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Review

Spinal interneuronal networks in the cat: Elementary components

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ABSTRACT

This review summarises features of networks of commissural interneurons co-ordinating muscle activity on both sides of the body as an example of feline elementary spinal interneuronal networks. The main feature of these elementary networks is that they are interconnected and incorporated into more complex networks as their building blocks. Links between networks of commissural interneurons and other networks are quite direct, with mono- and disynaptic input from the reticulospinal and vestibulospinal neurones, disynaptic from the contralateral and ipsilateral corticospinal neurones and fastigial neurones, di- or oligosynaptic from the mesencephalic locomotor region and mono-, di- or oligosynaptic from muscle afferents. The most direct links between commissural interneurons and motoneurons are likewise simple: monosynaptic and disynaptic via premotor interneurons with input from muscle afferents. By such connections, a particular elementary interneuronal network may subserve a wide range of movements, from simple reflex and postural adjustments to complex centrally initiated phasic and rhythmic movements, including voluntary movements and locomotion. Other common features of the commissural and other interneuronal networks investigated so far is that input from several sources is distributed to their constituent neurones in a semi-random fashion and that there are several possibilities of interactions between neurones both within and between various populations. Neurones of a particular elementary network are located at well-defined sites but intermixed with neurones of other networks and distributed over considerable lengths of the spinal cord, which precludes the topography to be used as their distinguishing feature.

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Abbreviations: 4-AP, 4-aminopyridine; 5-HT, 5-hydroxytryptamine (serotonin); c, commissural; co, contralateral; EPSP, excitatory postsynaptic potential; FN, fastigial nucleus; GABA, gamma aminobutyric acid; GS, gastrocnemius–soleus; i, ipsilateral; IPSP, inhibitory postsynaptic potential; L, lumbar; LVN, lateral vestibular nucleus; MLF, medial longitudinal fascicle; MLR, mesencephalic locomotor region; MN, motoneuron; NA, noradrenaline; Q, quadriceps nerve; PT, pyramidal tract; RS, reticulospinal; T, threshold; VS, vestibulospinal

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1. Introduction

Elementary spinal interneuronal networks are very simple. In the simplest cases, there are just one or two interneurons in series between input neurones and motoneurons. However, even in the simplest networks, there is a number of interneurons of each kind in parallel and these neurones integrate somewhat different combinations of information, from not only their main sources of input, e.g. muscle and skin afferents, but also from other neuronal networks. They forward it also to somewhat different combinations of their target neurones, including interneurons of other neuronal networks. Because of their links with other networks, all elementary networks may thus be considered to be components of more complex networks.

This arrangement may be illustrated with any of the previously investigated networks of spinal interneurons, from Renshaw cells and interneurons mediating Ia reciprocal inhibition, which were among the first interneurons to be analysed (for references, see Jankowska, 1992), through cervical propriospinal neurones (Lundberg, 1979) and interneurons mediating reflex actions of group II muscle spindle afferents (Jankowska et al., 2002a,b), to mention only those known in most detail. In this review it will be illustrated with the recently investigated networks of commissural interneurons. These networks have become of particular interest as being attributed a critical role in locomotor networks (for references, see Buchanan, 1999; Grillner, 2003; Kiehn, 2006; Soffe et al., 1984) because they are needed to adjust rhythmic activity of neurones on both sides of the spinal cord and because they are one of the major targets of reticulospinal neurones that are involved in initiation of locomotion. There is also a growing body of evidence that commissural interneurons may be of critical importance for other centrally or reflexly initiated phasic movements, including voluntary movements and postural adjustments, and that individual commissural interneurons may contribute to several of these movements.

2. Networks of commissural interneurons as examples of spinal elementary networks

2.1. Functional differentiation of the population of commissural interneurons

As other spinal interneuronal populations, the population of commissural interneurons is not homogenous. It includes

subpopulations of both excitatory (glutamatergic) and inhibitory (glycinergic) neurones (Bannatyne et al., 2003, 2006; Butt and Kiehn, 2003; Nissen et al., 2005; Roberts et al., 1988; Sugiuchi et al., 1995), at different locations (Bannatyne et al., 2003, 2006; Harrison et al., 1986; Huang et al., 2000; Kiehn and Butt, 2003; Lu et al., 2001; Ohta et al., 1991; Stokke et al., 2002), with different target cells (Bannatyne et al., 2003, 2006; Birinyi et al., 2003; Butt et al., 2002; Butt and Kiehn, 2003; Matsuyama et al., 2006, 2004a,b; Stokke et al., 2002) and with different types of input (Harrison et al., 1986; Jankowska et al., 2005a,b,c; Jankowska and Noga, 1990). For instance, commissural interneurons of the L3–L6 segments that target contralateral motoneurons in caudal lumbar segments fall into two main subpopulations, those with monosynaptic input from reticulospinal (RS) neurones, vestibulospinal (VS) neurones and group I afferents, and those with monosynaptic input from group II muscle afferents (Jankowska et al., 2005a,b,c).

