selected genes involved in pituitary-testicular function on  
832 reproductive hormones and phenotype in aging men. *J Clin Endocrinol Metab.*  
833 2010;95(4):1898-908.

834 40. Haiman CA, Dossus L, Setiawan VW, Stram DO, Dunning AM, Thomas G, Thun MJ,  
835 Albanes D, Altshuler D, Ardanaz E, Boeing H, Buring J, Burt N, Calle EE, Chanock S,  
836 Clavel-Chapelon F, Colditz GA, Cox DG, Feigelson HS, Hankinson SE, Hayes RB,  
837 Henderson BE, Hirschhorn JN, Hoover R, Hunter DJ, Kaaks R, Kolonel LN, Le Marchand L,  
838 Lenner P, Lund E, Panico S, Peeters PH, Pike MC, Riboli E, Tjonneland A, Travis R,  
839 Trichopoulos D, Wacholder S, Ziegler RG. Genetic variation at the CYP19A1 locus predicts  
840 circulating estrogen levels but not breast cancer risk in postmenopausal women. *Cancer Res.*  
841 2007;67(5):1893-7.

842 41. Jin G, Sun J, Kim ST, Feng J, Wang Z, Tao S, Chen Z, Purcell L, Smith S, Isaacs WB,  
843 Rittmaster RS, Zheng SL, Condreay LD, Xu J. Genome-wide association study identifies a  
844 new locus JMJD1C at 10q21 that may influence serum androgen levels in men. *Hum Mol*  
845 *Genet.* 2012;21(23):5222-8.

846 42. Martinez-Garay I, Jablonka S, Sutajova M, Steuernagel P, Gal A, Kutsche K. A new gene  
847 family (FAM9) of low-copy repeats in Xp22.3 expressed exclusively in testis: implications  
848 for recombinations in this region. *Genomics.* 2002;80(3):259-67.

849 43. Oliveira LM, Seminara SB, Beranova M, Hayes FJ, Valkenburgh SB, Schipani E, Costa  
850 EM, Latronico AC, Crowley WF, Jr., Vallejo M. The importance of autosomal genes in  
851 Kallmann syndrome: genotype-phenotype correlations and neuroendocrine characteristics. *J*  
852 *Clin Endocrinol Metab.* 2001;86(4):1532-8.

853 44. Day FR, Bulik-Sullivan B, Hinds DA, Finucane HK, Murabito JM, Tung JY, Ong KK,  
854 Perry JR. Shared genetic aetiology of puberty timing between sexes and with health-related  
855 outcomes. *Nat Commun.* 2015;6:8842.

856 45. Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, Pfeffer S, Rice A,  
857 Kamphorst AO, Landthaler M, Lin C, Socci ND, Hermida L, Fulci V, Chiaretti S, Foa R,  
858 Schliwka J, Fuchs U, Novosel A, Muller RU, Schermer B, Bissels U, Inman J, Phan Q, Chien

859 M, Weir DB, Choksi R, De Vita G, Frezzetti D, Trompeter HI, Hornung V, Teng G,  
860 Hartmann G, Palkovits M, Di Lauro R, Wernet P, Macino G, Rogler CE, Nagle JW, Ju J,  
861 Papavasiliou FN, Benzing T, Lichter P, Tam W, Brownstein MJ, Bosio A, Borkhardt A,  
862 Russo JJ, Sander C, Zavolan M, Tuschl T. A mammalian microRNA expression atlas based  
863 on small RNA library sequencing. *Cell*. 2007;129(7):1401-14.

864 46. Pu D, Xing Y, Gao Y, Gu L, Wu J. Gene variation and premature ovarian failure: a meta-  
865 analysis. *Eur J Obstet Gynecol Reprod Biol*. 2014;182C:226-37.