In the adult cat, rat and mouse, the majority of commissural interneurons are located in lamina VIII on one side of the grey matter (Harrison et al., 1986; Hoover and Durkovic, 1992; Puskar and Antal, 1997; Stokke et al., 2002) and target neurones on the other side (Bannatyne et al., 2003; Matsuyama et al., 2006, 2004a, b; Nissen et al., 2005), as illustrated in Figs. 1A,B. This is true for both excitatory and inhibitory lamina VIII commissural interneurons but occasional bilateral projections have recently been reported (in one out of 34 lamina VIII neurones analysed by Matsuyama (2006). Bilateral projections have also been found in the case of two groups of interneurons with input from group II afferents: inhibitory (but not excitatory) dorsal horn interneurons (Bannatyne et al., 2006; Figs. 1C and D) and excitatory (but not inhibitory) lamina VII interneurons (B.A. Bannatyne, D.J. Maxwell K. Stecina, I. Hammar and E. Jankowska, unpublished). Contralateral projections have also been demonstrated for unidentified, primarily inhibitory dorsal horn neurones in the adult rat (Petko and Antal, 2000; Petko et al., 2004), but without specifying whether the same neurones projected ipsilaterally.

In contrast to projections in adult animals, projections of lamina VIII interneurons in neonatal animals appear to be more often bilateral, at least as judged by anatomical studies using the Golgi technique (Cajal, 1953; Scheibel and Scheibel, 1966), with examples in Figs. 1E and F. This may indicate that ipsilateral axon collaterals of these interneurons withdraw at some stage during the development. Bilateral projections may also be more frequent in genetically modified EphA4 knockout mice exhibiting synchronous (rabbit- or kangaroo-like) rather than alternating gait (Dottori et al., 1998; Kullander et al., 2003).

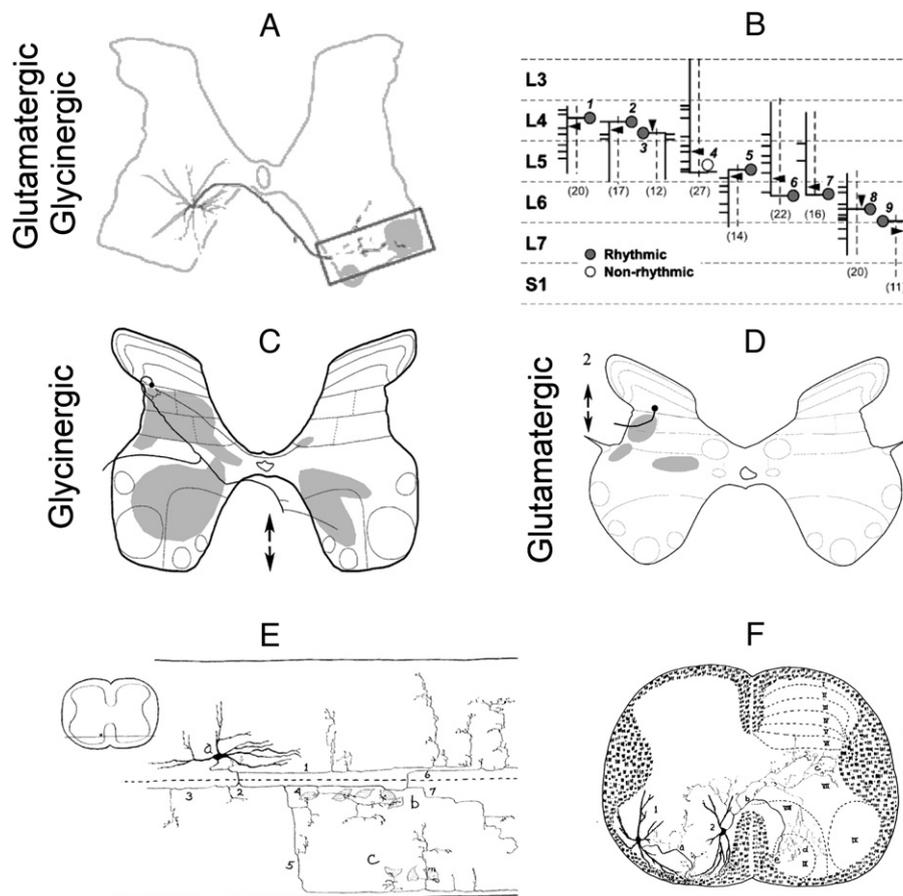


Fig. 1 – Axonal projections of commissural interneurons. (A) An example of exclusively contralateral projections of lamina VIII neurobiotin-labelled interneurons with input from reticulospinal neurons in adult cat, with the trajectory of the main axonal branches, as indicated, and terminal projection areas shaded (modified from Fig. 9 in [Bannatyne et al., 2003](#)). **(B)** Summary of axonal branching of intra-axonally labelled adult laminae VIII interneurons with input from reticulospinal neurons; all collaterals given of contralaterally (to the left of the dashed lines indicating the midline (modified from Fig. 6 in [Matsuyama et al., 2004b](#)). **(C, D)** Bilateral projections of inhibitory dorsal horn interneurons with input from group II afferents but only ipsilateral of excitatory ones (modified from Figs. 7 and 8 in [Bannatyne et al., 2006](#)). Shaded areas in panels C and D indicate the total terminal projected areas of the reconstructed axonal branches. **(E, F)** Bilateral projections of lamina VIII commissural interneurons in a newborn kitten and mouse (Golgi staining; modified from Figs. 5 and 8 in [Scheibel and Scheibel, 1966](#)).

2.2. Network connections indicated by patterns of input to lamina VIII commissural interneurons targeted by reticulospinal neurones

Network connections of commissural interneurons have been found to be generally very extensive but differ for various subpopulations of these neurones. [Fig. 2](#) summarizes some of the connections for interneurons with monosynaptic input from RS and VS neurones. The figure shows that commissural interneurons monosynaptically excited by RS neurones (A) ([Jankowska et al., 2005a,b,c, 2003](#)) may be disynaptically excited by both ipsilateral (B) and contralateral (C) pyramidal tract (PT) neurones ([Cabaj et al., 2006; Edgley et al., 2004; Jankowska et al., 2006](#)), and thus, incorporated in networks of neurones initiating voluntary movements. Rhythmic activation of many of these neurones by stimuli applied in the mesencephalic locomotor region (MLR) (F) ([Matsuyama et al., 2004a,b](#)) shows that they are also incorporated in networks of locomotion.

Commissural interneurons monosynaptically excited by VS neurones (E) ([Krutki et al., 2003](#)) will contribute to postural adjustments evoked by signals from the vestibular receptors and neck afferents and, as they are disynaptically excited by fastigial neurones (FN) (F) ([Matsuyama and Jankowska, 2004](#)), they are also incorporated in the networks of motor control and learning that depend on the cerebellum.

[Fig. 2](#) shows only the most direct connections, monosynaptic and disynaptic, but there are also possibilities of an additional indirect coupling at any of the linking sites and of incorporation of the network of these neurones into even more complex neuronal systems, e.g. of locomotion steered by cortical neurones ([Matsuyama et al., 2004a,b](#)) or scratching initiated by contralateral afferents ([Stein et al., 1995](#)).

Synaptic actions evoked by only one source of input to commissural interneurons were generally weak. However, in several cases, the neurones responded not only with small intracellularly recorded EPSPs but also with extracellularly

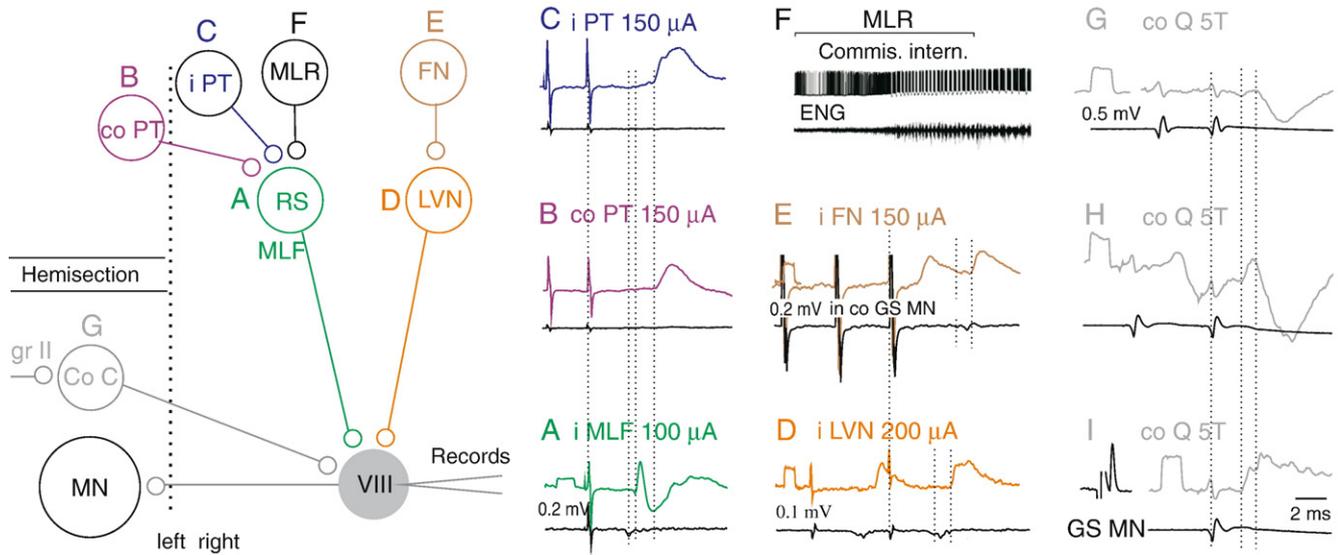


Fig. 2 – Incorporation of commissural interneurons with monosynaptic input from reticulospinal neurons into other spinal networks. The grey circle represents commissural interneurons targeting contralateral motoneurons. Other circles represent commissural interneurons (Co C) on the opposite side of the spinal grey matter, reticulospinal (RS) neurons with axons in the ipsilateral medial longitudinal fascicle (MLF), neurons in the lateral vestibular nucleus (LVN), ipsilateral and contralateral pyramidal tract (PT) neurons, and neurons in the ipsilateral mesencephalic locomotor region (MLR) and cerebellar fastigial nucleus (FN). Records A–D and G–I are from commissural interneurons, which were antidromically activated from the contralateral GS motor nuclei, while those in panel E are from a GS motoneurone and in panel F from an unspecified commissural neurone. They show PSPs (or action potentials in panel F) evoked by stimulation of the indicated structures. Dotted lines in A–E indicate: stimulus artefacts, descending volleys following the MLF or VS stimuli, onset of monosynaptic EPSPs and onset of di- or trisynaptically evoked EPSPs. Dotted lines in panels G–I indicate afferent volleys from the Q nerve and onset of disynaptic EPSPs and IPSPs evoked by them. The records are: A–C from Fig. 6 in Jankowska et al. (2006); D from Fig. 1 in Krutki et al. (2003); E from Fig. 5 in Matsuyama and Jankowska (2004); F from Fig. 8 in Matsuyama et al. (2004b); G–I from Figs. 5 and 7 in Jankowska et al. (2005a,b,c).

recorded action potentials, showing that these weak synaptic actions may nonetheless be suprathreshold. Furthermore, activation of commissural interneurons was considerably facilitated when 2 sources of input were jointly activated (e.g. MLF and VS, or both ipsilateral and contralateral PT neurons; Edgley et al., 2004; Jankowska et al., 2005a,b,c; Krutki et al., 2003). Synaptic transmission in their networks could also be enhanced, e.g. by the K^+ channel blocker 4-AP (Jankowska et al., 2005a,b,c) and by monoamines (Hammar et al., 2004), as illustrated in Fig. 3.