866 47. Zhai G, Teumer A, Stolk L, Perry JR, Vandenput L, Coviello AD, Koster A, Bell JT,  
867 Bhasin S, Eriksson J, Eriksson A, Ernst F, Ferrucci L, Frayling TM, Glass D, Grundberg E,  
868 Haring R, Hedman AK, Hofman A, Kiel DP, Kroemer HK, Liu Y, Lunetta KL, Maggio M,  
869 Lorentzon M, Mangino M, Melzer D, Miljkovic I, Nica A, Penninx BW, Vasani RS,  
870 Rivadeneira F, Small KS, Soranzo N, Uitterlinden AG, Volzke H, Wilson SG, Xi L, Zhuang  
871 WV, Harris TB, Murabito JM, Ohlsson C, Murray A, de Jong FH, Spector TD, Wallaschowski  
872 H. Eight common genetic variants associated with serum DHEAS levels suggest a key role in  
873 ageing mechanisms. *PLoS Genet*. 2011;7(4):e1002025.

874 48. Tivesten A, Vandenput L, Carlzon D, Nilsson M, Karlsson MK, Ljunggren O, Barrett-  
875 Connor E, Mellstrom D, Ohlsson C. Dehydroepiandrosterone and its sulfate predict the 5-year  
876 risk of coronary heart disease events in elderly men. *J Am Coll Cardiol*. 2014;64(17):1801-10.

877 49. Yan J, Li Q, Mao AP, Hu MM, Shu HB. TRIM4 modulates type I interferon induction  
878 and cellular antiviral response by targeting RIG-I for K63-linked ubiquitination. *J Mol Cell*  
879 *Biol*. 2014;6(2):154-63.

880 50. Krone N, Arlt W. Genetics of congenital adrenal hyperplasia. *Best Pract Res Clin*  
881 *Endocrinol Metab*. 2009;23(2):181-92.

882 51. Johnell O, Kanis JA, Oden A, Johansson H, De Laet C, Delmas P, Eisman JA, Fujiwara S,  
883 Kroger H, Mellstrom D, Meunier PJ, Melton LJ, 3rd, O'Neill T, Pols H, Reeve J, Silman A,  
884 Tenenhouse A. Predictive value of BMD for hip and other fractures. *J Bone Miner Res.*

885



886 **Legends to figures**

887 **Figure 1A-C. Manhattan plots for the genome-wide meta-analysis results.**

888 (A) E2 adjusted for age and BMI, (B) E2 adjusted for, age, BMI, testosterone and SHBG and  
889 (C) E1 adjusted for age and BMI. Red line indicates  $p = 5 \times 10^{-8}$ . Genome-wide significant  
890 loci are indicated by green color. In the analysis of E1, one SNP on chromosome 1 reached  
891 the threshold for genome-wide significance ( $p < 5 \times 10^{-8}$ ), but had a minor allele frequency of  
892  $< 0.01$  in all but two cohorts. Therefore this SNP was discarded from further analyses.

893 **Figure 2. Proposed mechanisms underlying the associations between genome wide**  
894 **significant SNPs and serum levels of E2 and T**

895 SNPs associated with elevated levels of both E2 and T are expected to be located upstream of  
896 T. SNPs associated with elevated levels of E2 but no increase in T levels are expected to be  
897 affecting aromatase activity or estradiol clearance. The allele associated with increased serum  
898 E2 is given for each SNP. The proposed effect of E2 on BMD is also indicated; upwards  
899 arrow represents increase, downwards arrow represents decrease and arrow in parentheses  
900 represents non-significant decrease.

901 **Figure 3. Forest plot of Mendelian Randomization analyses showing the effect of E2 on**  
902 **BMD.**

903 Effect size of E2 on BMD expressed as SD increase in BMD per pg/ml E2. The horizontal  
904 lines represent confidence interval; the central vertical line represents precision. The values  
905 are based on a meta-analysis of all five E2 associated SNPs (rs727479, rs2899472,  
906 rs16964258, rs5934505, rs5951794). The horizontal axis shows the scale of the effects.  
907 LSBMD = lumbar spine BMD, FNBMD = femoral neck BMD.

908