Strengthening and prolonging activation of commissural interneurons by delayed actions of some sources of input to them might be particularly important for triggering voltage-dependent persistent inward current and plateau potentials in motoneurons (Houngaard et al., 1988; Hultborn, 1999; Hultborn et al., 2003; Schwindt and Crill, 1980), which in turn give rise to long-lasting discharges. This is because the critical time of depolarization needed for the persistent inward current in motoneurons apparently exceeds 10 ms for even the strongest sources of the depolarization and over 100 ms for the weaker ones (Hultborn et al., 2003; Lee and Heckman, 1996; Schwindt and Crill, 1980). Doubling or tripling of the period of activation of commissural interneurons, and hence, of their actions on motoneurons, by utilising several parallel pathways with different overall conduction time might thus be beneficial for this purpose.

2.3. Network connections indicated by output from lamina VIII commissural interneurons activated by reticulospinal neurons

Reconstruction of terminal branching of intracellularly labelled lamina VIII interneurons projecting to contralateral hindlimb motor nuclei revealed that their projection areas are not only in lamina IX but extend to laminae VI–VIII (Bannatyne et al., 2003,2006; Matsuyama et al., 2006, 2004a,b; see also Birinyi et al., 2003), and overlap with the areas of location of several populations of premotor interneurons (for reference, see Jankowska, 1992).

At locations indicated in Fig. 4E, commissural interneurons might target interneurons mediating nonreciprocal inhibition from group I muscle afferents and interneurons in pathways from group II afferents. Intracellular records from interneurons in this area have shown that stimuli applied in the MLF did indeed evoke disynaptic EPSPs (Figs. 5A and D) or IPSPs (Figs. 5B and E) in interneurons with input from group Ib or II afferents (Cabaj et al., 2006). These EPSPs and IPSPs were evoked at the same latencies as in motoneurons (Figs. 5G and H), indicating that both excitatory and inhibitory lamina VIII commissural neurons act in parallel on motoneurons and on these interneurons. Interneurons excited by commissural interneurons might thus add to their actions on motoneurons with an

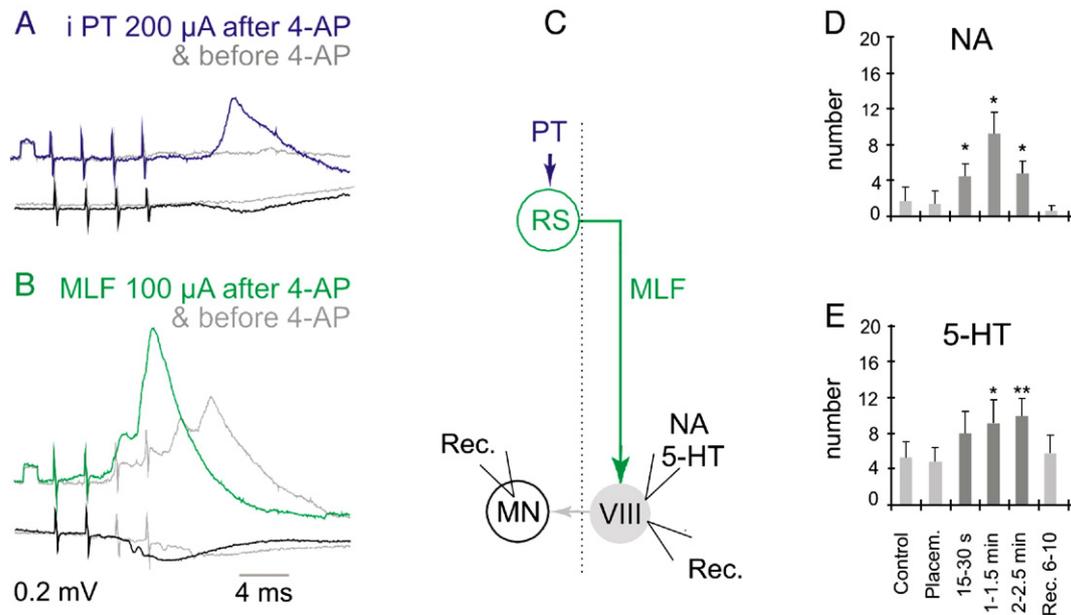


Fig. 3 – Enhancement of synaptic actions in the double crossed pathways between ipsilateral PT neurones and motoneurons. (A, B) Enhancement of EPSPs evoked by pyramidal tract (PT) and medial longitudinal fascicle (MLF) stimuli in a GS motoneurone after systemic application of the K^+ channel blocker 4-aminopyridine (4-AP) (modified from Fig. 2 in Jankowska et al., 2005a,b,c). (C) Diagram of connections in these pathways, via reticulospinal (RS) neurones and lamina VIII commissural interneurons, and sites of recording from a motoneurone (MN) in panels A and B and of recording from interneurons and of ionophoresis in panels D and E. (D, E) Histograms of mean numbers of spike potentials evoked in commissural interneurons by 20 stimuli applied within the MLF before (light grey; control and placement), during (dark grey) and after (light grey; recovery) ionophoresis of noradrenaline (NA) and serotonin (5-HT) (modified from Fig. 4 in Hammar et al., 2004).

additional synaptic delay, as indicated in Fig. 5I. Excitatory premotor interneurons would amplify excitatory actions of commissural interneurons, whereas inhibitory interneurons might either amplify inhibitory actions or counteract the excitatory ones. The illustrated records are from interneurons with characteristics of premotor interneurons but which have not been identified as exciting or inhibiting motoneurons. However, evidence that both excitatory and inhibitory premotor interneurons mediate actions of commissural interneurons has been provided by the demonstration that disynaptic EPSPs and/or IPSPs evoked by group Ia, Ib or II afferents in motoneurons are enhanced by a preceding stimulation of neuronal systems that activate commissural interneurons (Cabaj et al., 2006). This is illustrated in Fig. 5J–L with enhancement of disynaptic IPSPs evoked from Ib afferents.

As a consequence of their actions on premotor interneurons, commissural interneurons might modify the operation of the whole networks of these premotor interneurons; they might not only enhance PSPs evoked by them in motoneurons but also the degree of mutual inhibitory interactions between Ib interneurons (Brink et al., 1983) or between Ib interneurons and group II interneurons (Edgley and Jankowska, 1987).

At locations indicated in Fig. 4B, commissural interneurons might target contralateral commissural interneurons and interneurons mediating Ia reciprocal inhibition. Their actions on such interneurons were demonstrated by disynaptic or trisynaptic actions of contralateral afferents on commissural interneurons (Harrison et al., 1986) and by enhancement of disynaptic IPSPs evoked in motoneurons by Ia afferents and of

disynaptic or trisynaptic EPSPs and IPSPs evoked from contralateral group II afferents (via Ia inhibitory interneurons and commissural interneurons, respectively; Jankowska et al., 2005a,b,c). The Ia interneurons were, in addition, shown to be the last order interneurons of IPSPs evoked in motoneurons via contralateral commissural interneurons activated from the MLF and VS as well as from both PTs. The evidence was depression of these IPSPs by a preceding activation of Renshaw cells (Figs. 6B–D), which counteract actions of Ia inhibitory interneurons (Hultborn et al., 1971) but do not depress direct actions of commissural interneurons (Fig. 6A).

By exciting Ia inhibitory interneurons, commissural interneurons would also influence operation of their whole networks; enhancing actions of Ia interneurons on interneurons that mediate inhibition of agonists and counteracting the depression of their actions by Renshaw cells (Fig. 6 right diagram; Hultborn et al., 1976a,b). As Ia inhibitory interneurons are last-order interneurons of several contralateral and ipsilateral neuronal networks (see Lindstrom, 1973), the network of commissural interneurons would work in concert with them, e.g. during postural adjustments evoked by signals from the vestibular receptors (by ipsilateral VS neurones; Hultborn and Udo, 1972), and voluntary movements (by PT neurones; Hultborn et al., 1976a,b; Hultborn and Udo, 1972) or fictive locomotion (Degtyarenko et al., 1998; Feldman and Orlovsky, 1975) or scratching (Deliagina and Orlovsky, 1980).

At locations indicated in Fig. 4B, commissural interneurons might also target contralateral Renshaw cells, as postulated by

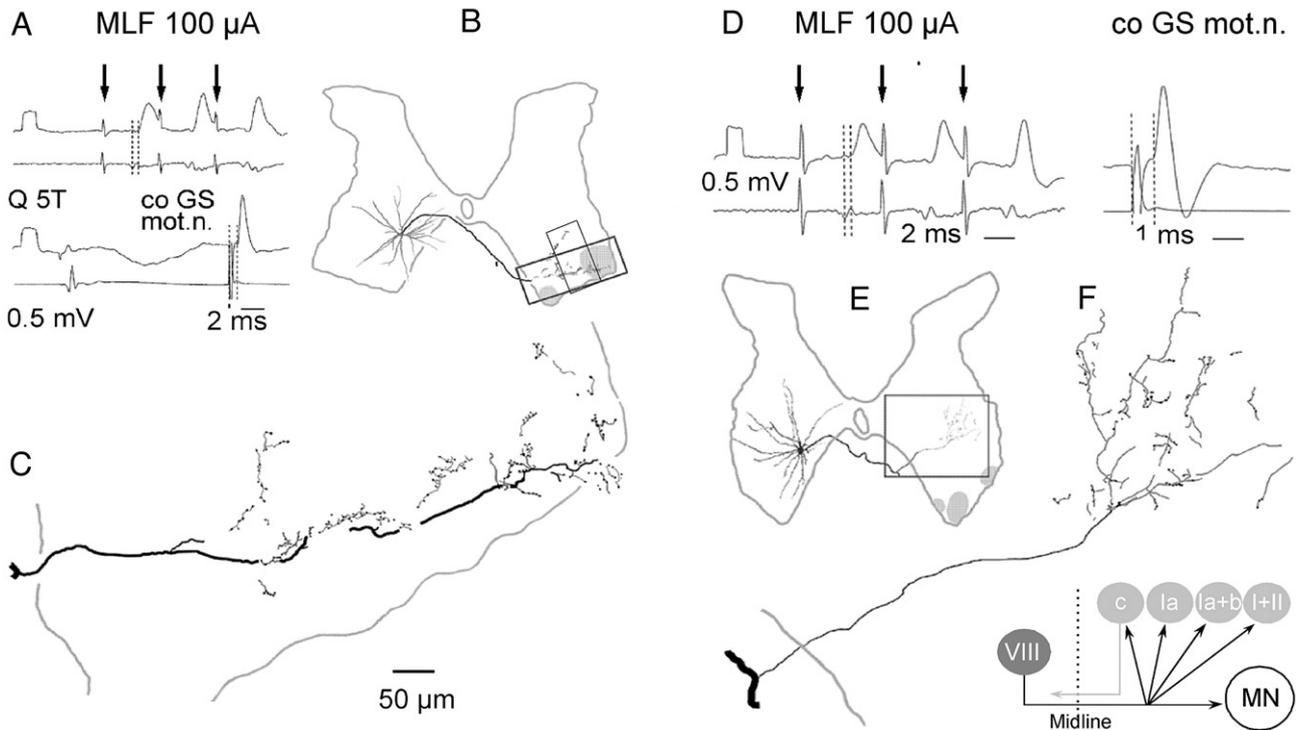


Fig. 4 – Terminal projection areas of lamina VIII commissural interneurons with input from RS neurones outside motor nuclei. (A, D) Records from two excitatory interneurons with monosynaptic EPSPs from the MLF (<1 ms latency from the descending volleys indicated by the first dotted lines). Both were antidromically activated from the gastrocnemius–soleus (GS) motor nuclei, but only the first was found to project to motor nuclei in the L5 segment (B) and their projections outside motor nuclei differed. In the L5 segment (B) they were in the ventral horn dorsal and medial to motor nuclei (upper box) and in the L4 segment (E) in the more dorsal part of lamina VII. In both areas, a considerable number of terminals were found (C and F) (modified from Figs. 7 and 9 in Bannatyne et al., 2003). Diagram shows hypothetical target cells (light grey) of lamina VIII commissural interneurons (dark grey) outside motor nuclei; dotted line indicates midline.

Nishimaru et al. (2006). However, as stated by the authors, more studies would be needed before relations between commissural interneurons and Renshaw cells are established.

2.4. Network connections indicated by input to and output from commissural interneurons activated by group II muscle afferents

Only a small proportion of commissural interneurons with monosynaptic input from ipsilateral group II muscle afferents were found to be excited by reticulospinal neurones, and if so, only di- or polysynaptically. The network of these neurones appears therefore to be only to a small extent, and only indirectly, incorporated in the networks steered by descending tract neurones illustrated in Fig. 2. This is also indicated by differences in modulatory actions of monoamines (Hammar et al., 2004) and of presynaptically acting GABAergic neurones (Edgley et al., 2003; Jankowska et al., 2002a,b) on commissural interneurons with monosynaptic input from group II afferents and from the MLF.

Activity of subpopulations of commissural interneurons with monosynaptic input from ipsilateral group II muscle afferents on both sides of the spinal cord might nevertheless be linked. This is indicated by disynaptic EPSPs and IPSPs from contralateral group II afferents in commissural interneurons

with monosynaptic input from the ipsilateral MLF (illustrated in Fig. 2G) or from the contralateral MLF in commissural interneurons with monosynaptic input from group II afferents (Jankowska et al., 2005a,b,c), which implicate monosynaptic actions of contralaterally located commissural interneurons which mediate them. These observations extend the original evidence for mutual interactions between networks of commissural interneurons on both sides of the body based on demonstration of disynaptic EPSPs and IPSPs from contralateral group I afferents (Harrison et al., 1986) and are substantiated by morphological demonstration of synaptic contacts between commissural interneurons on both sides of the spinal cord (Birinyi et al., 2003; Matsuyama et al., 2006), even if the interconnected neurones have not been specified.

The subpopulations of commissural interneurons with input from the MLF and from group II muscle afferents likewise appear to target the same populations of premotor interneurons, those mediating actions of ipsilateral group Ia, Ib and II muscle afferents on motoneurons. This is indicated by an effective facilitation of IPSPs evoked by Ia afferents (Bruggencate et al., 1969) and of EPSPs or IPSPs evoked by group Ib and II afferents (Cabaj et al., 2006) by conditioning stimulation of contralateral group II afferents as of the MLF. Disynaptic EPSPs recorded in premotor interneurons with group II input from

contralateral group II afferents fully supported this conclusion (Arya et al., 1991; Bajwa et al., 1992).

3. Comparison of internal organization of elementary interneuronal networks

One of the common features of the so far analysed elementary interneuronal networks is that input to each interneuronal population is drawn from a number of sources, and that input from any of these sources is distributed to several populations, although in different combinations, e.g. both Ia and Ib afferents provide input to interneurons mediating non-reciprocal inhibition of motoneurons (Jankowska et al., 1981), but Ia afferents are the main source of peripheral input to interneurons mediating

reciprocal inhibition (Hultborn, 1972), while Ib afferents excite also interneurons in pathways from group II afferents. In a given population, input from different sources appears to be distributed, with practically at random connections between afferents, or neurones that provide it, and individual interneurons of this population (Edgley, 2001; Harrison and Jankowska, 1985; Harrison et al., 1986; Hultborn, 1972; Jankowska et al., 2005a,b,c).

Another common feature is that of internal control of operation of neurones within the network. Mutual inhibitory interactions have been found between Renshaw cells (Ryall, 1970), Ia inhibitory interneurons (Hultborn et al., 1976a,b), Ib interneurons (Brink et al., 1983), group II interneurons (Edgley and Jankowska, 1987) and commissural interneurons (Harrison et al., 1986; Jankowska et al., 2005a,b,c). In several cases, they were found between interneurons with the same input. e.g. from

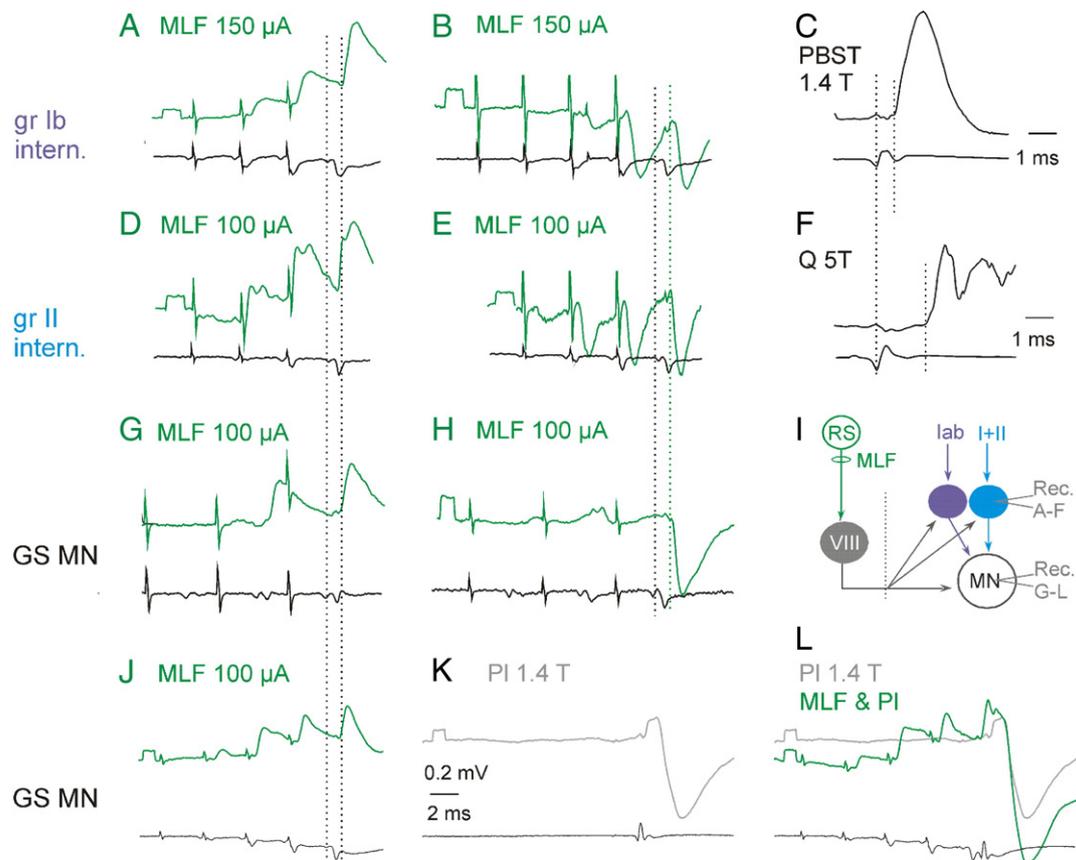


Fig. 5 – Examples of disynaptic PSPs evoked from the MLF via commissural interneurons—at the same latency in motoneurons and in interneurons in pathways from group Ib and II afferents, and thus, indicating collateral actions of commissural interneurons on other premotor interneurons. (A–F) Intracellular records (upper traces) from four interneurons with input from group Ib afferents (illustrated in panel C) or group II afferents (illustrated in panel F) in which disynaptic EPSPs (left panels) or IPSPs (middle panels) were evoked from the MLF. As judged by antidromic activation from motor nuclei, such interneurons were premotor interneurons in pathways from group Ib or II afferents. (G, H) Records from two GS motoneurons aligned with respect to MLF volleys evoked by the third stimulus; the volleys are indicated by the first dotted lines and the onset of the PSPs by the second. (I) Diagram showing collateral connections between MLF and the two kinds of interneurons illustrated in panels A–F. The circles represent subpopulations of interneurons of the various populations but the indicated connections apply to individual interneurons of these subpopulations. (J–L) Evidence that disynaptic Ib IPSPs evoked in motoneurons (K) are mediated by interneurons co-excited from the MLF because they are facilitated (L) when preceded by MLF stimuli. Note much greater amplitude of IPSPs evoked when stimulation of Ib afferents was preceded by stimulation of the MLF (black traces in panel L) than when it was not (K and grey traces in panel L). Such records substantiate connections indicated in panel I and postulated on the basis of the data illustrated in Fig. 4 (modified from Figs. 3, 4, and 8 in Cabaj et al., 2006).

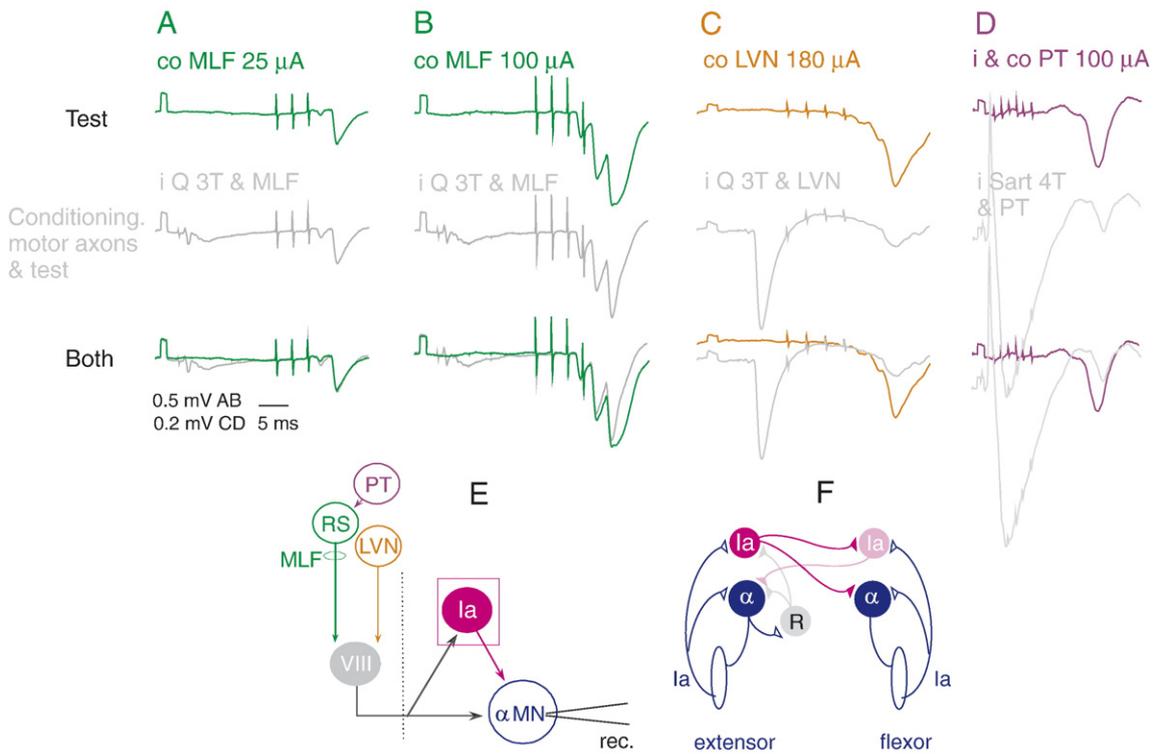


Fig. 6 – Depression of IPSPs evoked in motoneurons from the MLF, LVN and PT by Renshaw cells activated by stimulation of motor axons, as the evidence that the IPSPs are mediated by Ia inhibitory interneurons. Top row, test IPSPs alone. Middle row, IPSPs evoked by the same stimuli but after conditioning stimulation of a muscle nerve followed by activation of Renshaw cells. Bottom row, superposition of the test (black) and conditioned (grey, smaller) IPSPs. All the records are averages of 20 single records. (Left diagram) Pathways via which IPSPs illustrated in panels A–D were evoked. These from reticulospinal (RS) neurones with axons in the medial longitudinal fascicle (MLF) excited by pyramidal tract (PT) neurones or from vestibulospinal (VS) neurones via lamina VIII commissural interneurons and Ia inhibitory interneurons, the latter two represented by the grey circles labelled VIII and Ia, to alpha motoneurons. Right diagram, network of Ia inhibitory interneurons (Ia) and Renshaw cells (R) with which the networks to the left were linked. (A and B, modified from Fig. 2 in Jankowska et al., 2005c).

group Ib afferents (Harrison and Jankowska, 1985) or group II afferents (Edgley and Jankowska, 1987) of the same nerve, and might represent a kind of negative feedback. In other cases, they might reflect interactions between interneurons subserving opposite actions, for example, Ia inhibition of either flexors or extensors (Hultborn et al., 1976a,b) or commissural interneurons on the left and right sides (Edgley and Jankowska, 1987; Harrison et al., 1986). Mutual excitatory interactions have also been found but these might be more likely between interneurons of distinct neuronal populations (Bannatyne et al., 2006; Jankowska et al., 2002a,b).

4. Which neurones do and which do not belong to a neuronal network

Boundaries between different neuronal networks may be considered as not being sharp, especially when individual neurones form part of different networks under different circumstances and when neuronal networks change their configuration and elements depending on which movements they subserve. It may thus be a matter of personal preferences whether different kinds of neurones are classified as belonging to the same, or to different, networks. However, independently of how spinal interneuronal

networks are defined, there is, no doubt, that their constituent neurones cannot be distinguished by mere topographical factors. Interneurons that appear to belong to one interneuronal population are, as a rule, intermixed with other types of neurones, are distributed over considerable lengths of the spinal cord and do not form nuclear complexes in any parts of the spinal grey matter, even if they are preferentially located in more rostral or more caudal segments, and in more dorsal or more ventral Rexed's laminae.

Although neither functional nor morphological features allow an unequivocal delimitation of the interneuronal networks discussed above, the general conclusion that elementary interneuronal networks are building blocks of larger networks would hold true. It might therefore be more heuristic to look for these elementary networks while analyzing more complex networks rather than to try to identify constituent neurones of the complex networks “from scratch.”

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